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U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

09/857123

INTERNATIONAL APPLICATION NO.
PCT/EP99/09284INTERNATIONAL FILING DATE
30 November 1999PRIORITY DATE CLAIMED
1 December 1998

TITLE OF INVENTION

HUMAN VANILLOID RECEPTORS AND THEIR USES

APPLICANT(S) FOR DO/EO/US

DELANY, Natalie Samantha
SANSEAU, Philippe

TATE, Simon Nicholas

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (24) indicated below.
4. The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. is attached hereto (required only if not communicated by the International Bureau).
 - b. has been communicated by the International Bureau.
 - c. is not required, as the application was filed in the United States Receiving Office (RO/US).
6. An English language translation of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. is attached hereto.
 - b. has been previously submitted under 35 U.S.C. 154(d)(4).
7. Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. are attached hereto (required only if not communicated by the International Bureau).
 - b. have been communicated by the International Bureau.
 - c. have not been made; however, the time limit for making such amendments has NOT expired.
 - d. have not been made and will not be made.
8. An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. An English language translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).
11. A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. A copy of the International Search Report (PCT/ISA/210).

Items 13 to 20 below concern document(s) or information included:

13. An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. A **FIRST** preliminary amendment.
16. A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. A substitute specification.
18. A change of power of attorney and/or address letter.
19. A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.
20. A second copy of the published international application under 35 U.S.C. 154(d)(4).
21. A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
22. Certificate of Mailing by Express Mail
23. Other items or information:

Copy of Request (PCT/RO/101)

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 091857123	INTERNATIONAL APPLICATION NO. PCT/EP99/09284	ATTORNEY'S DOCKET NUMBER PG3606USW
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24. The following fees are submitted:

BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :

- | | |
|--|-----------|
| <input type="checkbox"/> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO | \$1000.00 |
| <input checked="" type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO | \$860.00 |
| <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO | \$710.00 |
| <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) | \$690.00 |
| <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) | \$100.00 |

CALCULATIONS PTO USE ONLY

ENTER APPROPRIATE BASIC FEE AMOUNT =

\$860.00

Surcharge of \$130.00 for furnishing the oath or declaration later than 20 30 months from the earliest claimed priority date (37 CFR 1.492 (e)).

\$0.00

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	
Total claims	51 - 20 =	31	x \$18.00	\$558.00
Independent claims	5 - 3 =	2	x \$80.00	\$160.00
Multiple Dependent Claims (check if applicable).			<input type="checkbox"/>	\$0.00

TOTAL OF ABOVE CALCULATIONS =

\$1,578.00

Applicant claims small entity status. (See 37 CFR 1.27). The fees indicated above are reduced by 1/2.

\$0.00

SUBTOTAL =

\$1,578.00

Processing fee of \$130.00 for furnishing the English translation later than 20 30 months from the earliest claimed priority date (37 CFR 1.492 (f)).

\$0.00

TOTAL NATIONAL FEE =

\$1,578.00

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable).

\$0.00

TOTAL FEES ENCLOSED =

\$1,578.00

<input type="checkbox"/> Amount to be: refunded	\$
<input type="checkbox"/> charged	\$

- a. A check in the amount of _____ to cover the above fees is enclosed.
- b. Please charge my Deposit Account No. 07-1392 in the amount of \$1,578.00 to cover the above fees. A duplicate copy of this sheet is enclosed.
- c. The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 07-1392. A duplicate copy of this sheet is enclosed.
- d. Fees are to be charged to a credit card. **WARNING:** Information on this form may become public. **Credit card information should not be included on this form.** Provide credit card information and authorization on PTO-2038.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:



23347
PATENT TRADEMARK OFFICE

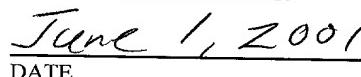

SIGNATURE

Frank Grassler

NAME _____

31,164

REGISTRATION NUMBER _____


DATE

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re: Application of: DELANY et al.
International Application No.: PCT/EP99/09284
International Filing Date: 30 November 1999
Title: HUMAN VANILLOID RECEPTORS AND THEIR USES

Honorable Commissioner of Patents
Washington, D.C. 20231

FIRST PRELIMINARY AMENDMENT

Dear Sir:

The above-identified application is being transmitted herewith for entry in the US National Phase under Chapter II of the PCT for the purpose of adding the priority information. Please amend the application as follows:

In the Abstract:

The Abstract has been placed on a separate sheet of paper according to US practice, as required under 37 CFR 1.72(b).

In the Specification:

On the first line of the specification, after the Title, please add:

--This application is filed pursuant to 35 U.S.C. §371 as a United States National Phase Application of International Application No. PCT/EP99/09284 filed 30 November 1999, which claims priority from Great Britain Application No. 9826359.3 filed 1 December 1998.--

In the Claims

14. (Amended) An expression vector comprising a nucleotide sequence according to claim 6, which is capable of expressing an hVR protein or a variant thereof.

23. (Amended) An antibody specific for a human vanilloid receptor (hVR) protein or a variant thereof as claimed in claim 1.

26. (Amended) A method for identification of a compound which exhibits hVR modulating activity comprising contacting a human vanilloid receptor (hVR) protein or a variant thereof according to claim 1 with a test compound and detecting modulating activity or inactivity.

45. (Amended) A method of producing an hVR protein or a variant thereof according to claim 1 comprising introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVR protein or a variant

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To be assigned

thereof, under conditions suitable for obtaining expression of the hVR protein or variant thereof.

50. (Amended) A human vanilloid receptor (hVR) protein according to claim 48, which is hVR1 or a variant thereof.

51. (Amended) A human vanilloid receptor (hVR) protein according to claim 48 which is hVR3 or a variant thereof.

REMARKS

Applicants have attached an abstract on a separate sheet of paper as required by US practice. Applicants have amended the specification for purposes of adding the priority information. Claims have been amended to eliminate multiple dependencies.

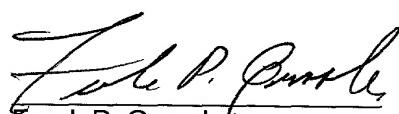
Attached hereto is a marked-up version of the changes made to the specification by the current amendment. The attached page is captioned "Version with markings to show changes made." Applicant respectfully requests the entry of the above preliminary amendments.

Examiner is invited and encouraged to contact the undersigned if such contact would facilitate prosecution of this application.

No fee is believed due in connection with this Amendment, however the Commissioner is hereby authorized to charge any under-payment to Deposit Account No. 07-1392.

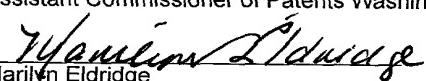
Respectfully submitted,

Date: June 1, 2001


Frank P. Grassler
Attorney of Record, Reg. No. 31,164

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CERTIFICATE OF EXPRESS MAILING (37 CFR 1.10)
I hereby certify that this paper (along with any referred to as being attached or enclosed) is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10 in an envelope addressed to: Assistant Commissioner of Patents Washington, D.C. 20231 on 6/01/01


Marilyn L. Eldridge

Version with markings to show changes

14. An expression vector comprising a nucleotide sequence according to [any one of] claim[s] 6 [to 13].
23. An antibody specific for a human vanilloid receptor (hVR) protein or a variant thereof as claimed in [any one of] claim[s] 1 [to 5].
26. A method for identification of a compound with exhibits hVR modulating activity comprising contacting a human vanilloid receptor (hVR) protein or a variant thereof according to [any one of] claim[s] 1 [to 5] with a test compound and detecting modulating activity or inactivity.
45. A method of producing an hVR protein or a variant thereof according to [any one of] claim[s] 1[-5] comprising introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVr protein or a variant thereof, under conditions suitable for obtaining expression of the hVR protein or variant thereof.
50. A human vanilloid receptor (hVR) protein according to claim 48 [or 49] which is hVR1 or a variant thereof.
51. A human vanilloid receptor (hVR) protein according to claim 48 [or 49] which is hVR3 or a variant thereof.

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WO 00/32766

PCT/EP99/09284

Rec'd PCT/PTO 01 JUN 2001

HUMAN VANILLOID RECEPTORS AND THEIR USES

Field of the Invention

The present invention relates to human vanilloid receptor (hVR) proteins and to related nucleotide sequences, expression vectors, cell lines, antibodies screening methods, compounds, methods of production and methods of treatment, as well as other related aspects.

Background of the Invention

Capsaicin, the irritant in hot peppers and a member of the vanilloid family activates a sub-group of sensory neurons: the nociceptors. These neurons transmit nociceptive and thermoceptive pain information back to pain-processing centres in the central nervous system such as the spinal cord and the brain. They are also sites for the release of pro-inflammatory mediators in the periphery (1). Nociceptors show heterogeneity in their sensitivity to capsaicin. Excitation and prolonged exposure of these neurons to capsaicin is followed by a refractory state known as desensitisation (2) when they become insensitive to capsaicin and other noxious stimuli (3). The long-term response to insensitivity could be explained by death of the nociceptors or destruction of its peripheral terminals (4). Because of the desensitisation phenomenon, capsaicin has been used therapeutically for decades as an analgesic agent for the treatment of pain in a range of disorders (5).

It has been speculated that the endogenous target for capsaicin plays an important function in the detection of painful stimuli. It has been shown by electrophysiological and biochemical studies that capsaicin induces a flux of cations in dorsal root ganglion (DRG) neurons (6,7). Because other vanilloid derivatives show responses in a dose dependent manner (8,9) a receptor is the most likely candidate to explain the mechanism. Therefore, based on indirect evidence it has been anticipated that these actions of capsaicin (excitation / desensitisation) are mediated by a specific membrane-bound receptor named vanilloid receptor (10).

Evidence for the existence of a vanilloid receptor came from binding experiments with resiniferatoxin (RTX), a capsaicin analog (11), and a competitive antagonist

of capsaicin, capsazepine (12). Vanilloid receptors have been visualised by using ($[^3\text{H}]$ -RTX) autoradiography in dorsal root ganglia (DRG) and spinal cord of different species including man (13,14).

5 Recently, a rat vanilloid receptor termed VR1 has been identified using an expression-cloning strategy to isolate the complementary DNA (cDNA) encoding the corresponding protein from a rat DRG cDNA library (15). The cDNA clone was completely sequenced. The rat VR1 cDNA has an open reading frame of 2,514 nucleotides and encodes for a protein of 838 amino acids with a predicted relative molecular mass of 95,000. Analysis of the amino acid sequence identified 6 potential transmembrane regions with a short hydrophobic stretch between the transmembrane regions 5 and 6. The N-terminus (amino terminal) contains three ankyrin repeat domains. No motifs have been identified at the C-terminus (carboxy terminal).

10 It has been noted that rat VR1 transfected cells exhibit an increase in calcium levels after heat treatment and it has been suggested that *in vivo* VR1 and vanilloid receptors are involved in detection of noxious heat (but not innocuous heat). It has also been proposed that protons could act as modulators of the vanilloid receptors (16, 17, 18).

20 While it has been recognised that the rat capsaicin receptor, VR1, is a member of the family of non-selective ion channels that are gated by ligands and that it is involved in pain sensation, the natural ligand of VR1 remains unknown. It is therefore suggested that human vanilloid receptor sub-types may provide targets for the development of novel analgesic agents (agonists and antagonists) and agents which may interact with other disorders.

30 Accordingly, it is an object of the present invention to locate and characterise human vanilloid receptors. Other objects of the present invention will become apparent from the following detailed description thereof.

Summary of the Invention

35 According to one embodiment of the present invention there is provided an isolated human vanilloid receptor (hVR) protein or a variant thereof. Preferably

the hVR protein is an hVR1 or hVR3 protein or a variant thereof. In a particularly preferred aspect of the invention the hVR protein has an amino acid sequence as shown in figure 3 or in figure 18.

- 5 According to another aspect of the invention, there is provided a human vanilloid receptor (hVR) protein or a variant thereof, preferably hVR1 or hVR3 or a variant thereof, for use in a method of screening for agents useful in the treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity, preferably hVR1 or hVR3 activity, in a human patient. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder such as asthma or chronic obstructive pulmonary disease (COPD), a urological disorder such as diabetic neuropathy, incontinence and interstitial cystitis, or an inflammatory disorder.
- 10 According to another aspect of the invention there is provided a nucleotide sequence encoding a human vanilloid receptor (hVR) protein or a variant thereof as hereinbefore described, or a nucleotide sequence that is complementary thereto. Preferably the nucleotide sequence encodes an hVR1, hVR3 protein or variant thereof or a nucleotide sequence which is complementary thereto. Particularly preferably the nucleotide sequence is as shown in figure 2 and figure 17.
- 15
- 20 According to another aspect of the invention there is provided an expression vector comprising a nucleic acid sequence as referred to above which is capable of expressing an hVR protein as hereinbefore described or a variant thereof, preferably hVR1 or hVR3 or a variant thereof. Preferably the expression vector is as displayed in figure 6 or figure 20.
- 25
- 30 According to another aspect of the invention there is provided a stable cell line comprising an expression vector as referred to above which is capable of expressing an hVR protein as hereinbefore described or a variant thereof, preferably hVR1 or hVR3 or a variant thereof. The stable cell line is preferably a

modified mammalian cell line, preferably HEK293, CHO, COS, HeLa or BHK although transient expression may be preferred in *Xenopus* oocytes.

5 According to another aspect of the invention there is provided an antibody specific for an hVR protein as hereinbefore described or a variant thereof, preferably specific for hVR1 or hVR3 or a variant thereof.

According to another aspect of the invention there is provided a method for identification of a compound which exhibits hVR modulating activity, comprising contacting an hVR protein as hereinbefore described or a variant thereof, preferably hVR1 or hVR3 or a variant thereof, with a test compound and detecting modulating activity or inactivity.

According to another aspect of the invention there is provided a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above.

20 According to another aspect of the invention there is provided a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above, for use in therapy.

According to another aspect of the invention there is provided the use of a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above, in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity, preferably hVR1 activity or hVR3 activity, in a human patient. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder such as asthma or chronic obstructive pulmonary disease (COPD), a urological disorder such as neuropathy, incontinence or interstitial cystitis, or an inflammatory disorder.

35 According to another aspect of the invention there is provided a method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR,

preferably hVR1 or hVR3, activity in a human patient which comprises administering to said patient an effective amount of a compound identifiable by the method referred to above. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain,
5 neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), respiratory disorders such as asthma and chronic obstructive pulmonary disease (COPD) and urological disorders including diabetic neuropathy, incontinence and interstitial cystitis and inflammatory disorders.

According to another aspect of the invention there is provided a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above, excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodial, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial, β-acaridial, scutigera, merulidial, anandamide and capsazepine.

According to another aspect of the invention there is provided a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above, excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodial, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial, β-acaridial, scutigera, merulidial, anandamide and capsazepine, for use in therapy.

According to another aspect of the invention there is provided the use of a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above, excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodial, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial, β-acaridial, scutigera, merulidial, anandamide and capsazepine, in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity, preferably hVR1 activity or hVR3 activity, in a human patient. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic

5 pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder such as asthma or chronic obstructive pulmonary disease (COPD), a urological disorder such as neuropathy, incontinence or interstitial cystitis, or an inflammatory disorder.

10 According to another aspect of the invention there is provided a method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR, preferably hVR1 or hVR3, activity in a human patient which comprises administering to said patient an effective amount of a compound identifiable by the method referred to above, excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodial, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial, β -acardial, scutigeral, merulidial, anandamide and capsazepine. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), respiratory disorders such as asthma and chronic obstructive pulmonary disease (COPD) and urological disorders including diabetic neuropathy, incontinence and interstitial cystitis and inflammatory disorders.

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According to another aspect of the invention there is provided a compound identified by the method referred to above.

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According to another aspect of the invention there is provided a compound identified by the method referred to above, for use in therapy.

25

According to another aspect of the invention there is provided the use of a compound identified by the method referred to above in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity, preferably hVR1 activity or hVR3 activity, in a human patient. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder such as asthma or chronic

obstructive pulmonary disease (COPD), a urological disorder such as neuropathy, incontinence or interstitial cystitis, or an inflammatory disorder.

According to another aspect of the invention there is provided a method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR, preferably hVR1 or hVR3, activity in a human patient which comprises administering to said patient an effective amount of a compound identified by the method referred to above. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), respiratory disorders such as asthma and chronic obstructive pulmonary disease (COPD) and urological disorders including diabetic neuropathy, incontinence and interstitial cystitis and inflammatory disorders.

According to another aspect of the invention there is provided a method of producing an hVR protein as hereinbefore described or a variant thereof, preferably hVR1 or hVR3 or a variant thereof, comprising introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVR protein or a variant thereof, preferably hVR1 or hVR3 or a variant thereof, under conditions suitable for obtaining expression of the hVR protein or a variant thereof, preferably hVR1 or hVR3 or a variant thereof.

Brief Description of the figures

Figure 1 is an alignment of hVR1 *in silico* derived clusters with rat VR1.

Figure 2 displays the human VR1 nucleotide sequence including the 5'UTR (nt – 773 to nt 0), coding region (nt 1 to 2517) and 3'UTR (nt 2518 to nt 3560).

Figure 3 illustrates the nucleotide and encoded amino acid sequence of the human VR1sequence.

Figure 4 depicts the amino acid sequence of the hVR1 gene, the shading denotes predicted trans-membrane regions (boxed) and the ankyrin binding domains (unboxed). The predicted phosphorylation sites are underlined.

Figure 5 is a comparison of the amino acid sequences of the rat (rVR1) and human (hVR1) vanilloid receptors.

Figure 6 illustrates constructs pBluescriptSK(+) (A) and pCIN5-new (B) with the full length hVR1 gene cloned via NotI and EcoRI restriction sites.

Figure 7 shows a Slot Blot hybridisation with hVR1 probe with positive labelling of both rat and human DRG mRNA.

5 Figure 8 displays a Western blot probed with anti-VR1 antibodies with the arrow indicating the VR1 specific protein.

Figure 9 shows localisation of VR1 in rat DRG tissue sections, the arrow points to VR1 expressing small diameter (<25 μ m) neurone cell bodies.

Figure 10 depicts the *in situ* localisation of VR1 in human DRG sections (A) and human skin (B).

Figure 11 illustrates the functional response to capsaicin and blockade by capsazepine (CPZ) (A) with the current voltage relationship plotted in (B) on human VR-1 channels, transiently expressed in HEK293T cells.

Figure 12 shows capsaicin-induced desensitisation of human VR-1 channels in the presence of 2mM external calcium (A), maximum current (65mV) against time (B) and current voltage relationship in the absence of Ca²⁺ (C).

Figure 13 shows the influx of calcium into transiently transfected HEK293T cells over a time course in the presence of agonist capsaicin, anandamide and resiniferatoxin in the absence (A, B, D and F) or presence (C, E, G) of the antagonist, capsezipine.

20 Figure 14 illustrates a graphical presentation the results shown in figure 13 examining the response of hVR1 transfected HEK293T cells over time before and after exposure to agonists: capsaicin, anandamide and resiniferatoxin in the absence (A, B, D and F) or presence (C, E, G) of the antagonist, capsezipine.

25 Figure 15 displays the proposed assay strategy to carry out drug screening.

Figure 16 displays an alignment of *in silico* derived hVR3 specific clusters with rat VR1.

Figure 17 depicts the hVR3 nucleotide sequence including the 5' UTR (nt -686 to nt 0) Coding region (nt1 to nt 2889), 3'UTR (nt 2890 to nt 3418).

30 Figure 18 shows the nucleotide and amino acid sequence of hVR3.

Figure 19 is of the amino acid sequence of hVR3, the shading denotes predicted trans-membrane regions (boxed) and the ankyrin binding domains (unboxed).

Figure 20 displays constructs pBluescriptSK(+) (A) and pCDNA3.1 (+) (B) with the full length hVR3 gene cloned via NotI and Xhol restriction sites.

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Figure 21 illustrates a multiple comparison of the amino acid sequences of the rat VR1 and the human vanilloid receptors: hVR1, hVRL-1 and hVR3.

Figure 22 Northern Blot hybridisation with hVR3 probe with strong signals detected in trachea (A), prostate (B), placenta, kidney and pancreas (C).

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Detailed Description of the Invention

Throughout the present specification and the accompanying claims the words "comprise" and "include" and variations such as "comprises", "comprising", "includes" and "including" are to be interpreted inclusively. That is, these words are intended to convey the possible inclusion of other elements or integers not specifically recited, where the context allows.

As referred to above, the present invention relates to isolated human vanilloid receptor (hVR) proteins, and in particular to the human vanilloid receptors which will be termed respectively human vanilloid receptors 1 and 3 (hVR1, and hVR3), sequence information for which is provided in figures 2 (hVR1) and 17 (hVR3). In the context of this invention the term "isolated" is intended to convey that the receptor protein is not in its native state, insofar as it has been purified at least to some extent or has been synthetically produced, for example by recombinant methods. The term "isolated" therefore includes the possibility of the receptor protein being in combination with other biological or non-biological material, such as cells, suspensions of cells or cell fragments, proteins, peptides, organic or inorganic solvents, or other materials where appropriate, but excludes the situation where the receptor protein is in a state as found in nature.

20

Routine methods, as further explained in the subsequent experimental section, can be employed to purify and/or synthesise the receptor proteins according to the invention. Such methods are well understood by persons skilled in the art, and include techniques such as those disclosed in Sambrook, J. et al. (28), the disclosure of which is included herein in its entirety by way of reference.

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By the term "variant" what is meant throughout the specification and claims is that other peptides or proteins which retain the same essential character of the human vanilloid receptor proteins for which sequence information is provided, are also intended to be included within the scope of the invention. For example,

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other peptides or proteins with greater than about 80%, preferably at least 90% and particularly preferably at least 95% homology with the sequences provided are considered as variants of the receptor proteins. Such variants may include the deletion, modification or addition of single amino acids or groups of amino acids within the protein sequence, as long as the peptide maintains the biological functionality of a human vanilloid receptor. This biological functionality can of course be assessed by conducting binding studies with known vanilloid modulators such as capsaicin, capsazepine (12) and resiniferatoxin (11).

Human VR1 is preferentially expressed in human dorsal root ganglia (DRG) and relative to hVR3 has the highest sequence homology with the rat VR1. Therefore, hVR1 is likely to be the human orthologue to rat VR1. hVR3 is less similar to rat VR1 and is expressed in a wider range of tissues. Nucleotide sequence analysis of hVR1 reveals a 2517bp open reading frame which encodes an 839 amino acid protein (see figures 2, 3 and 4). This deduced hVR1 protein sequence is 86 % identical to the rat VR1 (15) and shares many of its characteristics such as 6 transmembrane regions with an hydrophobic stretch between transmembrane 5 and 6 and an N-terminus which contains 3 ankyrin repeat domains. Similarly hVR3 has an open reading frame of 2889bp open reading frame which encodes a 963 amino acid protein (see figures 17, 18 and 19). The deduced hVR3 protein is 46 % identical to rat VR1 and 44 % identical to hVR1 sharing many of VR1's characteristics such as 6 transmembrane regions with an hydrophobic stretch between transmembrane 5 and 6 and an N-terminus which contains 3 ankyrin repeat domains.

The invention also includes nucleotide sequences which encode for human vanilloid receptor proteins or variants thereof as well as nucleotide sequences which are complementary thereto. Preferably the nucleotide sequence is a DNA sequence and most preferably, a cDNA sequence. Preferably the proteins are hVR1, hVR3 or variants thereof. Such nucleotides can be isolated or synthesised according to methods well known in the art. See reference 28, the disclosure of which is included herein in its entirety by way of reference.

The present invention also includes expression vectors which comprise nucleotide sequences encoding for the hVR, preferably hVR1 or hVR3, receptor

proteins or variants thereof. A further aspect of the invention relates to an expression vector comprising nucleotide sequences encoding for hVR1 or hVR3 receptor proteins or variants thereof. Such expression vectors are routinely constructed in the art of molecular biology and may for example involve the use of plasmid DNA and appropriate initiators, promoters, enhancers and other elements, such as for example polyadenylation signals which may be necessary, and which are positioned in the correct orientation, in order to allow for protein expression. Suitable vectors for use in practicing the present invention include pBluescript (Stratagene), pCR-Script (Stratagene), pCR2.1-TOPO (Invitrogen), pCRII-TOPO (Invitrogen), pCR-Blunt (Invitrogen), with vectors such as pCIN (32) (available from Clontech as pIRES-neo), pCDNA 3.1 (Invitrogen) or pCIneo (Promega) required for mammalian expression. Appropriate methods can be effected by following protocols described in many standard laboratory manuals (28, 29).

The invention also includes cell lines which have been modified to express the novel receptor. Such cell lines include transient, or preferably stable higher eukaryotic cell lines, such as mammalian cells or insect cells, lower eukaryotic cells, such as yeast or prokaryotic cells such as bacterial cells. Particular examples of cells which have been modified by insertion of vectors encoding for the receptor proteins according to the invention include HEK293T cells and *Xenopus* oocytes. Preferably the cell line selected will be one which is not only stable, but also allows for mature glycosylation and cell surface expression of the inventive receptor. Representative examples of appropriate hosts include animal cells such as HEK293, CHO, COS, HeLa and BHK.

It is also possible for the receptors of the invention to be transiently expressed in a cell line or on a membrane, such as for example in a baculovirus expression system. Such systems, which are adapted to express the receptors according to the invention, are also included within the scope of the present invention.

In particular, the functional hVR protein may include hVR receptor proteins selected from hVR1 and hVR3 and thereof or even other hVR protein subtypes or splice variants which have not yet been identified.

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According to another aspect, the present invention also relates to antibodies, preferably monoclonal antibodies, which have been raised by standard techniques and are specific for the receptor proteins or variants thereof according to the invention. Such antibodies could for example be useful in purification; isolation or screening involving immuno precipitation techniques and may be used as tools to further elucidate hVR, preferably hVR1 or hVR3, protein function, or indeed as therapeutic agents in their own right. Antibodies may also be raised against specific epitopes of the receptors according to the invention.

An important aspect of the present invention is the use of receptor proteins according to the invention in screening methods designed to identify compounds which act as receptor ligands and which may be useful to modulate receptor activity. In general terms, such screening methods will involve contacting the receptor protein concerned, preferably hVR1 or hVR3, with a test compound and then detecting modulation in the receptor activity, or indeed detecting receptor inactivity, which results. For further details on the screening strategy refer to figure 15. The present invention also includes within its scope those compounds which are identified as possessing useful hVR, preferably hVR1 or hVR3, modulation activity, by the screening methods referred to above. The screening methods comprehended by the invention are generally well known to persons skilled in the art. High throughput screens may include fluorescence based assays using the Fluorometric Imaging Plate Reader (FLIPR) with calcium sensitive dyes, and reporter gene assays using calcium sensitive photoproteins that emit light on the influx of calcium and can be detected using an Imaging system. Secondary screens may involve electrophysiological assays utilising patch clamp technology to identify small molecules, antibodies, peptides, proteins or other types of compounds that interact with hVR, preferably hVR1 or hVR3, to modulate activity. Tertiary screens may involve the study of modulators in well characterised rat and mouse models of pain. These models of pain include, but are not restricted to, intraplantar injection of inflammatory agents such as carageenan, formalin and complete freunds adjuvant (CFA). Models of neuropathic pain such as loose ligature of the sciatic nerve are also included. Other screens may involve the study of modulators in human volunteers subject to topically applied capsaicin.

Another aspect of the present invention is the use of compounds which have been identified by screening techniques referred to above in the treatment or prophylaxis of disorders which are responsive to modulation of hVR, preferably hVR1 or hVR3, receptor activity, in a human patient. By the term "modulation" what is meant is that there will be either agonism or antagonism at the receptor site which results from ligand binding of the compound at the receptor. By the term "modulation" what is meant is that there will be either agonism or antagonism at the receptor site which results from ligand binding of the compound at the receptor excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodial, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial, β -acaridial, scutigeral, merulidial, anandamide and capsazepine. hVR , preferably hVR1 and hVR3, proteins have been implicated in disorders of the central nervous system (CNS), gastrointestinal (GI) tract, lungs and bladder and therefore modulation of hVR, preferably hVR1 or hVR3, receptor activity in these tissues will result in a positive therapeutic outcome in relation to such disorders. In particular, the compounds which will be identified using the screening techniques according to the invention will have utility for treatment and/or prophylaxis of disorders such as pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, IBS, respiratory disorders such as asthma and COPD, urological disorders including diabetic neuropathy, incontinence and interstitial cystitis, and inflammatory disorders. It is to be understood however, that the mention of such disorders is by way of example only, and is not intended to be limiting on the scope of the invention.

The compounds which are identified according to the screening methods outlined above may be formulated with standard pharmaceutically acceptable carriers and/or excipients as is routine in the pharmaceutical art, and as fully described in Remmington's Pharmaceutical Sciences, Mack Publishing Company, Eastern Pennsylvania, 17th Ed, 1985, the disclosure of which is included herein in its entirety by way of reference.

The compounds may be administered via enteral or parenteral routes such as via oral, buccal, anal, pulmonary, intravenous, intraarterial, intramuscular, intraperitoneal, topical or other appropriate administration routes.

- 5 The present invention will be further explained, by way of examples, in the appended experimental section. Reference examples are provided.

Experimental details

10 **Reference Example A: Identification of related human ESTs (Expressed Sequence Tags) (19) to the rat VR1 sequence by *in silico* analysis**

The full-length rat VR1 amino acid sequence (15) was used as a query sequence using the tBlastn (20) alignment program to identify related human genes in the dbEST (21) and Incyte (Incyte Pharmaceuticals, Inc., 3174 Porter Drive, Palo Alto, California 94304, USA) databases. Several human ESTs were identified and those with similarities greater than 50% selected for further analysis. One of these ESTs was T12251 previously shown to have 68% amino-acid identity and 84% similarity over a region of 70 amino acids (15). Full-length cloning and functional characterisation of the gene represented by this cluster has been completed (30). This gene was denoted hVRL-1 and encoded a protein of 764 amino acid protein that was 48 % identical to the rat VR1 protein. All human ESTs from both databases were clustered to identify overlapping identical ESTs belonging to the same transcript. The GCG package (Wisconsin 20 Package Version 9.0, Genetics Computer Group (GCG), Madison, Wisconsin) and a program developed in house termed ESTBlast (22) were used to build up these clusters. In total, forty-three ESTs derived from different tissue sources and both EST databases were clustered into ten groups, one of these clusters represented hVRL-1. The remaining nine clusters have been named hVRa, hVRb, hVRc, hVRd, hVRe, hVRf, hVRg, hVRh and hVRi. For each EST the tissue source was assigned according to the annotations in the dbEST and Incyte databases. Since no obvious starting codon was present and the cluster sequences were shorter than the rat VR1 transcript none of these clusters were likely to represent a full-length vanilloid receptor transcript. Finally hVRg, hVRh and hVRi collapsed into a single contig. Sequence analysis has shown that

these cDNAs are likely to be chimeric. The 5' end has weak similarities with the rat VR1 gene but the 3' end is identical to a DNA binding protein. No more work was pursued with that transcript.

5 **Reference Example B: Isolation of the human orthologue to the rat VR1 gene (reference examples B1-B4):**

Reference Example B1: *In silico* assembly of human VR1

The consensus nucleotide sequences from the ten clusters were searched with the tBlastx program (20) against the rat VR1 sequences to identify the most likely open reading frames. Frame shifts were corrected when the sequence trace files were available. Each cluster was aligned against the rat VR1 amino-acid sequence according to the Blastx results. The Blastx alignment program (20) was used to compare the full-length rat VR1 protein with the amino-acid sequences of the ten clusters. The three clusters with the highest homology, displayed in figure 1, were aligned with the rat VR1 gene.

20 Cluster hVRa shared a high homology (70% identity and 75% similarity over a stretch of 107 amino acids) with the 5' of the rat VR1 sequence but did not seem to have a potential start codon. It contained two ESTs (EST1 and EST2) derived from the same tissue, bladder, and from the same patient. These two ESTs were selected for further investigation since this cluster was the most 5', had high homology with rat VR1 and the bladder tissue could be contaminated with sensory neurones. Both cDNA clones were ordered but only clone EST1 was received as EST2 failed the recovery procedure.

25 Cluster hVRb composed of two EST's (EST3 and EST4), with 89% identity and 92% similarity over 90 residues, showed the highest degree of homology to the rodent sequence. The overlap between both sequences was located towards the middle of the gene.

30 hVRc (EST5) also while having high homology (71% identity and 75% similarity over 65 residues) with rat VR1 was closely related to the C-terminus of the rat protein sequence.

Reference Example B2: Sequencing of clones

All DNA sequences were determined by automated DNA sequencing based on the dideoxy chain-termination method using the ABI 373A / 377 sequencers (Applied Biosystems). Sequence-specific primers were used with the 'Big-Dye' Terminator Cycle Sequencing kit (Applied Biosystems). The nucleotide sequence was analysed using programs from the University of Wisconsin Genetics Computer Group package.

More specifically when sequencing an EST clone, the following protocol was adopted. The EST1 clone was grown using standard procedures and DNA was isolated using Qiagen columns. SP6 (5' ATTTAGGTGACACTATAG) and T7 (5' TAATACGACTCACTATAGGG) primers flanking the cloning site were used to sequence both ends. Plasmid DNA (0.6 pmol) was used with 10.0 pmol of each primer for the dye terminator reaction. The SP6 end corresponded to the *in silico* derived EST sequence (identical to EST1). The T7 end did not have homologies with VR1 nor did it possess a long open reading frame or a polyadenylation motif. The size of the insert was determined by enzyme digestion of the DNA with the endonucleases NotI and EcoRI and calculated to be approximately 3kb.

Plasmid DNA (50ng) was used to amplify the insert by Polymerase Chain Reaction (PCR) with T7 and SP6 as primers. The PCR conditions included an initial hot-start at 94°C for 2 minutes, followed by 35 cycles at 94°C for 45 seconds, 50°C for 45 seconds and 72°C for 1 minute and terminated by 5 minutes at 72°C. The resulting PCR amplicon was separated on a 1.2% agarose gel and shown to be of ~3kb in size.

To fully sequence the PCR product the nuclease-Bal-31 technique was used where both strands of duplex DNA are degraded from both ends (23). After ethanol precipitation of the PCR product, the pellet was re-suspended in 30ml of 1X Bal-31 buffer (add buffer composition). A time-course digest with 2 units of Bal-31 enzyme (Roche Molecular Biochemicals) was carried out with 12 time points taken over 90 minutes (30 seconds, 1, 2, 3, 5, 7, 10, 15, 25, 45, 75 and 90 minutes). Three pools were made respectively from digests 1 to 4, 5 to 8 and 9

to 12. Each pool was blunt-ended and sub-cloned into the pCR-Script SK (+) plasmid from Stratagene at the SrfI site. After transformation, 16 colonies from each pool were screened by PCR with the flanking Reverse (5' GGAAACAGCTATGACCATG) and M13-20 (5' GTAAAACGACGGCCAGT) primers. The amplicons of 6 positive colonies per pool were subjected to direct sequencing (24) using the T3 (5' AATTAAACCCTCACTAAAGGG) and T7 primers. The DNA sequences obtained were assembled using the GCG package, translated and aligned against the rat VR1 gene using the Blast tools. After analysis, the 3079bp amplicon was shown to have 2 introns of 603bp and 1221bp. The latter intron was located at the 3'end of the PCR product. The coding sequence covered 1255 bp and was separated by the former intron. Therefore the clone EST1 was likely to be a partially spliced and incomplete cDNA.

The clone belonging to cluster 1b (EST3) and derived from a kidney cDNA library was ordered and sequenced using the Bal-31 technique. After assembly of the sequences using the GCG package an identical overlap was identified with the DNA sequence of the cluster hVRc. Moreover a 3'end with a polyadenylation signal and tail was identified. The complete sequence of the combined hVRb Bal-31 derived sequence and hVRc was 2063 bp (1020 bp of coding and 1043 bp of 3' untranslated sequence).

Reference Example B3: Amplification of the middle section of hVR1 using the Polymerase Chain Reaction

We formulated the hypothesis that both sequences (hVRa and hVRb/c) were part of a common transcript. If the human and rat VR1 were going to be similar, the 2 contigs should be separated by a gap of approximately 275bp. Primers were designed on both sides of the gap to amplify mRNA from brain tissues in order to clone the gap. A smear was obtained with the sense primer (5' TCTACTTCGGTGAACTGCC) and antisense (5' ACGGCAGGGAGTCATTCTTC). For specificity 50ng of the PCR product were amplified with the nested sense (5' CTGCAGAACTCCTGGCAGA) and antisense (5' GTCACCACCGCTGTGGAAAA) primers. The 900bp nested amplicon was sequenced and shown to be identical to hVRa at one end and

hVRb/c at the other end. The middle part of the PCR product was homologous to the rat VR1 sequence. This region corresponded to 91 amino acids. When the sequences of hVRa, hVRb/hVRc and the internal amplicon are combined the total length of the Open Reading Frame (ORF) is 824 amino acids followed by a 3' untranslated sequence of 1043 bp. The human amino acid sequence is 87% identical to the rat sequence over that part of the coding region. This sequence was termed hVR1 because of its high degree of identity with the rat VR1 sequence.

Reference Example B4: Isolation of the 5' Terminus of hVR1 by PAC isolation

Since no start codon was identified at the 5' end an additional strategy was designed to identify the full-length sequence. Two primers, sense (5' TCCTCTGGCTTCCAACCCGTT) and antisense (5' GAACTGGGCAGAAAGTGCCT) were designed to amplify a 150bp product from the first intron mentioned in reference example B2. A P1 Artificial Chromosome (PAC) genomic clone (25) was isolated by PCR screening of a PAC library (Genome Systems, St Louis, Missouri). PAC DNA was recovered by using standard plasmid isolation protocol (26). An anti-sense primer was designed (5' CTGGAGTTAGGGTCTCCATCC) to sequence the genomic clone towards the potential 5' end of the gene. An open reading frame with a starting codon was identified. The gene structure was confirmed by using the GenScan software (27). The complete gene has a nucleotide sequence of 2517bp (figure 2) and encoded a 839 amino acid protein (Figures 3 and 4). The gene was named hVR1. Multiple alignment of the amino acid sequence of hVR1 and rat VR1 shows a remarkable degree of identity and similarities between both sequences (figure 5). The rVR1 and hVR1 amino acid sequences are 86% identical. Moreover after protein analysis 6 trans-membrane domains and 3 ankyrin binding domains were identified in hVR1 as in the rat VR1 gene.

Example 1: Full-length Amplification of hVR1 from human DRG and assembly into cloning vectors

HVR1 was PCR amplified in three sections from human DRG template. The 5' fragment was amplified using a sense primer encoding a NotI site and a strong

Kozak motif followed by gene specific sequence (5' GTCATAGCGGCCGCGCCGCCACCATGAAGAAATGGAGCAGCAC) and an antisense primer (5' AGGCCCACTCGGTGAACCTTC). The thermo-cycling conditions used for this amplification included a hot start at 94°C for 4 mins, followed by 35 cycles of 94°C for 1 min, 54°C for 1 min and 72°C for 1 min. A final extension step of 72°C for 5 min completed the reaction. The resulting PCR products were separated on a 2% agarose gel and cloned into pCR®II-TOPO according to the manufacturers instructions supplied with the TOPO™ TA Cloning® kit (Invitrogen). The middle section of hVR1 was PCR amplified using the sense primer: 5' GACGAGCATGTACAATGAGA and antisense primer: 5' GTCACCACCGCTGTGGAAAA. The cycling conditions included a hot start at 94°C for 4 mins, followed by 35 cycles of 1 min at 94°C, 56°C and 72°C. A final extension step of 72°C for 5 min completed the reaction. A band of approximately 870 bp was excised from a 2 % agarose gel and cloned as detailed by the TOPO™ TA Cloning® kit into the vector pCR2.1®-TOPO. Finally the 3' end was PCR amplified with the sense primer: 5' TGTGGACAGCTACAGTGAGA and the antisense primer: 5'TGCACTGAATTGAGCACTGGTGTCCCTCAG which encoded an EcoRI site for cloning. The PCR conditions included a 90 sec hot start at 94°C followed by 35 cycles of 94°C for 50 sec, 50°C for 50 sec and 72°C for 50 sec. The cycling was completed with a 72°C step for 5 min. PCR products were separated on a 2% agarose gel and cloned into the vector pCR2.1®-TOPO.

Resulting clones for each of the three hVR1-fragments were taken for sequence analysis and separate clones coding a consensus sequence were used in the full length assembly of the gene. The NotI/DraIII (New England Biolabs) digested 5' end fragment ligated together with the middle DraIII/EcoRI fragment into a NotI/EcoRI restricted pBluescript SK (+) vector (Stratagene). Finally, the remaining 3' fragment was introduced into the resulting construct via MscI and EcoRI restriction sites, a map of the resulting construct is displayed in figure 6A.

Several clones were selected for sequence analysis to confirm that constructs still encoded the hVR1 consensus sequence. These were then digested with NotI/EcoRI and ligated into the mammalian expression vector pCIN5-new (a modified version of pCIN1 (32) having an IVS deletion as well as a 36 bp

deletion repositioning the start codon of neomycin phosphotransferase immediately after the upstream EMVC IRES) as illustrated in figure 6B.

Example 2: Chromosomal Localisation

The primers used to isolate the PAC clone (reference example B4) were selected for PCR on the G3 radiation hybrid panel from Stanford commercially available from Research Genetics (Huntsville, Alabama). The positive lanes and negative patterns were analysed using the public web server at Stanford University (<http://www-sghc.stanford.edu>). After analysis the hVR1 gene appears to be located on human chromosome 17 around marker SHGC-36073 (lod score=9.55).

Example 3: mRNA Distribution

The tissue distribution of hVR1 was established by slot-blot hybridisation. RNA was transferred onto a sheet of GeneScreen hybridisation transfer membrane (DUPONT) sandwiched in a slot blotter by suction via a vacuum pump. Once the membrane was rinsed in 2x SSC (3M sodium chloride and 0.3M sodium citrate pH7) for 2 min it was exposed to UV using an Ultraviolet crosslinker (Amersham Life Science) for 1min at 15000uW/cm² thus enabling cross-linkage of the RNA onto the membrane. The amounts of RNA on the blot are unknown. The probe was obtained by PCR amplification of a 260 bp product of the coding region of hVR1 with the following two primers: 5' TGTGGACAGCTACAGTGAGA and 5' GTGGAAAACCCGAACAAGA. Membranes were hybridised for 4 hr shaking at 60°C in a 10% dextran sulphate, 1% SDS (sodium dodecyl sulphate) and 1M NaCl solution. The probe was labelled with [α 32P]dCTP (Amersham) using the Rediprime™DNA labelling system (Amersham), so as to obtain approximately 500,000cpm of the labelled probe per ml of prehybridisation solution. Briefly 100ng of probe was boiled for 3 minutes (denaturization) and then cooled on ice for 2 minutes in a total volume of 45 μ l. This was added to the labelling tube from the kit together with 3 μ l of 32P dCTP followed by an incubation at 37°C for 30 minutes. 400 μ l of Herring Sperm DNA (Sigma) at a concentration of 8 μ g/ml was added to the labelled probe and heated at 99°C for 3 minutes followed by rapid cooling on ice. The labelled probe was added and mixed well in pre-hybridisation solution. The membranes were hybridised overnight at 55°C.

The membranes were then washed, first at room temperature in 2xSSC and 1% SDS for 5 minutes, followed by 2x SSC and 1% SDS for 30 min at 50°C. If necessary further washes with 1x SSC and 0.5% SDS or 0.1xSSC and 0.1% for 30 mins at the same temperature were carried out. The membranes were then exposed to Scientific Imaging Film AR (Kodak) using intensifying screens at – 70°C overnight and the film developed.

The results are shown on figure 7. Strong signals were observed with the positive controls (slots 4B and 5B). Signals are detected on the human DRG slots (1A and 1B). No signals were detected with the water control (slot 3B). Three multi-tissue northern blots (Clontech) with a wide range of tissues have also been hybridised with the same probe, however no signals were detected. RT-PCR was performed on various tissues with the primer combination used to amplify the probe. A strong band was detected in DRG RNA. Taken together these hybridisations suggest that hVR1 is specifically expressed in neuronal tissue and DRG in particular.

Example 4: Design and production of Anti-hVR1 Antibody

The peptides CHIFTTRSRTRLFGKGDSEEASC (peptide68) and CGSLKPEDAEVFKDSMVPGEK (peptide69) were synthesised by standard solid phase techniques and purified by gel filtration chromatography. These peptides were conjugated via their Cys residues to the carrier protein, Tuberculin PPD (purified protein derivative) using sulpho-SMCC (sulfosuccinimidyl 4-[N-maleimidomethyl]-cyclohexan-1-carboxylate). Rabbits, previously sensitised to Bacillus Calmette Guerin (BCG), were inoculated with the resulting conjugates emulsified in incomplete Freund's adjuvant at approx monthly intervals. Serum was prepared from blood samples taken 7 days after each immunisation. The specific antibody response was followed by indirect enzyme-linked immunosorbent assay (ELISA) using free peptide as antigen. Immunoglobulins were purified from high titre sera using immobilised peptide affinity columns (sulpholink Pierce). Rabbits designated M143, 144 and 145 received peptide68 conjugate, rabbits M146, 147 and 148, peptide69 conjugate.

The antibodies have been validated by specific staining of the recombinant protein expressed in HEK293 cells. Whole cell lysates were prepared in Sample

Buffer (4 ml dH₂O, 1 ml 0.5 M Tris-HCl, pH 6.8, 0.8 ml glycerol, 1.6 ml 10 % w/v SDS, 0.4 ml 2-β mercaptoethanol and 0.2 ml of 0.05 % w/v bromophenol blue) and proteins separated by SDS-PAGE and transferred to a nitrocellulose filter by electroblotting. Following incubation with the antisera, bound immunoglobulins were revealed using HRP-conjugated secondary antibodies and enhanced chemiluminescence (ECL) detection. The antisera showed specific binding to a protein(s) of the appropriate molecular weight(s) in extracts of VR1 transfected cells, but not in control extracts, this is illustrated in figure 8.

Example 5: *In situ* localisation of hVR1 using specific antibody

The purified immunoglobulins have been used for immunohistochemical staining of rat DRG tissue sections. Fixed cryosections of DRG were incubated with antibodies for 48h at 4°C at concentrations between 0.1 to 0.5µg/ml. Following a washing step, bound antibodies were detected by indirect immunofluorescence. The antibodies recognised exclusively small diameter cell bodies of the peripheral sensory neurones as displayed in figure 9. This observation has been extended to human DRG tissues for the anti-peptide68 peptide antibodies demonstrating cross-reactivity with the human sequence as expected. Figure 10A demonstrates labelling of DRG cell bodies with an arrow that points to small diameter neuronal cell body) and in figure 10B the arrow points to labelled neurones innervating human skin.

Example 6: Mammalian Cell Expression (examples 6a-6b)

Example 6a: Transient expression of hVR1 in mammalian cells

HEK293 cells were plated onto a 6 well plate, containing poly-l-lysine coated coverslips, at 5 x 10⁴ cells per well. Next day, fresh media was added to the cells (50% confluent). CalPhos Mammalian Transfection Protocol (Clontech, K2051-1) was used for DNA transfection. For each well of cells, solution A was made up containing 8ug hVR1pCIN5, 2µg pEYFP-N1 reporter DNA, 12.4 µl calcium solution and water to 100µl. Solution B (hepes buffered saline) was slowly vortexed while solution A was added dropwise. The mixture was incubated at room temperature for 20 minutes, and then added to cells. The plate was slowly rocked to distribute the solution. The cells were incubated at 37°C for 5 hours, and then washed with phosphate buffered saline. Fresh culture

medium was added and the plate was incubated 24-48 hours for functional analysis.

Example 6b: Stable expression of hVR1 in mammalian cells

- 5 HEK293 cells were plated onto a 6 well plate at 1×10^5 cells per well. Next day, fresh media was added to the cells (50% confluent). CalPhos Mammalian Transfection Protocol (Clontech, K2051-1) was used for DNA transfection. For each well of cells, solution A was made up containing 2 μ g hVR1pCIN5, 12.4 μ l 2M calcium solution and water to 100 μ l. Solution B (hepes buffered saline) was slowly vortexed while solution A was added dropwise. The mixture was incubated at room temperature for 20 minutes, and then added to cells. The plate was slowly rocked to distribute the solution. The cells were incubated at 37°C for 5 hours, and then washed with phosphate buffered saline. Fresh culture medium was added and the plate was incubated 48 hours at 37°C, 5% CO₂. Cells were harvested into 100mm dishes in selection medium containing 800 μ g/ml geneticin. Cells were then incubated and fed at 4 day intervals. In total around 10 days selection is required for each single cell to multiply into a visible clone. Well-separated clones were each picked (with a gilson tip) into separate wells of a 96 well plate, containing maintenance medium (400 μ g/ml geneticin).
- 10 Cells were expanded into flasks for freezing stocks and functional analysis.
- 15 Stable cells may be plated at 1×10^5 cells onto poly-L-lysine coated coverslips in 6 well plate, for calcium imaging next day.
- 20

Example 7: Functional Analysis of hVR1(examples 7a-7c):

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Example 7a: Electrophysiology using patch clamp methods

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The activation of human VR-1 channels transiently expressed in HEK293T cells by capsaicin was investigated. Cells grown on poly-L-lysine-coated glass coverslips were placed in a recording chamber (0.5ml) and superfused with extracellular solution (2ml min⁻¹). The extracellular solution contained: NaCl (140mM), KCl (5mM), MgCl₂ (2mM), CaCl₂ (2mM), 4-(2-hydroxethyl)-1-piperazineethanesulphonic acid (HEPES, 10mM) and glucose (10mM). The pH was adjusted to 7.4 with NaOH and osmolarity ranged from 310-320mOsm l⁻¹. Patch pipettes (borosilicate glass) were pulled using a Sutter P-97 electrode puller. The pipettes were filled with an internal solution consisting of: CsCl

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(140mM), ethylene glycol-bis(β-aminoethyl ether) *N,N,N',N'*-tetra acetic acid Cs salt (Cs-EGTA, 5mM) and HEPES (10mM). The pH was adjusted to 7.25 using CsOH and the osmolarity ranged from 275-290 mOsm. When filled with this internal solution, patch electrodes had resistances of 2-5 MΩ.

5 Currents were recorded using standard whole-cell voltage clamp recording techniques (31) at room temperature (21-23°C) using an Axopatch 200A amplifier and signals were sampled at 2 or 0.1 kHz. The majority of series resistance errors (80-85%) were minimized with compensation circuitry. Membrane potentials were not corrected for junction potentials (<4 mV). Voltage pulses and data collection were performed on-line using pClamp8 software (Axon Instruments) interfaced with amplifiers. Membrane potentials were maintained at -60mV between protocols.

10 Capsaicin or capsazepine (CPZ) were applied, using a 'fast-flow system', directly onto the recording cell (<1s to equilibrate). The effects of capsaicin were measured either by application during constant recording while holding the membrane potential at -60mV to elicit an inward current, or applying voltage ramps (-100 to +60mV) in the absence and presence of capsaicin. Similarly both these methods of recording currents evoked by the application of capsaicin 15 were used to demonstrate the blockade by the antagonist (CPZ).

20 Figure 11A reveals that application of capsaicin (1 μM), on human VR1 channels transiently expressed in HEK293T cells, produces an inward current when the membrane was held at a potential of -60mV. This response was abolished by 1μM CPZ and the blockade was partially reversible.

25 In the presence of 1 μM capsaicin, voltage ramps (-100 to +70mV) produced a current-voltage relationship demonstrating a substantial outward rectification. Addition of 1μM CPZ completely blocked the current (figure 11B). Again, only partial recovery was observed, especially for the inward currents evoked by 30 negative potentials.

35 Capsaicin-induced desensitisation of human VR-1 channels in the presence of 2mM external calcium is illustrated in figure 12. Voltage ramps (-100 to +70) were applied and the addition of capsaicin (1μM) evoked an outwardly rectifying current. Repeated additions of capsaicin resulted in a progressive 'rundown' in

the size of the response (figure 12A). Figure 12B shows a plot of the current elicited at a potential of +65mV against time illustrating the 'rundown' in current amplitude. Voltage ramps were applied every 20s and capsaicin added at 2min intervals for approximately 40s. By the 6th addition the current had reduced about 4-fold.

When the external calcium was replaced with 5mM EGTA the size of the current increased dramatically (figure 12C). However, when calcium was re-applied to the external solution, the current evoked by capsaicin (1 μ M) was approximately equivalent to that of the 6th addition shown in (figure 12A).

Example 7b: Calcium Imaging with HEK293 expressing hVR1

HEK293 cells expressing hVR1 transiently or stably, were plated onto poly-L-lysine coated cover slips at 1×10^5 cells per well. They were analysed on the following day by calcium imaging (QuantiCell 700, Applied Imaging). On the day of experiment, WASH buffer was prepared by adding CaCl₂ to extracellular medium (ECM) to a final concentration of 2mM, (ECM contains 125mM NaCl, 5mM KCl, 2mM MgCl₂, 0.5mM NaH₂PO₄, 5mM NaHCO₃, 10mM Hepes, 10mM glucose, 0.1% BSA, pH7.4). The calcium sensitive dye solution was prepared by adding 50 μ l 5% pluronic F-127 in DMSO (Molecular Probes) to a vial of fura2-AM (Molecular Probes). After mixing, 20 μ l of the fura2-AM solution was added to 10ml WASH. 1.5 ml was then added to cells, which were then incubated at 37°C for 30 minutes. The plate was washed three times with WASH. 1ml WASH was added and stored in dark. Agonists and antagonists were prepared in WASH at 5x their required assay concentrations. The reagents and assay temperature was kept at 37°C. For the transiently transfected cells, the YFP reporter DNA fluorescence (490nm excitation) was used to identify the transfected cells. Cells were initially imaged in 400 μ l WASH (or 300 μ l WASH plus 100 μ l antagonist e.g. capsazepine). After approximately 1 min, 100 μ l agonist (e.g. capsaicin, anadamide or resiniferatoxin) at 5 x the desired concentration was added to give final 1x concentration. A sequence of images (340/380nm excitation) were taken to monitor calcium influx response in cells before (30-60 secs), and after the addition of agonist (2-5 mins). Figure 13 displays time courses taken for each of the tests set up to look at the affect of the different agonists mentioned above in the presence or absence of the rat VR1 antagonist, capsazepine. The Imager

also plots graphs of respective calcium concentration (nM) versus time (seconds) as shown in figure 14. After the addition of agonist (e.g. capsaicin, indicated by the vertical arrow on graph), the cells expressing hVR1 are stimulated to influx calcium. This is shown by the appearance of peak on the trace. The peak height correlates with hVR1 expression level. Varying levels of expression is sometimes seen depending on which cells are selected for the graph. Similar experiments may be accomplished to examine the response of protons and heat.

Example 7c: Use of a FLIPR assay with VR1

FLIPR (Fluorometric Imaging Plate Reader) is a high throughput fluorescence-based drug discovery tool for functional cell analysis. Intracellular calcium is monitored with the calcium sensitive dye, fluo3-AM. HEK293 cells stably expressing rat VR1 were plated into a 96 well, poly-L-lysine treated FLIPR plate at 3×10^4 cells per well. On the following day, the plate was processed for FLIPR. FBP buffer was prepared (15 μ M Probenecid (calcium ATPase pump blocker) in 1x FLIPR buffer (145mM NaCl, 5mM KCl, 1mM MgCl₂, 2mM CaCl₂, 10mM glucose, 20mM Hepes). FBP buffer pH was then adjusted to 7.4 with NaOH. 400 μ l DMSO was added to a vial of fluo3-AM (Cambridge Bioscience, F-1241). The fluo3-AM solution was incubated at 37°C for 10 min and vortexed. LOAD was prepared by adding 20 μ l of fluo3-AM solution and 20 μ l 20% pluronic F-127 in DMSO (Cambridge Bioscience, P-3000) into 10 ml FBP. The 96 well plate containing cells was flicked off to remove cell medium. 100 μ l LOAD was added per well. Cells were then incubated at 37°C for 60 minutes. Capsaicin (a rVR1 agonist) and capsazepine (CPZ, a rVR1 antagonist) were prepared at 10x the desired final assay concentrations in FBP. The plate was flicked to remove LOAD from cells, and 180 μ l FBP was added per well. The FLIPR machine added 20 μ l capsaicin per well to give a final 1x concentration. Cells were monitored for 70 seconds after agonist addition. The FLIPR traces (fluorescence change (counts) versus time (seconds)) were produced for each well. Peaks indicate capsaicin-gated calcium influx, by cells expressing rVR1. The peak height correlates with the rVR1 expression level. To measure antagonism of the VR1 response 20 μ l 10x antagonist CPZ was added into wells to give a final 1x concentration. The plate was incubated for 15 minutes at room temperature prior reading in the FLIPR. The FLIPR traces recorded for each well show that the

peak heights are reduced in cells pre-incubated in CPZ. The same FLIPR assay may be used to monitor the response of human VR1 on exposure to agonists and antagonists.

5 **Example 8: Example of a screen using human VR1.**

FLIPR assay technology may be utilised to screen for hVR1 modulators according to the procedure described in figure 15. Human VR1 may be gated with protons, capsaicin or heat.

10 **Reference Example C: Identification and partial characterisation of additional human vanilloid receptors (reference examples C1-C3):**

Reference Example C1: Identification and characterisation of a novel vanilloid-like receptor, hVR3

15 ESTs belonging to the remaining clusters were characterised by *in silico* cloning (reference example A). The following clones were used during this process: - EST6/EST7 (hVRd), -EST8. (hVRe), - EST9/EST10. (hVRf). These EST clusters have been aligned with rat VR1 in figure 16, note that this diagram is not to scale.

20 **Reference Example C2: Sequencing of clones**

Further sequencing, as detailed in reference example B2, and *in silico* cloning, enabled clusters hVRd, hVRe and hVRf to collapse forming a single contig of 583 amino acids. This sequence was named hVR3 and has 49 % identity with the rat VR1 sequence. It was unlikely that this single contig was a full-length vanilloid receptor transcript as no obvious starting codon was present and it was shorter than the rat VR1 transcript.

25 **Reference Example C3: Identification of the 5' terminus of hVR3**

30 Two primers (sense primer 5' ATGGCCACCAGCAGGGTTAC and antisense primer 5' TCTGCCAGGTTCCAGCTG) designed to PCR amplify an amplicon stretching the 3' end of hVR3 and its 3'utr were used to isolate a genomic PAC clone (Genome Systems. St Louis, Missouri). The hVR3 specific PAC clone was then used as template to generate a library. This was achieved by sonicating 6 μ g of Qiagen purified PAC construct, size selecting fragmented DNA of 500-

2000bp. These resulting fragments were then blunt ended and cloned into the vector pCR®-Blunt as detailed in the manufacturers protocol supplied with the Zero Blunt™ PCR cloning kit (Invitrogen). Clones were then sequenced (reference example B2) to identify the complete 5' end of the hVR3 transcript.

5 The full-length nucleotide sequence of the hVR3 gene is displayed in figure 17. Figure 18 illustrates both nucleotide and encoded amino acid sequence of the human VR1 and figure 19 depicts the amino acid sequence of the hVR3 gene with shaded regions denoting predicted trans-membrane regions (boxed) and the ankyrin binding domains (unboxed).

10

Example 9: Full-length Amplification of hVR3 from human kidney template

Human kidney was used as a source of template for the PCR amplification of hVR3. Primers used for amplification were designed to isolate the gene in three fragments. Primers designed to isolate the 5' end included a sense primer encoding a NotI site and a strong Kozak motif followed by gene specific sequence (5' GTCATAGCGGCCGCGGCCACCATGCCAGGGTAGTTGGAC

15

and antisense primer (5' CACCTCTTGTGTCACTGGA). The PCR conditions used were a hot start at 94°C for 4 mins, followed by 35 cycles of 94°C for 1 min, 56°C for 1 min and 72°C for 1 min and finally one cycle at 72°C for 5 min. The

20

resulting PCR products were separated on a 2% agarose gel and cloned into pCR®II-TOPO according to the manufacturers instructions supplied with the TOPO™ TA Cloning® kit (Invitrogen). The middle fragment was PCR generated using sense and antisense primers 5' CAAATCTGCGCATGAAGTTCCAG and 5' GCCACGAGAAGTTCCACGTAGTG respectively in the presence of 5% DMSO.

25

PCR thermo-cycling required 35 cycles of 1 min at 94°C, 58°C and 72°C for successful amplification of the fragment which was then excised from a 2% agarose gel for cloning into the pCRII®-TOPO vector. Finally the 3' fragment was amplified with a sense primer 5' GCTGCTCCCATTCTGCTGA and an antisense primer 5' TGCACTCTCGAGAAATGAGTGGGCAGAGAAC encoding

30

a Xhol restriction site. This fragment was successfully amplified using a hot start at 94°C for 4 min followed by 35 cycles of 94°C for 50 sec, 48°C for 50 sec and 72°C for 2 min. The cycling was completed with a 72°C step for 5 min. The amplified fragment was excised from a 2% agarose gel and clone into the pCRII®-TOPO vector.

35

Resulting clones for each of the three PCR generated hVR3-fragments were taken for sequence analysis and separate clones coding a consensus sequence were used in the full-length assembly of the gene. The DralII restriction site of the pBluescript SK (+) vector (Stratagene) was firstly abolished by digestion with DralII followed by a blunt ending step using T₄ DNA polymerase (New England Biolabs). This modified vector was then restricted to enable the ligation of both a NotI/Ncol 5' fragment and Ncol/ EcoRI middle fragment. Finally, the remaining 3' fragment was introduced into the resulting construct via DralII and Xhol sites (figure 20A).

Several clones were selected for sequence analysis to confirm that the constructs still encoded the hVR3 consensus sequence. These were then digested with NotI/Xhol and ligated into the mammalian expression vector pCDNA3.1 (+) (Invitrogen) as seen in figure 20B. The resulting hVR3 consensus sequence is shown in the multiple alignment along with the full-length sequence of hVR1 and the published hVRL-1 in figure 21.

Example 10: Chromosomal localisation

The 3' terminus, including the 3' UTR sequence of hVR3 was used to design two primers to amplify a product of 360 bp: sense primer 5' ATGGCCACCAGCAGGGTTAC and antisense primer 5' TCTGCCAGGTTCCAGCTG. The G3 radiation hybrid panel from Stanford University (Research Genetics, Huntsville, Alabama) was screened by PCR. The positive and negative lanes were analysed using the public web server at Stanford University (<http://www-sghc.stanford.edu>). After analysis the hVR3 gene appears to be located on human chromosome 12 around markers D12S177E (lod score=15) and D12S1893 (lod score=14).

Example 11: mRNA distribution

The following primers (5' ACAAGAAGGCCGGACATGCGG and 5' ATCTCGTGGCGGTTCTCAAT) were used to obtain a PCR product from the coding region of hVR3. This amplicon was used as a probe on multi-tissue northern blots, the protocol of which is detailed in example 3, to determine the tissue distribution of the gene (figures 22A, 22B and 22C). A transcript of approximately 3.8 kb was detected in the following tissues (the intensities of the

signals are indicated in brackets): trachea (very strong), kidney (strong), pancreas (strong), prostate (strong), placenta (strong), bone marrow (weak), adrenal gland (weak), lymph node (weak), spinal cord (weak), thyroid (weak), stomach (weak), lung (weak) and liver (weak).

5 Since these commercial blots (Clontech, Palo Alto, California, USA) should have
the same amount of RNA it is interesting to note the very strong signal in the
trachea lane (figure 22A). This could indicate the potential of hVR3 as a target
for respiratory pathologies. It was shown by RT-PCR with the primer
10 combination used to produce the probe that the gene is not expressed in DRG.

Example 12: Riboprobe generation for the in situ localisation of hVR3

The same probe, which was specific to hVR3 in Northern blot analysis (example 11), was used to generate a riboprobe. This hVR3 specific probe was cloned into the T7 and SP6 encoding pCR^{II}-TOPO vector (Invitrogen). This construct was then used in the *in vitro* transcription of DIG labelled RNA strands from the vectors promoters as described in the manufacturers instructions as detailed in the DIG RNA labelling kit (Roche Molecular Biochemicals). This riboprobe may be used to identify the cellular localisation of hVR3 present in tissues such as trachea, lung, pancreas, prostate, placenta and kidney.

Example 13: Mammalian Cell Expression of hVR3

Expression of hVR3 may be accomplished by transfecting a mammalian cell line such as: HEK283T, HEK293, CHO, COS, HeLa and BHK. A detailed method for both transient and stable transfection is detailed in example 6.

Example 14: Functional Analysis of hVR3

The functional analysis of hVR3 may be studied using the electrophysiology, calcium imaging and FLIPR methods as detailed in examples 7a to 7c.

Example 15: Example of a drug screen using human VR3.

A stable cell line expressing hVR3 may be used in a drug screen such as a selectivity screen using test compounds that have been identified to have an agonistic or antagonistic action on hVR1. FLIPR assay technology may be utilised to screen for hVR3 modulators as proposed in figure 15.

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Changed the margins in cases where the sequence text was "wrapped" down to the next line.

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128	Gly	Arg	Pro	Gly	Phe	Tyr	Phe	Gly	Glu	Leu	Pro	Leu	Ser	Leu	Ala	Ala			
129		245				250					255								
131	tgc	acc	aac	cag	ctg	ggc	atc	gtg	aag	ttc	ctg	ctg	cag	aac	tcc	tgg		1593	
132	Cys	Thr	Asn	Gln	Leu	Gly	Ile	Val	Lys	Phe	Leu	Leu	Gln	Asn	Ser	Trp			
133		260			265				270										
135	cag	acg	gcc	gac	atc	agc	gcc	agg	gac	tcg	gtg	ggc	aac	acg	gtg	ctg		1641	
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137	275				280					285									
139	cac	gcc	ctg	gtg	gag	gtg	gcc	gac	aac	acg	gcc	gac	aac	acg	aag	ttt		1689	
140	His	Ala	Leu	Val	Glu	Val	Ala	Asp	Asn	Thr	Ala	Asp	Asn	Thr	Lys	Phe			
141	290				295					300							305		
143	gtg	acg	agc	atg	tac	aat	gag	att	ctg	atc	ctg	ggg	gcc	aaa	ctg	cac		1737	
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RAW SEQUENCE LISTING
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152	Leu	Ala	Leu	Ala	Ala	Gly	Thr	Gly	Lys	Ile	Gly	Val	Leu	Ala	Tyr	Ile	
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158	Leu	Gln	Arg	Glu	Ile	Gln	Glu	Pro	Glu	Cys	Arg	His	Leu	Ser	Arg	Lys	
159		355				360						365					
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162	Phe	Thr	Glu	Trp	Ala	Tyr	Gly	Pro	Val	His	Ser	Ser	Leu	Tyr	Asp	Leu	
163	370				375						380				385		
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166	Ser	Cys	Ile	Asp	Thr	Cys	Glu	Lys	Asn	Ser	Val	Leu	Glu	Val	Ile	Ala	
167				390				395							400		
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173	ccg	ctg	aac	cga	ctc	ctg	cag	gac	aag	tgg	gac	aga	ttc	gtc	aag	cgc	2073
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175		420				425						430					
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271	aga	cac	tgg	aag	aac	ttt	gcc	ctg	gtc	ccc	ttt	tta	aga	gag	gca	agt	3177	
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279	ttt	tca	ggg	tct	ctg	aag	cca	gag	gac	gtc	ttc	aag	agt	cct		3273		
280	Phe	Ser	Gly	Ser	Leu	Lys	Pro	Glu	Asp	Ala	Glu	Val	Phe	Lys	Ser	Pro		
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284	Ala	Ala	Ser	Gly	Glu	Lys												
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 293 atctcctaacc agactttcag gtttttactc actttactaa acagtttgcg tggctcagt 3564
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 317 atcagctcaa ttgctgtgcg ggttaaaaact acagaaccac atcccaaagg tacctggtaa 4224
 319 gaatgttgc aagatcttcc atttcttagga accccagtcg tgcttctccg caatggcaca 4284
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 335 Gln Lys Asp Thr Cys Pro Asp Pro Leu Asp Gly Asp Pro Asn Ser Arg
 336 20 25 30
 338 Pro Pro Pro Ala Lys Pro Gln Leu Ser Thr Ala Lys Ser Arg Thr Arg
 339 35 40 45
 341 Leu Phe Gly Lys Gly Asp Ser Glu Glu Ala Phe Pro Val Asp Cys Pro
 342 50 55 60
 344 His Glu Glu Gly Glu Leu Asp Ser Cys Pro Thr Ile Thr Val Ser Pro
 345 65 70 75 80
 347 Val Ile Thr Ile Gln Arg Pro Gly Asp Gly Pro Thr Gly Ala Arg Leu
 348 85 90 95
 351 Leu Ser Gln Asp Ser Val Ala Ala Ser Thr Glu Lys Thr Leu Arg Leu
 352 100 105 110
 354 Tyr Asp Arg Arg Ser Ile Phe Glu Ala Val Ala Gln Asn Asn Cys Gln
 355 115 120 125
 357 Asp Leu Glu Ser Leu Leu Leu Phe Leu Gln Lys Ser Lys Lys His Leu
 358 130 135 140
 360 Thr Asp Asn Glu Phe Lys Asp Pro Glu Thr Gly Lys Thr Cys Leu Leu
 361 145 150 155 160
 363 Lys Ala Met Leu Asn Leu His Asp Gly Gln Asn Thr Thr Ile Pro Leu
 364 165 170 175
 366 Leu Leu Glu Ile Ala Arg Gln Thr Asp Ser Leu Lys Glu Leu Val Asn
 367 180 185 190
 369 Ala Ser Tyr Thr Asp Ser Tyr Tyr Lys Gly Gln Thr Ala Leu His Ile
 370 195 200 205

VERIFICATION SUMMARY
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L:13 M:270 C: Current Application Number differs, Replaced Application Number
L:14 M:271 C: Current Filing Date differs, Replaced Current Filing Date

PCT09

RAW SEQUENCE LISTING
PATENT APPLICATION: US/09/857,123

DATE: 08/30/2001
TIME: 07:44:20

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Does Not Comply
Corrected Diskette Needed

4 <110> APPLICANT: Glaxo Group Ltd
5 Tate, Simon N
6 Delany, Natalie S
7 Sanseau, P
9 <120> TITLE OF INVENTION: Novel Receptors
11 <130> FILE REFERENCE: PG3606
C--> 13 <140> CURRENT APPLICATION NUMBER: US/09/857,123
C--> 14 <141> CURRENT FILING DATE: 2001-07-30
16 <150> PRIOR APPLICATION NUMBER: GB 9826359.3
17 <151> PRIOR FILING DATE: 1998-12-01
19 <160> NUMBER OF SEQ ID NOS: 40
21 <170> SOFTWARE: PatentIn Ver. 2.1

ERRORED SEQUENCES

1759 <210> SEQ ID NO: 40
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VERIFICATION SUMMARY
PATENT APPLICATION: US/09/857,123

DATE: 08/30/2001
TIME: 07:44:21

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L:14 M:271 C: Current Filing Date differs, Replaced Current Filing Date
L:1772 M:254 E: No. of Bases conflict, LENGTH:Input:1 Counted:20 SEQ:40

Claims

1. An isolated human vanilloid receptor (hVR) protein or a variant thereof.
- 5 2. An isolated human vanilloid receptor (hVR) protein according to claim 1 which is hVR1 or a variant thereof.
- 10 3. An isolated human vanilloid receptor (hVR) protein according to claim 1 which is hVR3 or a variant thereof.
- 15 4. An isolated human vanilloid receptor (hVR) protein according to claim 2 having an amino acid sequence as shown in Figure 3.
5. An isolated human vanilloid receptor (hVR) protein according to claim 3 having an amino acid sequence as shown in Figure 18.
- 20 6. A nucleotide sequence encoding a human vanilloid receptor (hVR) protein or a variant thereof, or a nucleotide sequence which is complementary thereto.
7. A nucleotide sequence according to claim 6 encoding for an hVR1 protein or a variant thereof, or a nucleotide sequence which is complementary thereto.
- 25 8. A nucleotide sequence according to claim 6 encoding for an hVR3 protein or a variant thereof, or a nucleotide sequence which is complementary thereto.
9. A nucleotide sequence according to claim 6 which is a cDNA sequence.
- 30 10. A nucleotide sequence according to claim 7 which is a cDNA sequence
11. A nucleotide sequence according to claim 8 which is a cDNA sequence

12. A nucleotide sequence according to claim 7 as shown in Figure 2.
13. A nucleotide sequence according to claim 8 as shown in Figure 17.
- 5 14. An expression vector comprising a nucleotide sequence according to any one of claims 6 to 13, which is capable of expressing an hVR protein or a variant thereof.
- 10 15. An expression vector according to claim 14 which is capable of expressing an hVR1 protein or a variant thereof.
16. An expression vector according to claim 14 which is capable of expressing an hVR3 protein or a variant thereof.
- 15 17. A stable cell line comprising an expression vector according to claim 14.
18. A stable cell line comprising an expression vector according to claim 15.
- 20 19. A stable cell line comprising an expression vector according to claim 16.
- 25 20. A stable cell line according to claim 17 which is a modified HEK293, CHO, COS, HeLa or BHK cell line.
21. A stable cell line according to claim 18 which is a modified HEK293, CHO, COS, HeLa or BHK cell line.
- 30 22. A stable cell line according to claim 19 which is a modified HEK293, CHO, COS, HeLa or BHK cell line.
23. An antibody specific for a human vanilloid receptor (hVR) protein or a variant thereof as claimed in any one of claims 1 to 5.

24. An antibody according to claim 23 which is specific for hVR1 or a variant thereof.

5 25. An antibody according to claim 23 which is specific for hVR3 or a variant thereof.

10 26. A method for identification of a compound which exhibits hVR modulating activity comprising contacting a human vanilloid receptor (hVR) protein or a variant thereof according to any one of claims 1 to 5 with a test compound and detecting modulating activity or inactivity.

15 27. A compound which modulates hVR activity, identifiable by a method according to claim 26.

20 28. A compound according to claim 27 for use in therapy.

25 29. The use of a compound according to claim 27 in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity in a human patient.

30 30. The use according to claim 28 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

35 31. A method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR activity in a human patient which comprises administering to said patient an effective amount of a compound according to claim 27.

32. A method according to claim 31 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain,

rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

5

33. A compound which modulates hVR activity, identifiable by a method according to claim 26, excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodal, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial, β -acaridial, scutigeral, merulidial, anandamide and capsazepine.

10

34. A compound according to claim 33 for use in therapy.

15

35. The use of a compound according to claim 33 in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity in a human patient.

20

36. The use according to claim 35 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

25

37. A method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR activity in a human patient which comprises administering to said patient an effective amount of a compound according to claim 33.

30

38. A method according to claim 37 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a

35

urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

39. A compound identified by the method according to claim 26.

5

40. A compound according to claim 39 for use in therapy.

10

41. The use of a compound according to claim 39 in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity in a human patient.

15

42. The use according to claim 41 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

20

43. A method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR activity in a human patient which comprises administering to said patient an effective amount of a compound according to claim 39.

25

44. A method according to claim 43 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

30

45. A method of producing an hVR protein or a variant thereof according to any one of claims 1-5 comprising introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVR protein or

a variant thereof, under conditions suitable for obtaining expression of the hVR protein or variant thereof.

46. A method of producing an hVR1 protein or a variant thereof comprising

5 introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVR1 protein or a variant thereof, under conditions suitable for obtaining expression of the hVR1 protein or variant thereof.

47. A method of producing an hVR3 protein or a variant thereof comprising

10 introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVR3 protein or a variant thereof, under conditions suitable for obtaining expression of the hVR3 protein or variant thereof.

48. A human vanilloid receptor (hVR) protein or a variant thereof for use in

15 a method of screening for agents useful in the treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity in a human patient

49. A human vanilloid receptor (hVR) protein according to claim 48

20 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

25

50. A human vanilloid receptor (hVR) protein according to claim 48 or 49 which is hVR1 or a variant thereof.

30 51. A human vanilloid receptor (hVR) protein according to claim 48 or 49 which is hVR3 or a variant thereof.

FIG. 1
ALIGNMENT OF HUMAN VR1 IN SILICO DERIVED CLUSTERS WITH RAT VR1

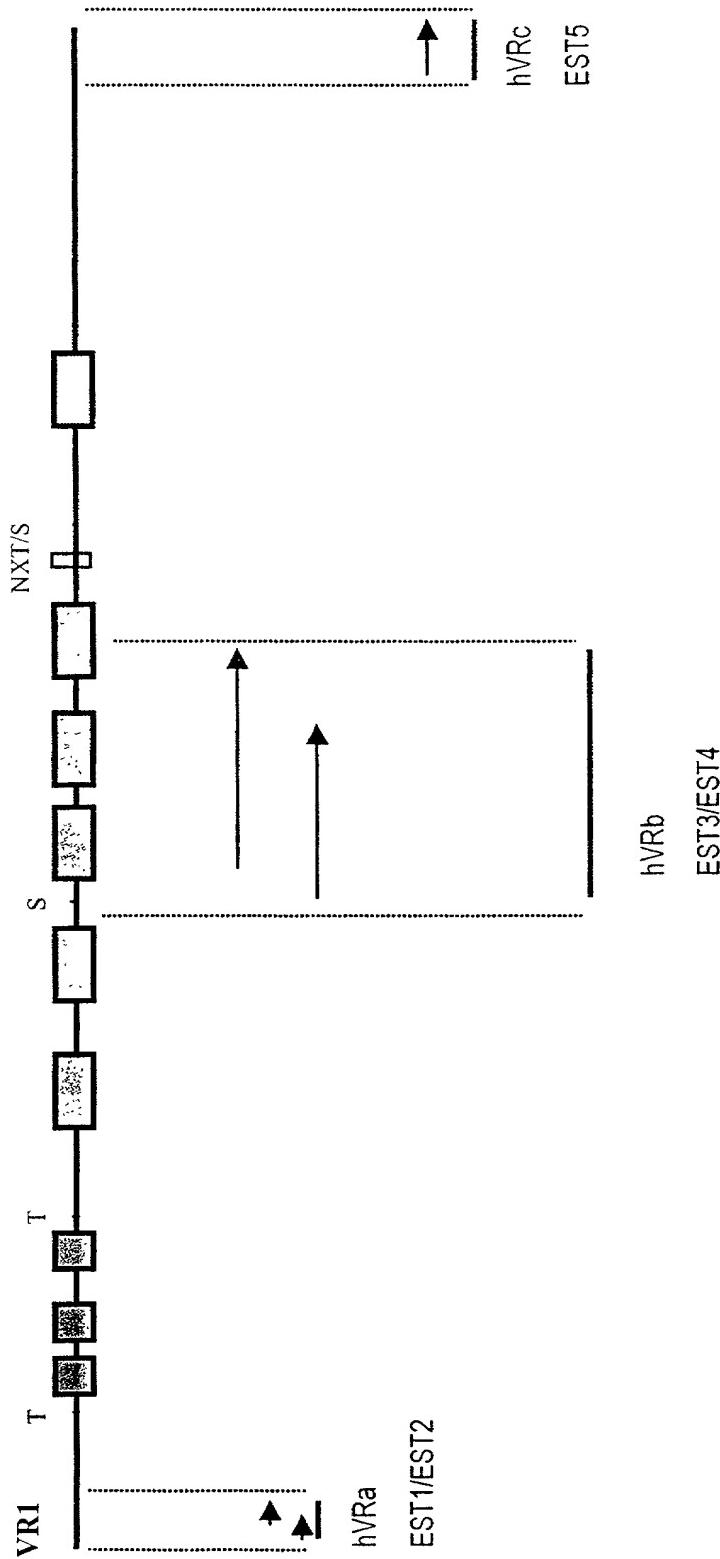


FIG. 2

hVR1 SEQUENCE INCLUDING THE 5'UTR (nt -773 TO nt 0), CODING REGION (nt 1 TO 2517) AND 3'UTR (nt 2518 TO nt 3560)

-773	cccccagccaggcttaaccattca	-714
-713	aaggccagaagcttgcacagatgttgcattcataaaaatgcaaaaggccaaaatccaaaatct	-654
-653	tgtataagctcagtggctgtggcagcgagggttgaagagcaaaggcaggccgggacacctgg	-594
-593	ctgatgatgtgtggacccgttgcacagcaggcccgcagtgcgggtgtgggtgtgggtggg	-534
-533	ccagtctctgccgctcacccattccaggacacagtcgtctggctttctggactgag	-474
-473	ccatcctcatcaccgagatcctccctgaattcagccacgacagccacccggccgttt	-414
-413	ccttgttctgtgtggaaaggaggcagcgcgggtggtatcaacctcaccctgcagaggag	-354
-353	gcacctgaggcccagagacgaggaggatgggtctaaccagaaccacagatggctctga	-294
-293	gcggggggctgtccaccctccaggccacgtcagtgccgcaggactgcctggccct	-234
-233	gttaggcctgctcacctctgaggcctctgggtgagagggttcagtcctggaaacacttca	-174
-173	gttctaggggctggggcagcagcaagtggagtttgggtaccctgcttcacaggc	-114
-113	ccttggcaaggaggcagggtgggtctaaggacaaggcagtcctactttggagtcaacc	-54
-53	ccggcgtggctgctgcagggtcacactggccacagaggatccagcaaggATGAAG	6
7	AAATGGAGCAGCACAGACTTGGGGCAGCTGCGGACCACTCCAAAAGGACACCTGCCA	66
67	GACCCCCCTGGATGGAGACCCCTAACCTCCAGGCCACCTCCAGCCAAGCCCCAGCTCTCCACG	126
127	GCCAAGAGCCGCACCCGGCTTTGGGAAGGGTGACTCGGAGGAGGCTTCCGGTGGAT	186
187	TGCCCTCACGAGGAAGGTGAGCTGGACTCCTGCCGACCACAGTCAGCCCTGTTATC	246
247	ACCATCCAGAGGCCAGGAGACGGCCCCACCGGTGCCAGGCTGCTGTCCCAGGACTCTGTC	306

307	GCCGCCAGCACCGAGAAGACCCCTCAGGCTCTATGATGCAGGAGTATCTTGAAGCCGTT	366
367	GCTCAGAATAACTGCCAGGATCTGGAGAGCCTGCTGCTCTTCCTGCAGAAGAGCAAGAAG	426
427	CACCTCACAGACAACGAGTTCAAAGACCCCTGAGACAGGGAAAGACCTGTCTGCTGAAAGCC	486
487	ATGCTCAACCTGCACGACGGACAGAACACCACATCCCCCTGCTCCTGGAGATCGCGCGG	546
547	AAACGGACAGCCTGAAGGAGCTTGTCAACGCCAGCTACACGGACAGCTACTACAAGGGC	606
607	CAGACAGCACTGCACATGCCATCGAGAGACGCAACATGGCCCTGGTACCCCTGGTG	666
667	GAGAACGGAGCAGACGTCCAGGCTGCAGGCCATGGGGACTTCTTAAGAAAACCAAAGGG	726
727	CGGCCTGGATTCTACTTCGGTGAAC TGCCCCTGTCCTGGCCCGTGCACCAACCAGCTG	786
787	GGCATCGTGAAGTTCCCTGCTGCAGAACTCCCTGGCAGACGGCCGACATCAGGCCAGGGAC	846
847	TCGGTGGCAACACGGTGCTGCACGCCCTGGTGGAGGTGGCCGACAACACGGCCGACAAC	906
907	ACGAAGTTGTGACGGAGCATGTACAATGAGATTCTGATCCTGGGGCCAAACTGCACCCG	966
967	ACGCTGAAGCTGGAGGAGCTACCAACAAGAAGGGATGACGCCGCTGGCTCTGGCAGCT	1026
1027	GGGACCGGAAAGATCGGGTCTTGGCTATATTCTCCAGCGGGAGATCCAGGAGCCGAG	1086
1087	TGCAGGCACCTGTCCAGGAAGTTACCGAGTGGCCCTACGGCCGTGCACCTCGCTG	1146
1147	TACGACCTGTCCTGCATCGACACCTGCGAGAAGAACTCGGTGCTGGAGGTGATGCCCTAC	1206
1207	AGCAGCAGCGAGACCCCTAACGCCACGACATGCTTGGTGGAGCCGCTGAACCGACTC	1266
1267	CTGCAGGACAAGTGGACAGATTCTGTCAGCGCATTTCTACTTCAACTCCTGGTCTAC	1326
1327	TGCCTGTACATGATCATCTTACCATGGCTGCCTACTACAGGCCGTGGATGGCTTGCCT	1386
1387	CCCTTTAAGATGGAAAAAATTGGAGACTATTCGAGTTACTGGAGAGATCCTGTCTGTG	1446

FIG. 2 CONT'D

1447	TTAGGAGGAGTCTACTTCTTTCCGAGGGATTCACTATTTCTGCAGAGGCCGGCGTCG	1506
1507	ATGAAGACCCCTGTTGTGGACAGCTACAGTGAGATGCTTTCTGCAGTCAGTGTTC	1566
1567	ATGCTGGCCACCGTGGTGCTGTACTTCAGCCACCTCAAGGAGTATGTGGCTTCATGGTA	1626
1627	TTCTCCCTGGCCTGGGCTGGACCAACATGCTCTACTACACCCGGTTCCAGCAGATG	1686
1687	GGCATCTATGCCGTATGATAAGAAGATGATCCTGAGAGACCTGTGCCGTTCATGTT	1746
1747	GTCTACATCGTCTTCTTGTTCGGTTTCCACAGCGGTGGTGACGCTGATTGAAGACGGG	1806
1807	AAGAATGACTCCCTGCCGTCTGAGTCCACGTCGCACAGGTGGCGGGGCCTGCCTGCAGG	1866
1867	CCCCCGATAGCTCCTACAACACGCTGTACTCCACCTGCCTGGAGCTGTTCAAGTTCACC	1926
1927	ATCGGCATGGCGACCTGGAGTTCACTGAGAACTATGACTTCAAGGCTGTCTTCATCATH	1986
1987	CTGCTGCTGGCTATGTAATTCTCACCTACATCCTCCTGCTAACATGCTCATGCCCTC	2046
2047	ATGGGTGAGACTGTCAACAAGATCGCACAGGAGAGCAAGAACATCTGAAAGCTGCAGAGA	2106
2107	GCCATCACCATCCTGGACACGGAGAAGAGCTTCCTTAAGTGCATGAGGAAGGCCTCCGC	2166
2167	TCAGGCAAGCTGCTGCAGGTGGGTACACACCTGATGGCAAGGACGACTACCGGTGGTGC	2226
2227	TTCAGGGTGGACGAGGTGAACCTGGACCACTGGAACACCAACGTGGCATTCAACGAA	2286
2287	GACCCGGCAACTGTGAGGGCGTCAAGCGCACCTGAGCTTCTCCCTGCGGTCAAGCAGA	2346
2347	GTTTCAGGCAGACACTGGAAGAACTTGGCCCTGGTCCCCCTTTAAGAGAGGCAACTGCT	2406
2407	CGAGATAGGCAGTCTGCTCAGCCCGAGGAAGTTATCTGCGACAGTTTCAGGGTCTCTG	2466
2467	AAGCCAGAGGACGCTGAGGTCTCAAGAGTCCTGCCGTTCCGGGAGAACGtggacgt	2526
2527	cacgcagacagcactgtcaacactgggccttaggagaccccggtgccacgggggctgct	2586

FIG.2 CONT'D

2587	gagggaacaccagtgtctgtcagcagccctggcctggctgtgcctgccagcatgttcc	2646
2647	caaatctgtgctggacaagctgtggaaagcgtttttggaaagcatggggagtgtatgtacat	2706
2707	ccaaccgtcactgtccccaaagtgaatctoctaacagactttcaggttttactcaactta	2766
2767	ctaaacagttggatggtcagttctactggacatgttaggccttggattttttttgatt	2826
2827	ttatttttctgtgagacagagttcacttttgcggcaggctggagtgcagtgggttg	2886
2887	atcttggctcactgcaacctctgtccgggttcaagcgatttttgcaggcttc	2946
2947	aagtagcttggattacaggtgagcactaccacgccccggctaattttgtattttaatag	3006
3007	agacggggtttcacatgttggccaggctggctcgaacttttgcacgtgatctgc	3066
3067	ccgccttggcctccaaaagtgtggattacaggtgtgagccgctgcgcctggccttctt	3126
3127	tgattttatattattaggagcaaaagtaaaatgaagcccaggaaaacaccttggaaacaa	3186
3187	actttcccttgcattttgcacggcccttctctgtgcgcgtgccttctt	3246
3247	acctggccgggtgggtgggtgtttccctggagaagatgggggaggctg	3306
3307	tcccactcccagcttgcagaatcaagctgtgcagcagtgccttcatcctt	3366
3367	acgatcaatcacagtcctccagaagatcagctcaattgtgtgcaggtaaaactacagaa	3426
3427	ccacatccaaaaggtaacctggtaagaatgtttgaaagatttccatccatggaaacccca	3486
3487	gtcctgcttctccgaatggcacatgttccactccatccatactggcatcctcaaataa	3546
3547	acagatatgtataaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaa	3591

FIG. 2 CONT'D

FIG. 3

NUCLEOTIDE AND AMINO ACID SEQUENCE OF hVR1 INCLUDING
THE 5'UTR (nt -773 TO nt 0), CODING REGION (nt 0 TO 2517) AND
3'UTR (nt 2518 TO nt 3560)

-773	cccccagccacacacacacacacacacacacacacacacacacacaggcttaaccattca	-714
-713	aaggccagaagcttgcacagatgttattcataaaaatgcaaaagccaaaatccaaaatct	-654
-653	tgtataagctcagtggctgtggcagcgaggttgaagagcaaaggcaggccggcacctgg	-594
-593	ctgatgtatgtgtggacccgttgcacagcagggccgcagtgcggtgtgggtgtgggtgg	-534
-533	ccagtctctgccgctcaccctattccaggacacagtctgcttggctttctggactgag	-474
-473	ccatcctcatcaccgagatcctccctgaattcagcccacgacagccacccggccgttt	-414
-413	ccttgttctgtgtggaaaggaggcagcgcggtggttatcaacctcaccctgcagaggag	-354
-353	gcacctgaggcccagagacgaggaggatgggtctaaccacagaaccacagatggctctga	-294
-293	gccgggggcctgtccaccctcccaggccacgtcagtggccgcaggactgcctggccct	-234
-233	gctaggcctgctcacctctgaggcctctgggtgagagttcagtcctggaaacacttca	-174
-173	gttctaggggctggggcagcagcaagttggagttttgggtaccctgcttcacaggc	-114
-113	ccttggcaaggaggcaggtgggtctaaggacaagcagtccattacttggagtcacc	-54
-53	ccggcgtggctgcaggttcacactggccacagaggatccagcaaggATGAAG	6
1	M K	2
7	AAATGGAGCAGCACAGACTTGGGGCAGCTGCGGACCCACTCCAAAAGGACACCTGCCA	66
3	K W S S T D L G A A A D P L Q K D T C P	22
67	GACCCCCCTGGATGGAGACCCTAACCTCCAGGCCACCTCCAGCCAAGCCCCAGCTCTCCACG	126
23	D P L D G D P N S R P P P A K P Q L S T	42
127	GCCAAGAGCCGCACCCGGCTTTGGAAAGGGTGAECTCGGAGGAGGCTTCCGGTGGAT	186
43	A K S R T R L F G K G D S E E A F P V D	62
187	TGCCCTCACGAGGAAGGTGAGCTGGACTCCTGCCGACCATCACAGTCAGCCCTGTTATC	246
63	C P H E E G E L D S C P T I T V S P V I	82
247	ACCATCCAGAGGCCAGGAGACGGCCCCACCGGTGCCAGGCTGCTGTCCCAGGACTCTGTC	306
83	T I Q R P G D G P T G A R L L S Q D S V	102
307	GCCGCCAGCACCAGAGAACCCCTCAGGCTCTATGATGCAGGAGTATCTTGAGCCGTT	366
103	A A S T E K T L R L Y D R R S I F E A V	122
367	GCTCAGAATAACTGCCAGGATCTGGAGAGCCTGCTGCTCTCCTGCAGAAGAGCAAGAAG	426
123	A Q N N C Q D L E S L L L F L Q K S K K	142
427	CACCTCACAGACAACGAGTTCAAAGACCCCTGAGACAGGAAAGACCTGTCTGCTGAAAGCC	486
143	H L T D N E F K D P E T G K T C L L K A	162
487	ATGCTCACCTGCACGGACAGAACACCACATCCCCCTGCTCCTGGAGATCGCGCGG	546

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163	M L N L H D G Q N T T I P L L L E I A R	182
547	CAAACGGACAGCCTGAAGGAGCTTGTCAACGCCAGCTACACGGACAGCTACTACAAGGGC	606
183	Q T D S L K E L V N A S Y T D S Y Y K G	202
607	CAGACAGCACTGCACATGCCATCGAGAGACGCAACATGGCCCTGGTGACCCCTGGTG	666
203	Q T A L H I A I E R R N M A L V T L L V	222
667	GAGAACGGAGCAGACGTCCAGGCTGCGGCCATGGGACTTAAAGAAAACCAAAGGG	726
223	E N G A D V Q A A A H G D F F K K T K G	242
727	CGGCCTGGATTCTACTTCGGTGAAC TGCCCTGTCCCTGGCCGCGTGCACCAACCAGCTG	786
243	R P G F Y F G E L P L S L A A C T N Q L	262
787	GGCATCGTGAAGTTCTGCTGCAGAACTCCTGGCAGACGGCCGACATCAGGCCAGGGAC	846
263	G I V K F L L Q N S W Q T A D I S A R D	282
847	TCCGTGGCAACACGGTGCACGCCCTGGTGGAGGTGGCCGACAACACGGCCGACAAC	906
283	S V G N T V L H A L V E V A D N T A D N	302
907	ACGAAGTTGTGACGAGCATGTACAATGAGATTCTGATCCTGGGGCCAAACTGCACCCG	966
303	T K F V T S M Y N E I L I L G A K L H P	322
967	ACGCTGAAGCTGGAGGAGCTACCAACAAGAAGGAATGACGCCGCTGGCTCTGGCAGCT	1026
323	T L K L E E L T N K K G M T P L A L A A	342
1027	GGGACCGGAAAGATCGGGTCTGGCTATATTCTCCAGGGGAGATCCAGGAGCCGAG	1086
343	G T G K I G V L A Y I L Q R E I Q E P E	362
1087	TGCAGGCACCTGTCCAGGAAGTCACCGAGTGGCCCTACGGGCCGTGCACTCCTCGCTG	1146
363	C R H L S R K F T E W A Y G P V H S S L	382
1147	TACGACCTGTCCTGCATCGACACCTGCGAGAAAGAAACTCGGTGCTGGAGGTGATGCC	1206
383	Y D L S C I D T C E K N S V L E V I A Y	402
1207	AGCAGCAGCGAGACCCCTAACGCCACGACATGCTTGGTGGAGCCGCTGAACCGACTC	1266
403	S S S E T P N R H D M L L V E P L N R L	422
1267	CTGCAGGACAAGTGGACAGATTGTCAAGGCATCTCTACTTCAACTTCCCTGGTCTAC	1326
423	L Q D K W D R F V K R I F Y F N F L V Y	442
1327	TGCCTGTACATGATCATCTCACCATGGCTGCCTACTACAGGCCGTGGATGGCTGCCT	1386
443	C L Y M I I F T M A A Y Y R P V D G L P	462
1387	CCCTTTAAGATGGAAAAATTGGAGACTATTCGAGTTACTGGAGAGATCCTGTCTG	1446
463	P F K M E K I G D Y F R V T G E I L S V	482
1447	TTAGGAGGAGTCTACTTCTTCCGAGGGATTCACTGAGATGCTTTCTGCAGAGGCCGCG	1506
483	L G G V Y F F F R G I Q Y F L Q R R P S	502
1507	ATGAAGACCCCTGTTGTGGACAGCTACAGTGAGATGCTTTCTGCAGTCAGTCAGT	1566
503	M K T L F V D S Y S E M L F F L Q S L F	522
1567	ATGCTGGCCACCGTGGTGTACTTCAGCCACCTCAAGGAGTATGTGGCTCCATGGTA	1626
523	M L A T V V L Y F S H L K E Y V A S M V	542
1627	TTCTCCCTGGCCTTGGCTGGACCAACATGCTACTACACCCGGTTCCAGCAGATG	1686

FIG. 3 CONT'D

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543	F S L A L G W T N M L Y Y T R G F Q Q M	562
1687	GGCATCTATGCCGTATGATAGAGAAGATGATCCTGAGAGACCTGTGCCGTTCATGTTT	1746
563	G I Y A V M I E K M I L R D L C R F M F	582
1747	GTCTACATCGTCTTCTTGTTCGGTTTCCACAGCGGTGGTACGCTGATTGAAGACGGG	1806
583	V Y I V F L F G F S T A V V T L I E D G	602
1807	AAGAATGACTCCCTGCCGTCTGAGTCCACGTGACAGGTGGCGGGGCCTGCCTGCAGG	1866
603	K N D S L P S E S T S H R W R G P A C R	622
1867	CCCCCGATAGCTCCTACAACAGCCTGTACTCCACCTGCCCTGGAGCTGTTCAAGTTCAC	1926
623	P P D S S Y N S L Y S T C L E L F K F T	642
1927	ATCGGCATGGCGACCTGGAGTTCACTGAGAACTATGACTTCAAGGCTGTTCATCATC	1986
643	I G M G D L E F T E N Y D F K A V F I I	662
1987	CTGCTGCTGGCCTATGTAATTCTCACCTACATCCTCCTGCTAACATGCTCATGCCCTC	2046
663	L L L A Y V I L T Y I L L L N M L I A L	682
2047	ATGGGTGAGACTGTCAACAAGATCGCACAGGAGACCAAGAACATCTGGAAGCTGCAGAGA	2106
683	M G E T V N K I A Q E S K N I W K L Q R	702
2107	GCCATACCACATCCTGGACACGGAGAAGAGCTTCCTTAAGTGCATGAGGAAGGCCTCCGC	2166
703	A I T I L D T E K S F L K C M R K A F R	722
2167	TCAGGCAAGCTGCTGCAGGTGGGTACACACCTGATGGCAAGGACGACTACCGTGTTGC	2226
723	S G K L L Q V G Y T P D G K D D Y R W C	742
2227	TTCAGGGTGGACGAGGTGAACGGACACCTGGAACACCAACGTGGCATCATCAACGAA	2286
743	F R V D E V N W T T W N T N V G I I N E	762
2287	GACCCGGCAACTGTGAGGGCGTCAAGCGCACCTGAGCTTCCTGCCGTCAAGCAGA	2346
763	D P G N C E G V K R T L S F S L R S S R	782
2347	GTTCAGGCAGACACTGGAAGAACCTTGCCTGGTCCCCCTTTAAGAGAGGCAAGTGT	2406
783	V S G R H W K N F A L V P L L R E A S A	802
2407	CGAGATAGGCAGTCTGCTCAGCCCCGAGGAAGTTATCTGCGACAGTTTCAGGGTCTCTG	2466
803	R D R Q S A Q P E E V Y L R Q F S G S L	822
2467	AAGCCAGAGGACGCTGAGGTCTCAAGAGTCCTGCCGTTCCGGGAGAACGtggacgt	2526
823	K P E D A E V F K S P A A S G E K	839
2527	cacgcagacagcactgtcaacactgggccttaggagaccccggtgccacgggggtgtct	2586
2587	gagggaacaccagtgtctgtcagcagcctggcctggctgtgcctcccagcatgttcc	2646
2647	caaatctgtgctggacaagctgtggaaagcgttcttggaaagcatggggagtgtatgtacat	2706
2707	ccaaccgtcactgtccccaaagtgaatctctaacaagactttcaggttttactcacttta	2766
2767	ctaaacagttggatggtcagtctactggacatgttagggcccttgatttttttgcatt	2826
2827	ttattctttctgtgagacagagttcactcttggccaggctggagtgcagtgggtgtg	2886
2887	atcttggctcactgcaacctgtctgtcccggttcaagcgattttctgtttcagtcctccc	2946

FIG. 3_{CONT'D}

SUBSTITUTE SHEET (RULE 26)

2947 aagttagcttggattacaggtgaggcactaccacgccccggctaattttgtatTTtaatAG 3006
3007 agacggggtttcaccatgttggccaggctggctcgaaactcttgacctcaggtgatctgc 3066
3067 ccgccttggcctcccaaagtgcgtggattacaggtgtgagccgctgcgctcggccttctt 3126
3127 tgattttatattattaggagcaaaagtaaatgaagcccaggaaaacacccTTggaaacAA 3186
3187 actcttccttgcgtggaaaatgcagaggcccttcctctgtgccgtgcttgctccttctt 3246
3247 acctgcccggtgggttgggggtgtgggtttactccctggagaagatgggggaggctg 3306
3307 tccccactcccagctctggcagaatcaagctgtgcagcagtgccttcttcattccttc 3366
3367 acgatcaatcacagtctccagaagatcagctcaattgtgtgcaggttaaaactacagaa 3426
3427 ccacatccaaaggtaacctggtaagaatgtttgaaagatcttccatttcttaggaacccca 3486
3487 gtcctgcttctccgcaatggcacatgctccactccatccatactggcatcctcaaataa 3546
3547 acagatatgtataaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaa 3591

FIG. 3 CONT'D

FIG. 4

AMINO ACID SEQUENCE OF hVR1

1 MKKWSSTD LG AAADPLQKDT CPDPLGD P N SRPPPAKPQL STAKSRTRLF
 51 GKGDSEEA FP VDCPHEEGEL DSCPTITVSP VITIQRPGDG PTGARLLSQD
 101 SVAAS TEKTL RLYDRRSIFE AVAQNNCQL E SLLLFLQKS KKHL TDNEFK
 151 DPETGKTCLL KAMLNLHD GQ NTTIPLL E I ARQTDSLKE L VNASYTDSYY
 201 KGQTALHIAI ERRNMALV TL LVENGADVQA AAH GDFFKKT KGRPGFY FGE
 251 LPLSLAACTN QLGIVKFLLQ NSWQTADISA RD S VGN TVLH ALVEVADNTA
 301 DNTKFVTSMY NEILILGAKL HPTLKLEELT NH KGMTPLAL AAGTGKIGVL
 351 AYILQREIQE PECRHLSRKF TEWAYGPVHS SLYDLSCIDT CEKNSVLEVI
 401 AYSSSETPNR HDMLLVEPLN RLLQDKWDRF VKR K I F Y C L Y M E F T H
 451 MAAYY RPVDG LPPFKMEKIG DYFRVTGEI D SVLGGVYFFF RGIOY FLQRR
 501 PSMKTLFV I S E Y S E M P E L O S I H M I A T V Y I E E S H LKEYVAS M V E S E A T G W I
 551 N M P Y V T R G E O T O M G I Y A V M I E K M I L R D T C R E M F V Y I V E E F G F F S T A V V T L I E
 601 DGKNDSL PSE STSHWRGP A CRPPDSSY NS LYSTCLELFK FTIGMDLE F
 651 T E N Y D E K A V F T I I I I I A Y V I L E T Y I L E T N M I T E A L M G E T V N K I A Q E S K N I W K L
 701 Q R A I T I L D T E K S F L K C M R K A F R S G K L L Q V G Y T P D G K D D Y R W C F R V D E V N W
 751 T T W N T N V G I I N E D P G N C X G V K R T L S F S L R S S R V S G R H W K N F A L V P L L R E A
 801 S A R D R Q S A Q P E E V Y L R Q F S G S L K P E D A E V F K S P A A S G E K *

Key

T/S predicted phosphorylation sites

 Transmembrane domains

 Ankyrin binding domains

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FIG. 5

COMPARISON OF THE AMINO ACID SEQUENCE OF THE RAT (VR1)
AND HUMAN (hVR1) VANILLOID PROTEINS.

VR1	MEQRASLDSEESPPQENSCLDPDDRDPNCKPPVKPHIFTTRSRTRLF	10	20	30	40	50	
hVR1	MKKWSSTDLGAAADPLQKDTCPDPLGDPNSRPPPAPQLSTAKSRTRLF		60	70	80	90	100
VR1	GKGDSEEASPLDCPYEEGGLASCPIITVSSVLTIQRPGDGPASVRPSSQD						
hVR1	GKGDSEEAFPVDCPHEEGELDSCPTITVSPVITIQRPGDGPAGARLLSQD						
VR1	SVSAG.EKPPRPLYDRRSIFDAVAQSNQCELESLLPFLQRSKRLTDSEFK	110	120	130	140	150	
hVR1	SVAASSTEKTLRLYDRRSIFEAVAQNNCQDLESLLLFLQSKKKHLTDNEFK	160	170	180	190	200	
VR1	DPETGKTCLLKAMLNHNGQNDTIALLLDVARAKTDSLKQFVNASYTDSDYY						
hVR1	DPETGKTCLLKAMLNHHDGQNTTIPLLLEIARQTDLSLKELVNASYTDSDYY	210	220	230	240	250	
VR1	KGQTALHIAIERRNMTLVTLLENGADVQAAANGDFFKTKGRPGFYFGE						
hVR1	KGQTALHIAIERRNMAVTLLENGADVQAAAHGDFFKTKGRPGFYFGE	260	270	280	290	300	
VR1	LPLSLAACTNQLAIVKFILLQNSWQPADISARDSVGNTVLHALVEADNTV						
hVR1	LPLSLAACTNQLGIVKFILLQNSWQTADISARDSVGNTVLHALVEADNTV	310	320	330	340	350	
VR1	DNTKFVTSMYNEILILGAKLHPTLKEEITNRKGTLPLALAASSGKIGVL						
hVR1	DNTKFVTSMYNEILILGAKLHPTLKEELTNKKGMTPLALAAGTGKIGVL	360	370	380	390	400	
VR1	AYILQREIHEPECRHLRSRKFTEWAYGPVHSSLYDLSCIDTCEKNSVLEVI						
hVR1	AYILQREIQEPECRHLRSRKFTEWAYGPVHSSLYDLSCIDTCEKNSVLEVI	410	420	430	440	450	
VR1	AYSSSETPNRHDMLLVEPLNRLIQLDKWDRFVKRIFYFNFFVYCLYMIIFT						
hVR1	AYSSSETPNRHDMLLVEPLNRLIQLDKWDRFVKRIFYFNFLVYCLYMIIFT	460	470	480	490	500	
VR1	AAAYYRPVEGLPPYKLKWTVGDFRVTGEILSVSGGVYFFFRRGIQYFLQR						
hVR1	MAAYYRPVDGLPPFKMEK.IGDYFRVTGEILSVLGGVYFFFRRGIQYFLQR	510	520	530	540	550	
VR1	RPSLKSLFVDSYSEILFEVOSLFMLVSVVLYFSQRKEYVASMVFSLAMGW						
hVR1	RPSMKTIFVDSYSEMLFFLQSLFMLATVVLYFSHLKEYVASMVFSLALGW	560	570	580	590	600	
VR1	TNMLYYTRGFQOMGIYAVMIEKMLRDLCKRFMFVYLVFLFGFSTAVVTLI						
hVR1	TNMLYYTRGFQOMGIYAVMIEKMLRDLCKRFMFVYIVFLFGFSTAVVTLI	610	620	630	640	650	
VR1	EDGKNNSLPMESTPHKCRGSACK.PGNSYNSLYSTCLELFKFTIGMDLE						
hVR1	EDGKNDSLPESTSHWRWRGPACRPPDSSYNSLYSTCLELFKFTIGMDLE	660	670	680	690	700	
VR1	FTE NYDFKAVFIILLAYVILTYIILLNLMLIALMGETVNKIAQESKNIWK						
hVR1	FTE NYDFKAVFIILLAYVILTYIILLNLMLIALMGETVNKIAQESKNIWK	710	720	730	740	750	
VR1	LQRAITILDTEKSFLKCMRKAFRSGKLLQVGTPDGKDDYRWCFRVDEVN						
hVR1	LQRAITILDTEKSFLKCMRKAFRSGKLLQVGTPDGKDDYRWCFRVDEVN	760	770	780	790	800	
VR1	WTWNTNVGIINEDPGNCEGVKRTLSFSLRSGRVSGRNWKNFALVPLL RD						
hVR1	WTWNTNVGIINEDPGNCEGVKRTLSFSLRSSRVSGRHWNFALVPLL RE	810	820	830			
VR1	ASTRDRHATQQEVOLKHYTGSILKPEDAEVFKDSMVPGEK						
hVR1	ASARDQSAQPEEVYLQFSGSLKPEDAEVFKSPAASGEK						

FIG. 6

FULL-LENGTH hRV1 CLONED INTO (A) pBLUESCRIPT SK(+) (hVR1pBSK)
AND (B) pCIN5-NEW (hVR1pCIN5) VIA NotI/EcoRI RESTRICTION SITES.

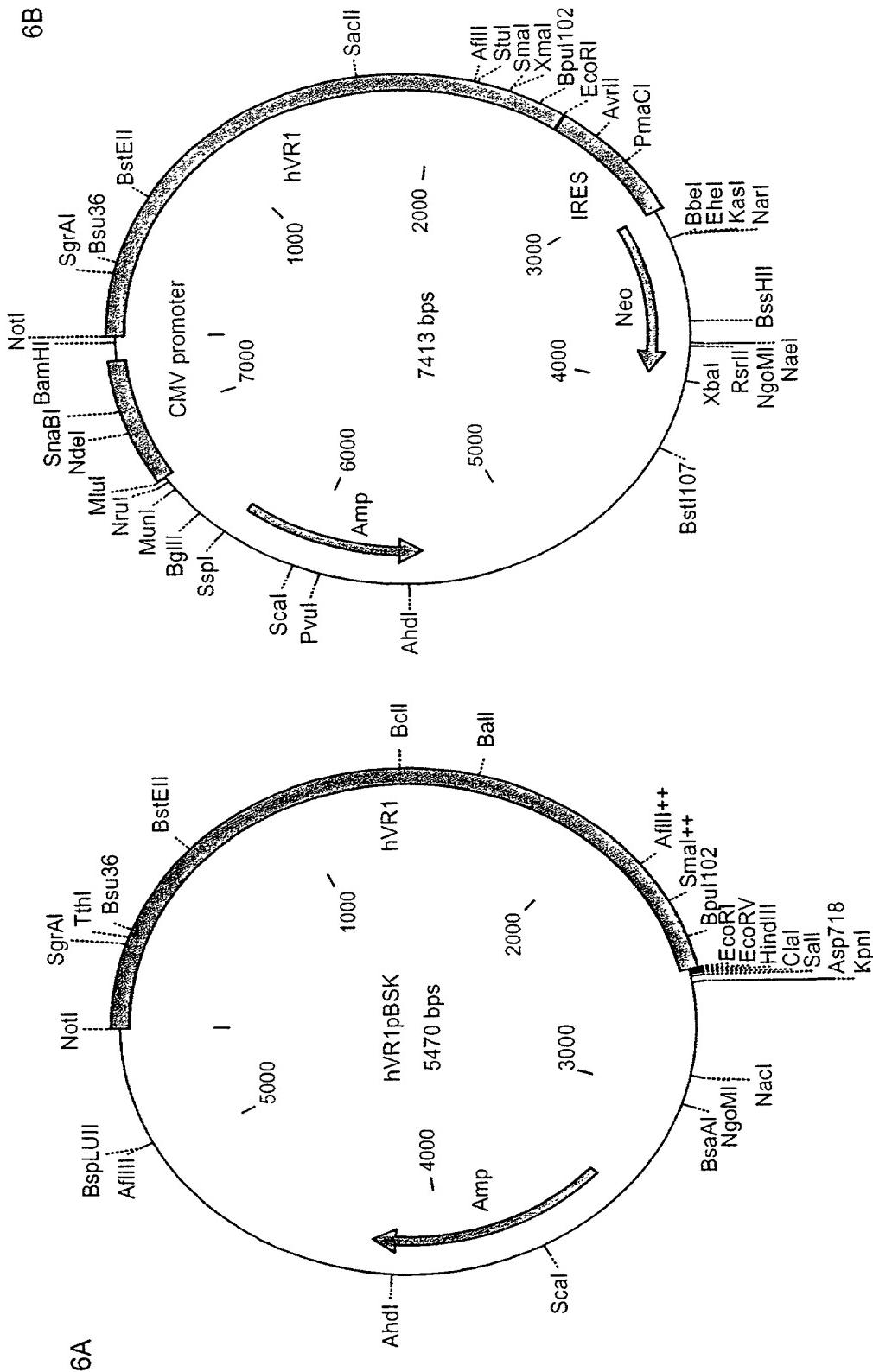
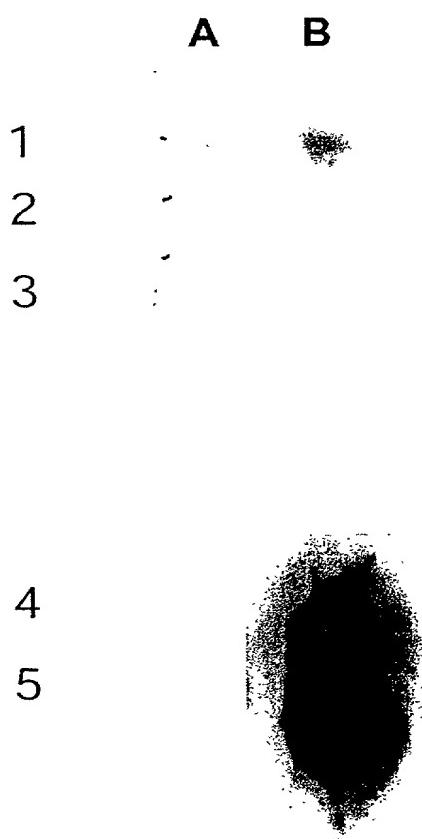


FIG. 7
SLOT HYBRIDISATION WITH hVR1 PROBE



Well

1A hDRG
2A rDRG

1B hDRG

3A Water

4B EST3 clone

5B 260bp Amplicon from Brain cDNA

FIG. 8

WESTERN BLOTH PROBED WITH ANTI-hVR1 ANTIBODIES.
ARROW POINTS TO hVR1 SPECIFIC BAND

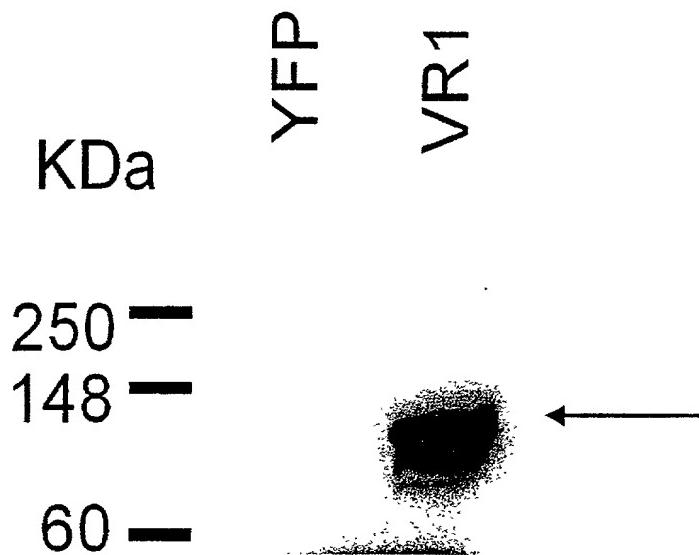
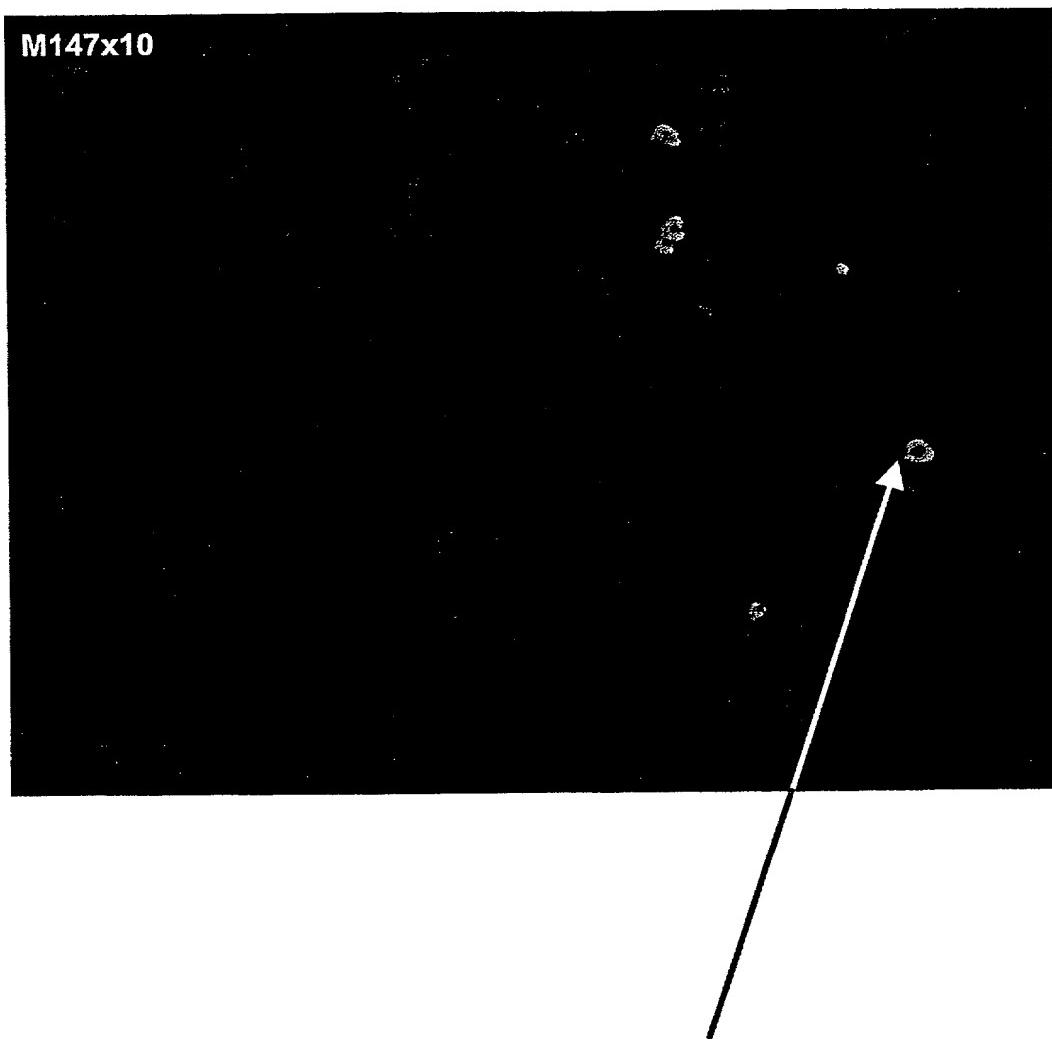


FIG. 9

IN SITU LOCALISATION OF VR1 IN RAT DRG TISSUE SECTIONS.
ARROW POINTS TO A VR1 EXPRESSING SMALL DIAMETER
($<25\mu\text{m}$) NEURONE CELL BODY, MAGNIFICATION USED 147x10.



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FIG. 10A

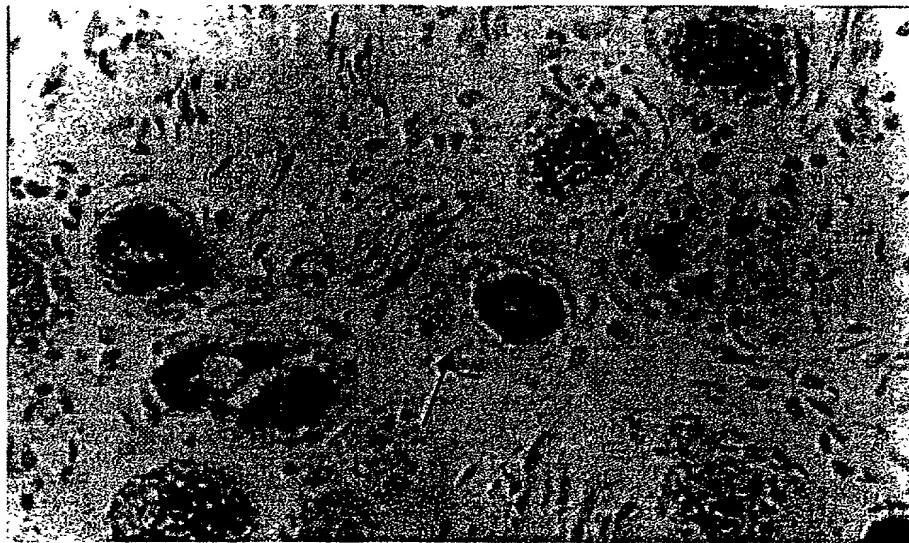


FIG. 10B

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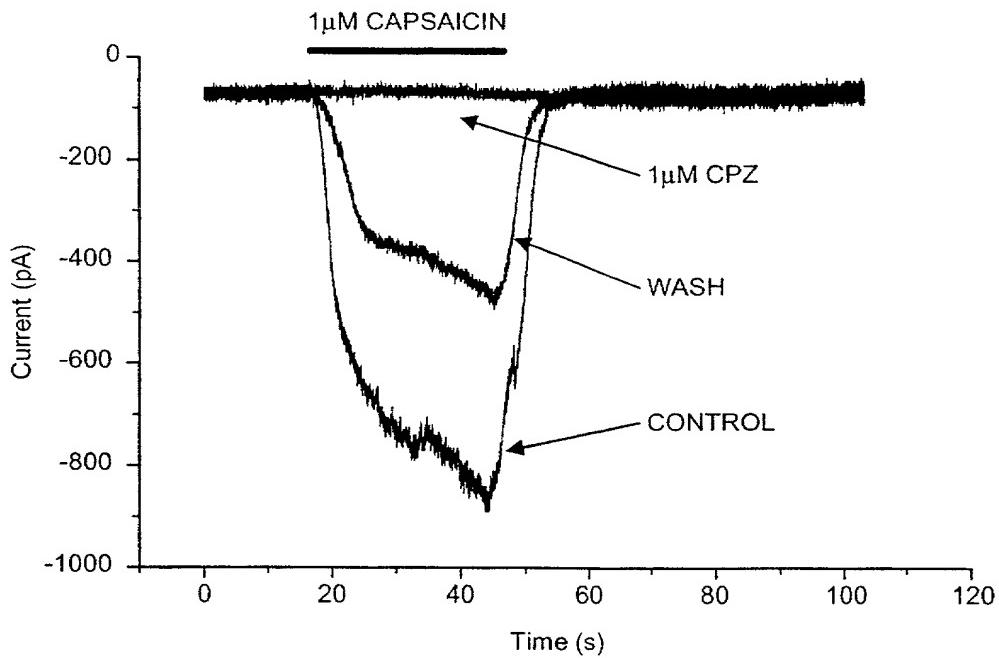
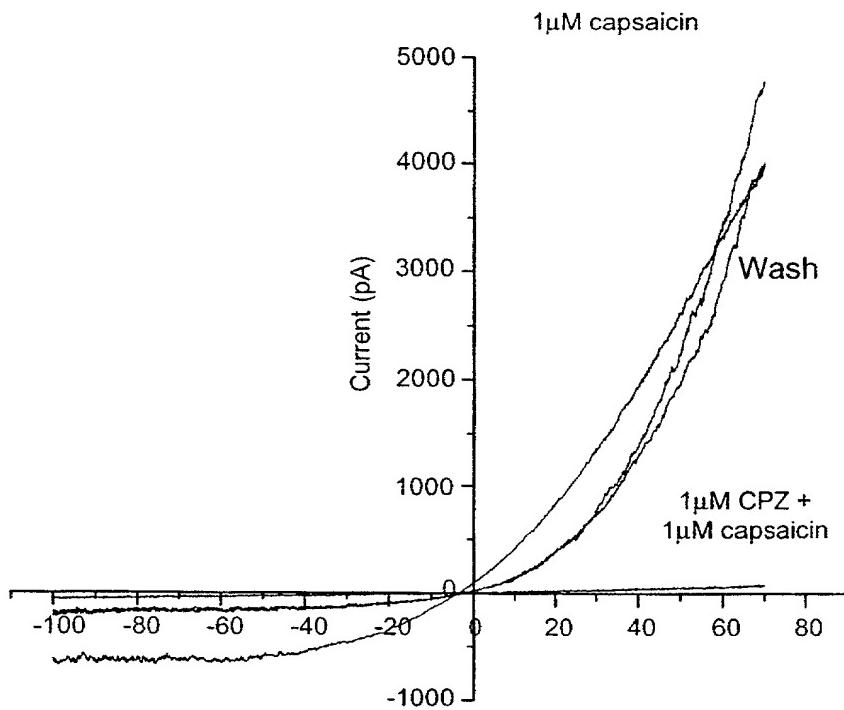


FIG. 11A



SOLUTIONS
OUTSIDE 140mM Na⁺ 2mM Ca²⁺
INSIDE 140mM Cs⁺

FIG. 11B

FIG. 12A

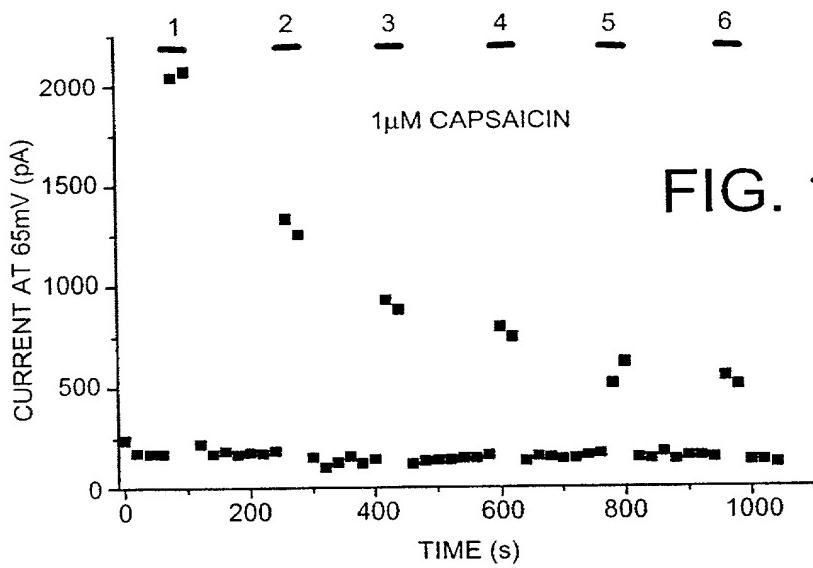
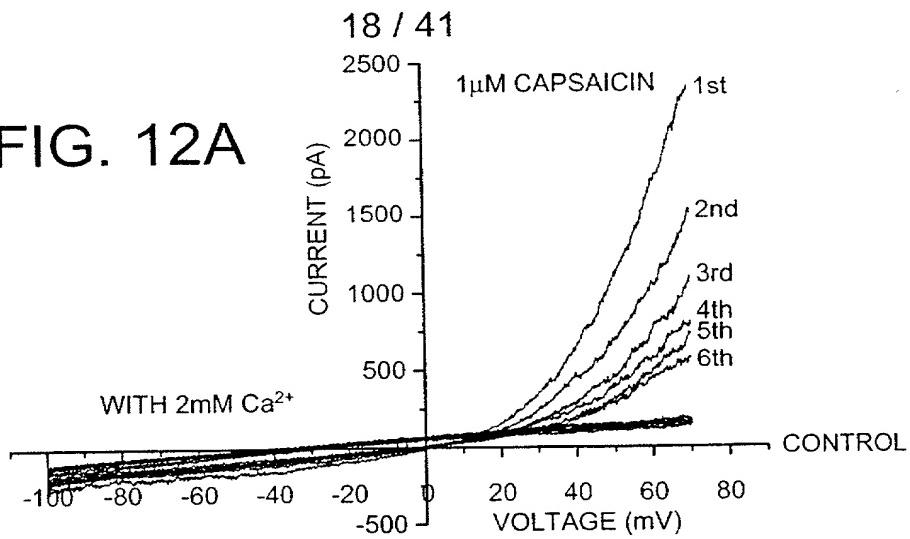
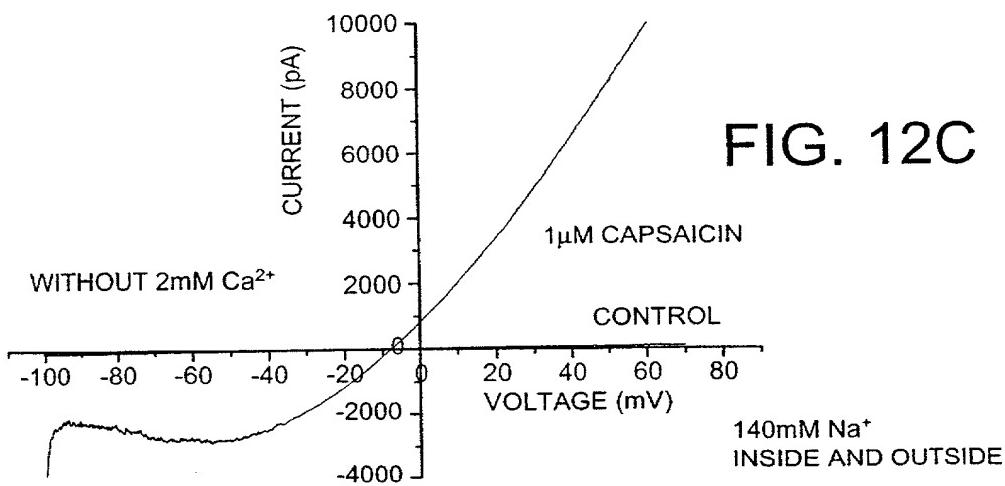
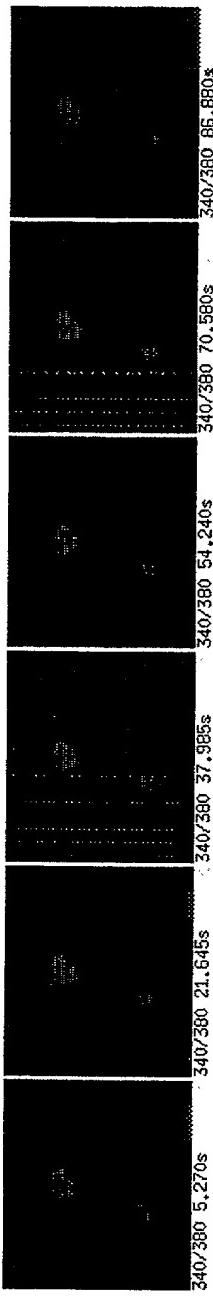


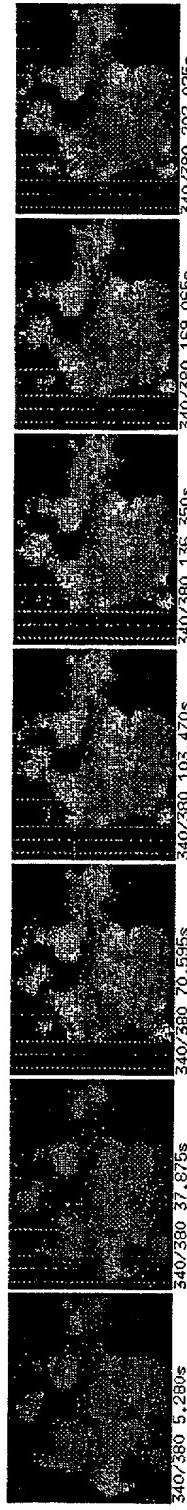
FIG. 12B



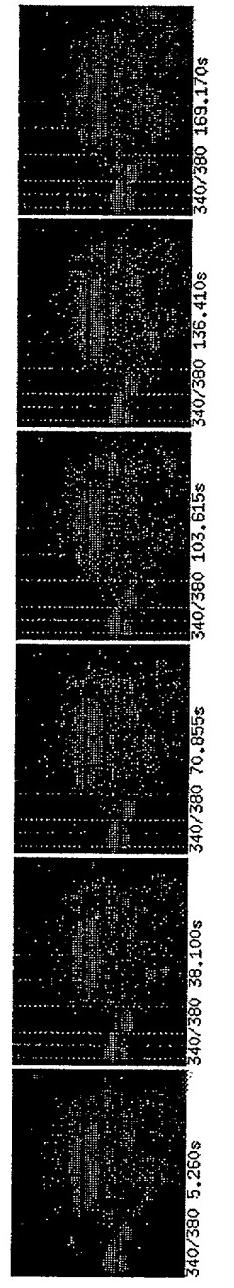
13A pCIN5-new in HEK293T, 24hr transient expression, stimulated with 3 μ M capsaicin at time point 52 secs of time course



13B hVR1pCIN5 in HEK293T, 24hr expression, stimulated with 1 μ M capsaicin at time point 52 seconds



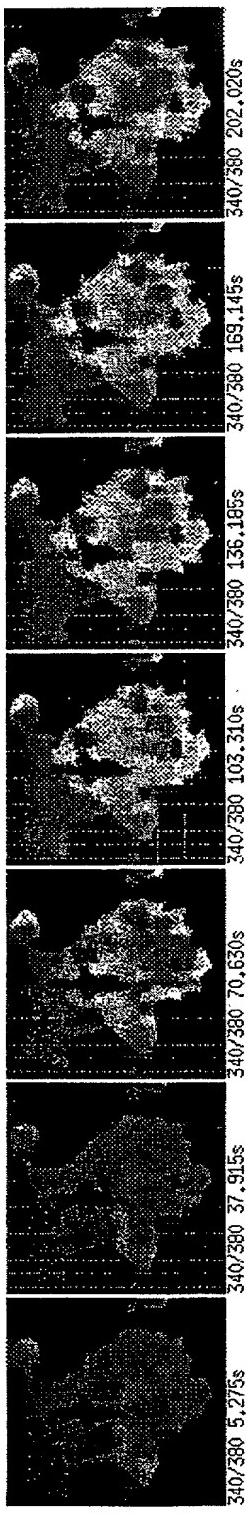
13C hVR1pCIN5 in HEK293T, 24hr transient expression, 20 min pre-incubation with 10 μ M capsazepine, stimulated with 1 μ M capsaicin at time point 52 seconds of time course



[nM Ca²⁺]

FIG. 13

13D hVR1pCIN5 in HEK293T, 24hr transient expression, stimulated with 10 μ M anandamide at time point 52 seconds



13E hVR1pCIN5 in HEK293T, 24hr transient expression, 20 min pre-incubation in 10 μ M capsazepine, stimulated with 10 μ M anandamide at time point 52 sec

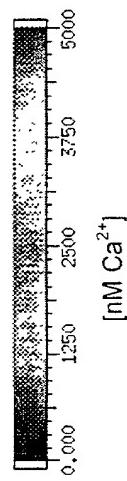
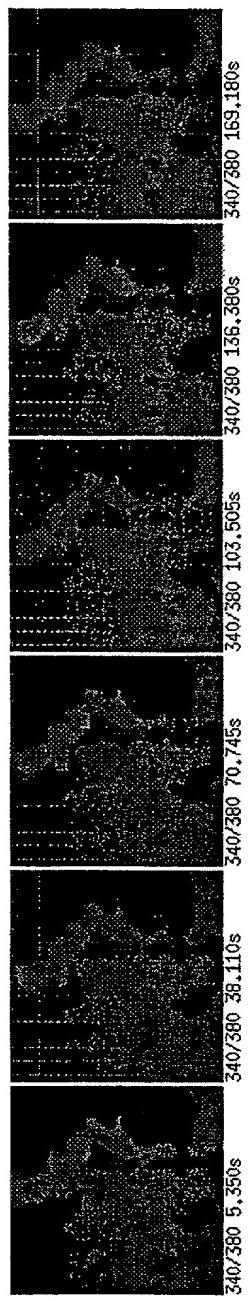
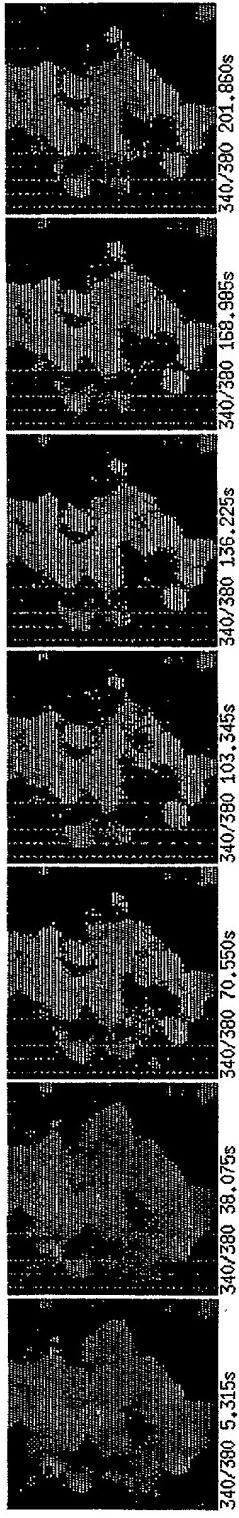


FIG. 13^{CONT'D}

13F hVR1pCIN5 in HEK293T cells, 24hr transient expression, stimulated with 1uM Resiniferatoxin at time point 52 seconds



13G hVR1pCIN5 in HEK293T, 24hr transient expression, 20 min pre-incubation with 10 uM capsazepine, stimulated with 1uM Resiniferatoxin at time point 52 seconds

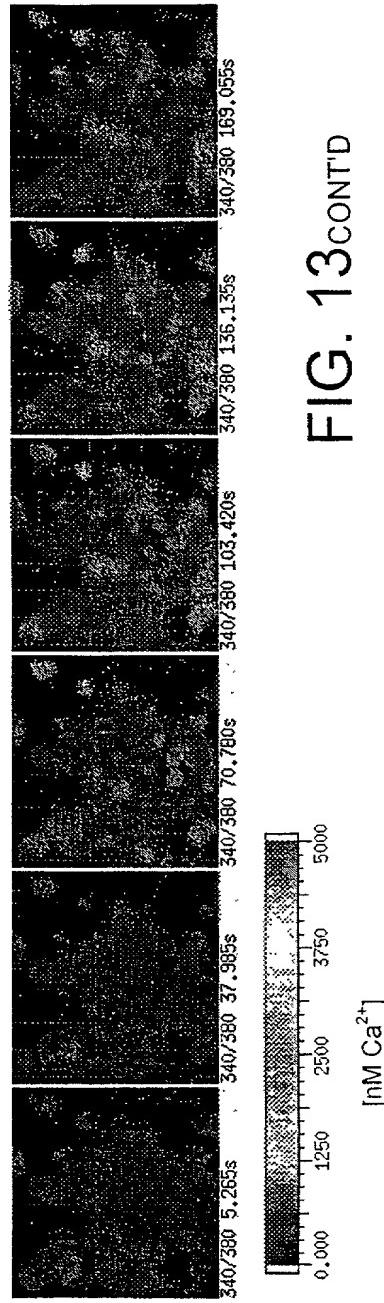


FIG. 13_{CONT'D}

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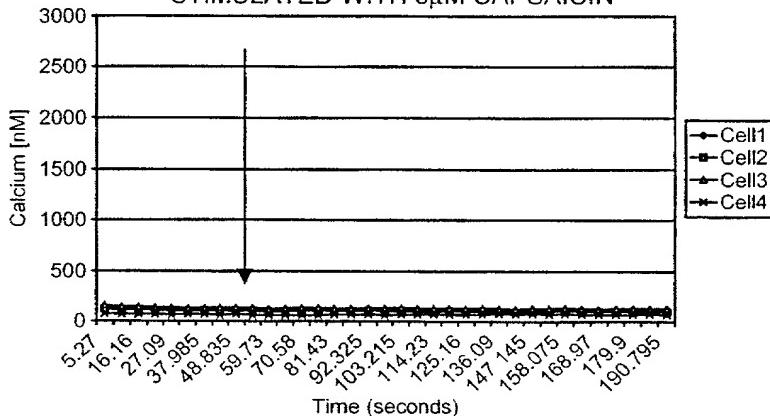
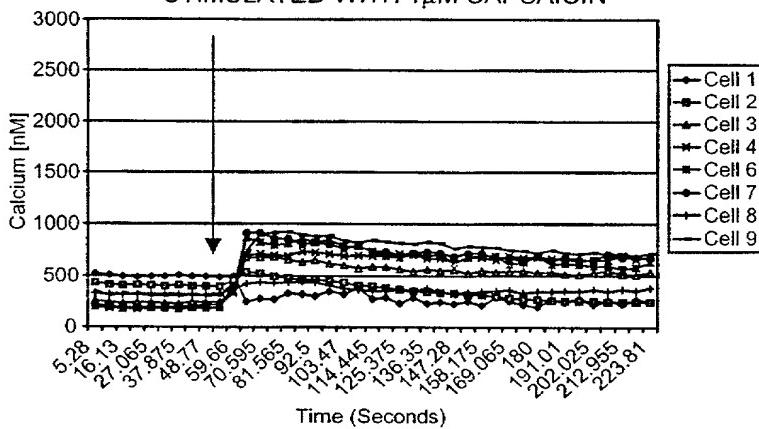
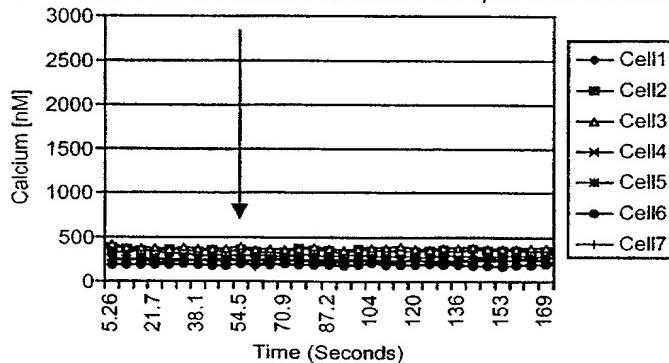
FIG. 14**EXPOSURE OF TRANSFECTED CELLS TO AGONISTS
(ADDITION INDICATED BY ARROW).****14A: pCIN5-NEW IN HEK293T, 24hr TRANSIENT EXPRESSION,
STIMULATED WITH 3 μ M CAPSAICIN****14B: hVR1pCIN5 IN HEK293T, 24hr EXPRESSION,
STIMULATED WITH 1 μ M CAPSAICIN****14C: hVR1pCIN5 IN HEK293T, 24hr TRANSIENT
EXPRESSION, 20 MIN PRE-INCUBATION WITH 10 μ M
CAPSAZEPINE, STIMULATION WITH 1 μ M CAPSIACIN**

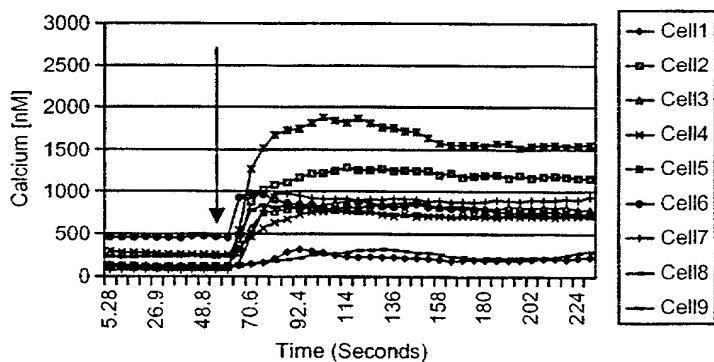
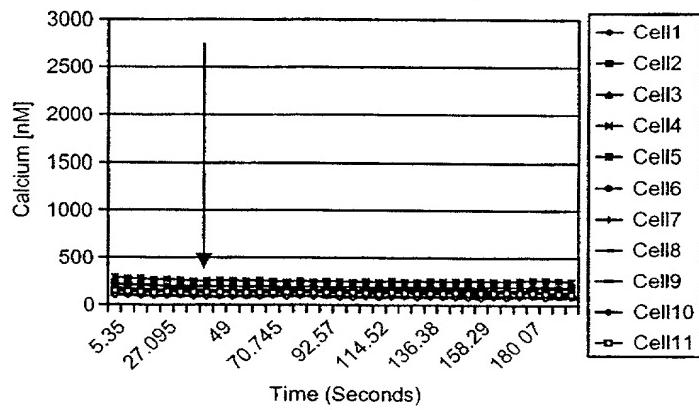
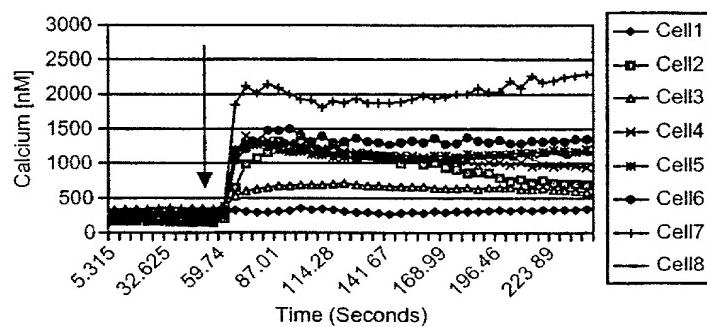
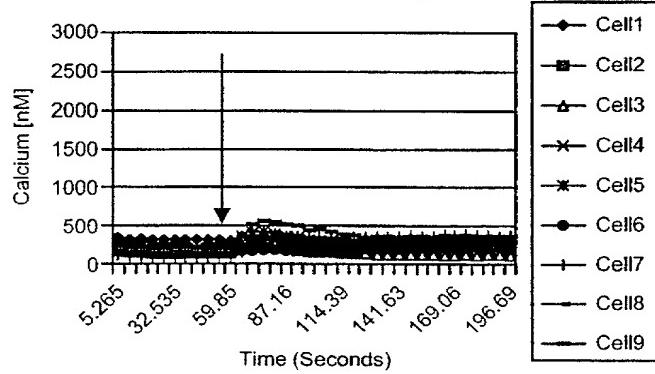
FIG. 14 CONT'D14D: hVR1pCIN5 IN HEK293T, 24hR TRANSIENT EXPRESSION, STIMULATION WITH 10 μ M ANANDAMIDE14E: hVR1pCIN5 IN HEK293T, 24hr TRANSIENT EXPRESSION, 20 MIN PRE-INCUBATION IN 10 μ M CAPAZEPINE, STIMULATED WITH 10 μ M ANANDAMIDE

FIG. 14_{CONT'D}

14F: hVR1pCIN5 IN HEK293T CELLS, 24hr TRANSIENT EXPRESSION, STIMULATED WITH 1 μ M RESINIFERATOXIN



14G: hVR1pCIN5 IN HEK293T, 24hr TRANSIENT EXPRESSION, 20 MIN PRE-INCUBATION WITH 10 μ M CAPSAZEPINE, STIMULATED WITH 1 μ M RESINIFERATOXIN



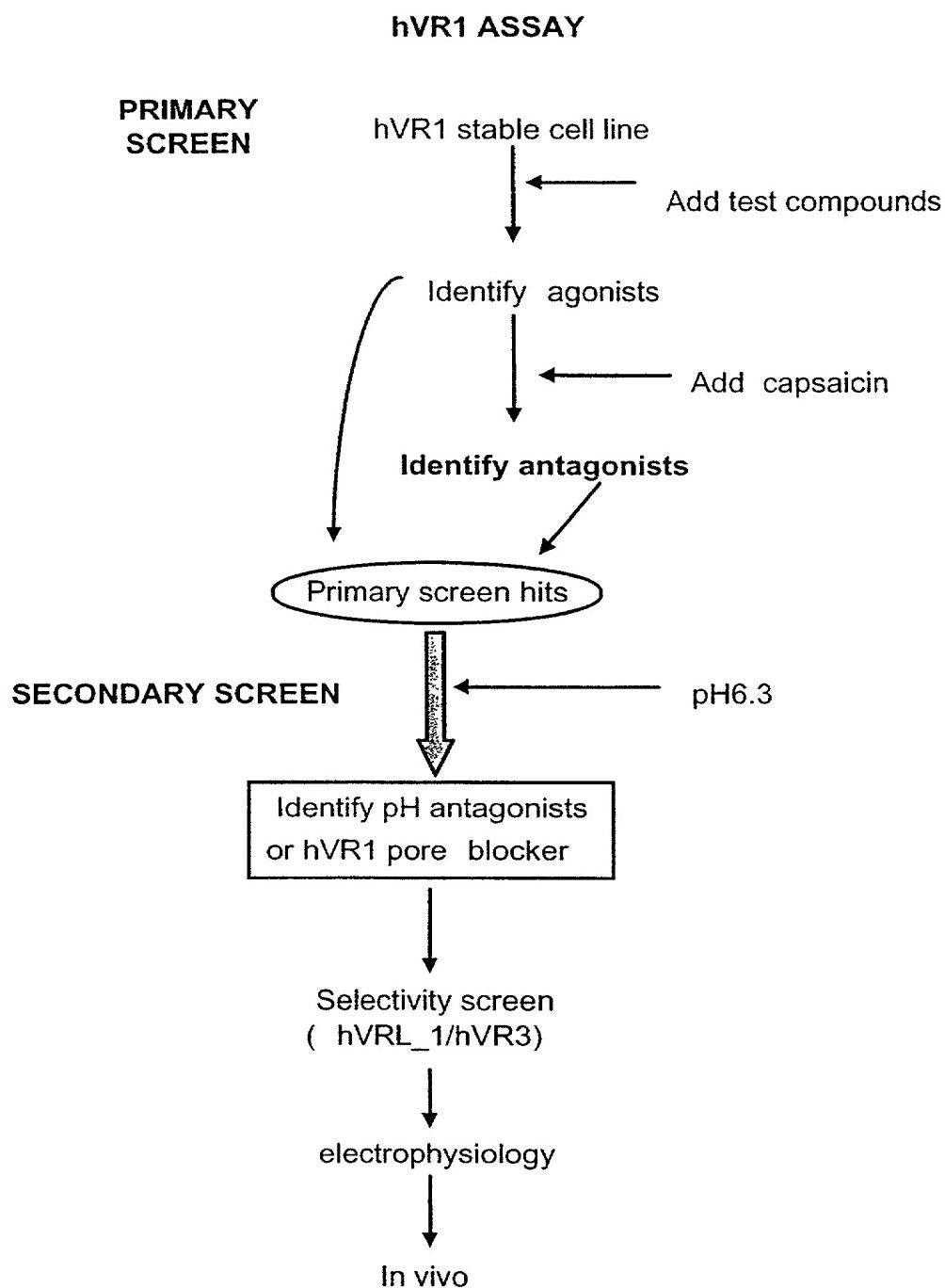
**FIG. 15**

FIG. 16
ALIGNMENT OF THE HUMAN VR3 IN S/L/CO CLUSTERS WITH RAT VR1

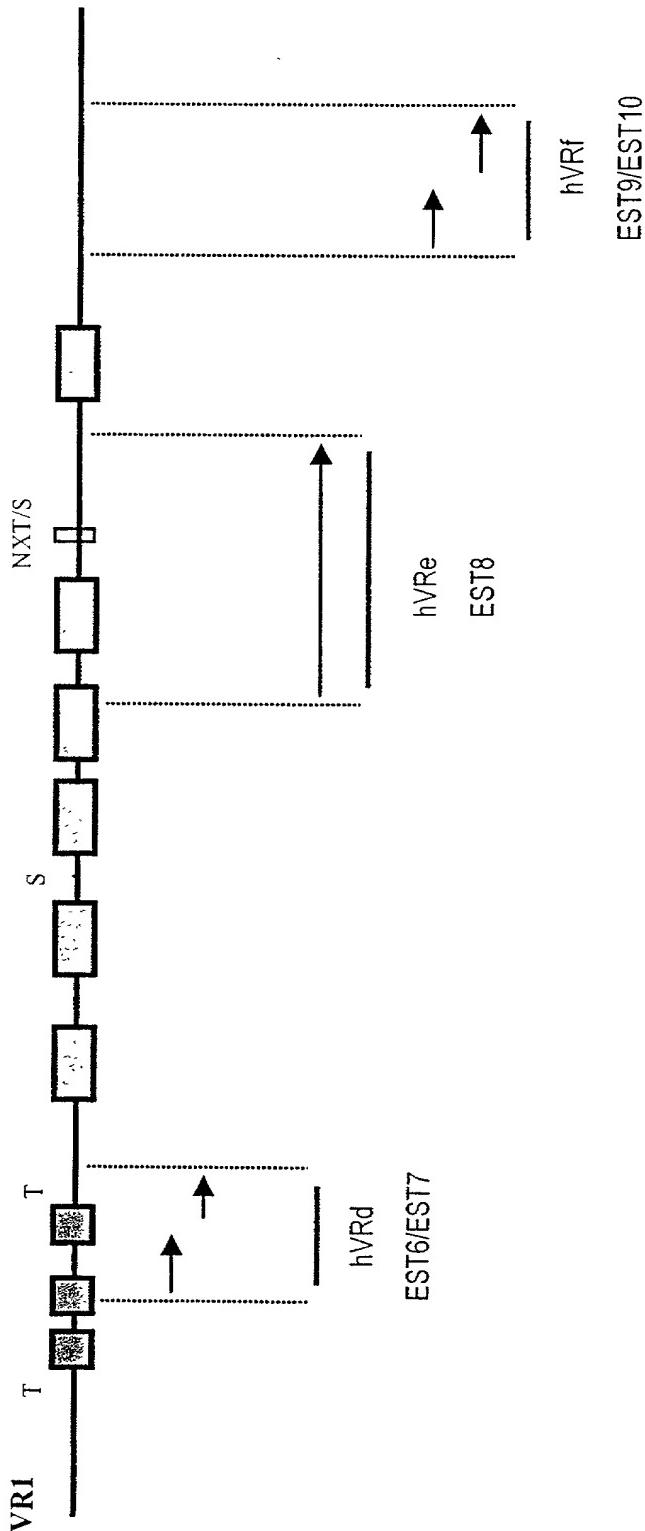


FIG. 17

**hVR3 SEQUENCE INCLUDING 5' UTR (nt -686 TO nt 0) CODING
REGION (nt1 TO nt 2889), 3'UTR (nt 2890 TO nt 3418)**

-684	ttacgcgttaagaaaatacccaagcttatgcatcaagcttggtagccgagctcgatccact	-625
-624	agtaccgcggccagtgtgttggaaattcaagggtgaggagaggagcatggatcctggagc	-565
-564	gagtgtgtcaggccaggggagggctttccagaggagcccagttgagctggAACACCAAGTG	-505
-504	gggaggagttgaccagcaaagggtgcaggagggatcagcactttgcactgggagcagag	-445
-444	tttgtgcactgggaagtcaactcaagtattggagccctcagtttctgttctgtaaaaatg	-385
-384	ggttcatcatgacagtgtttgtgaggaaaaggactgccggctacacagcaagtccaca	-325
-324	tggattttctgagccccctctgtgcctgaagccacggtaatggttctgccttagcagg	-265
-264	tgcttaccacgtgccaggcactgcactgcactggcactggactgcattgtccatg	-205
-204	aggcttggatatccccatcttacagatcaggaagctgaggctatgaaatgtcgacttgct	-145
-144	caatgtcatgaaatgactaagtgtggagccctggatttgcacttggctctctggggctcca	-85
-84	aagctggcttcttggtcagcagtagggctggatccaagttatgggtccagcttgac	-25
-24	cctgaagtccaccctttcagctaATGCCAGGGTAGTTGGACCTGGGCCAATTGTG	35
36	TTTCCAGGTTCGTGAAGAGAGGCTCCTGTCAGTTCCGCCTGAGGCTGGCGGCCAACCA	95
96	CATCTGGGAGTGGCCTCCCTGTGCCCTGTCAATTACAACGGTGGCTTGAAAGCAGCTGGC	155
156	AGCACTGCTGCTTGTCCACGTGGAGGGGGCTTCCTGGAGCCCCCGCCCTGGCCGGGTT	215
216	CTGCCTGACTCCCCTTCATCCCTTGAGGCTGAGCAGTGCAGACGGGCTGGGCCAGG	275
276	CATGGCGGATTCCAGCGAAGGCCCGCGCGGGGCCGGGAGGTGGCTGAGCTCCCCGG	335
336	GGATGAGAGTGGCACCCAGGTGGGAGGCTTTCTCTCCTGGCCAATCTGTT	395

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396	TGAGGGGGAGGATGGCTCCCTTCGCCCTCACCGGCTGATGCCAGTCGCCCTGCTGGCCC	455
456	AGGCAGATGGCGACCAAATCTGCGCATGAAGTTCCAGGGGCCCTCCGCAAGGGGTGCC	515
516	CAACCCCATCGATCTGCTGGAGTCCACCCCTATATGAGTCCTCGGTGGTGCCTGGGCCAA	575
576	GAAAGCACCACATGGACTCACTGTTGACTACGGCACCTATCGTCACCACCTCCAGTGACAA	635
636	CAAGAGGTGGAGGAAGAAGATCATAGAGAACGAGCCAGAGCCCCAAAGCCCCCTGCC	695
696	TCAGCCGCCCCCATCCTCAAAGTCTTCAACCGGCCATCCTCTTGACATCGTGTCCC	755
756	GGGCTCCACTGCTGACCTGGACGGCTGCTCCCATTCTTGCTGACCCACAAGAAACGCCT	815
816	AACTGATGAGGAGTTCGAGAGCCATCTACGGGAAGACCTGCCCTGCCCAAGGCCTTGCT	875
876	GAACCTGAGCAATGCCGCAACGACACCATCCCTGTGCTGCTGGACATCGCGGAGCGCAC	935
936	CGGCAACATGCCGGAGTTCATTAACTCGCCCTCCGTGACATCTACTATCGAGGTAGAC	995
996	AGCCCTGCACATGCCATTGAGCGTCGCTGCAAACACTACGTGGAACCTCTCGTGGCCA	1055
1056	GGGAGCTGATGTCCACGCCAGGGCGTGGCGCTTCTCCAGCCCCAAGGATGAGGGGG	1115
1116	CTACTTCTACTTGGGGAGCTGCCCTGTCGCTGGCTGCCATGACCAACCAGCCCCACAT	1175
1176	TGTCAACTACCTGACGGAGAACCCCCACAAGAACGGGACATGCCGCCAGGACTCGCG	1235
1236	AGGCAACACAGTGCATGCCCTGGTGGCCATTGCTGACAAACACCGTGAGAACACCAA	1295
1296	GTGGTACCAAGATGTACGACCTGCTGCTCAAGTGTGCCCGCTCTCCCCGACAG	1355
1356	CAACCTGGAGGCCGTGCTAACAAACGACGGCCTCTGCCCTCATGATGGCTGCCAAGAC	1415
1416	GGGCAAGATTGGATCTTCAGCACATCCGGGGAGGTGACGGATGAGGACACACG	1475
1476	GCACCTGTCCCGCAAGTCCAAGGACTGGGCCTATGGCCAGTGTATTCCCTCGCTTATGA	1535

FIG. 17 CONT'D

1536	CCTCTCCCTGGACACGTGCGGAAGAGGCCTCCGTGCTGGAGATCCTGGTGTACAA	1595
1596	CAGCAAGATTGAGAACGCCACGAGATGCTGGCTGTGGAGCCCATCAATGAACTGCTGCG	1655
1656	GGACAAGTGGCGGAAGTCGGGCCGTCTCCTTCTACATCAACGTGGTCTCCTACCTGTG	1715
1716	TGCCATGGTTATCTTCACTCTCACCGCCTACTACCAGCCGCTGGAGGGCACACCGCCGTA	1775
1776	CCCTTACCGCACCAACGGTGGACTACCTGCGGCTGGCTGGCGAGGTCAATTACGCTCTTAC	1835
1836	TGGGGTCCTGTTCTTCACCAACATCAAAGACTTGTTCATGAAGAAATGCCCTGGAGT	1895
1896	GAATTCTCTCTTCATTGATGGCTCCTCCAGCTGCTACTTCATCTACTCTGTCCCTGGT	1955
1956	GATCGTCTCAGCAGCCCTTACCTGGCAGGGATCGAGGCCTACCTGGCCATGATGGTCTT	2015
2016	TGCCCTGGCCTGGCTGGATGAATGCCCTTACTTCACCCGTGGCTGAAGCTGACGGG	2075
2076	GACCTATAGCATCATGATCCAGAAGATTCTTCAAGGACCTTCCGATTCTGCTCGT	2135
2136	CTACTTGCTCTTCATGATGGCTACGCTTCAGCCCTGGTCTCCCTCTGAACCGTGTGC	2195
2196	CAACATGAAGGTGTCAATGAGGACCAGACCAACTGCACAGTGCCACTTACCCCTCGT	2255
2256	CCGTGACAGCGAGACCTTCAGCACCTCCCTGGACCTGTTAAGCTGACCATGGCAT	2315
2316	GGGCGACCTGGAGATGCTGAGCAGCACCAAGTACCCGTGGTCTTCATCATCCTGCTGGT	2375
2376	GACCTACATCATCCTCACCTCTGTGCTGCTCCTCAACATGCTCATTGCCCTCATGGCGA	2435
2436	GACAGTGGCCAGGTCTCCAAGGAGAGCAAGCACATGGAAGCTGCAGTGGCCACCAC	2495
2496	CATCCTGGACATTGAGCGCTCCTCCCCGTATTCTGAGGAAGGCCTCCGCTCTGGGA	2555
2556	GATGGTCACCGTGGCAAGAGCTCGGACGGCACTCCTGACCGCAGGTGGTCTCAGGGT	2615
2616	GGATGAGGTGAACGGTCTCACTGGAACCAGAACTGGCATCATCAACGAGGACCCGGG	2675

FIG. 17 CONT'D

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2676	CAAGAATGAGACCTACCAGTATTATGGCTTCTCGCATACCGTGGGCCCTCCGAGGGA	2735
2736	TCGCTGGTCCTCGGTGGTACCCCGCGTGGTGGAACTGAACAAGAACTCGAACCCGGACGA	2795
2796	GGTGGTGGTGCCTCTGGACAGCATGGGGAAACCCCCGCTGCGATGCCACCAGCAGGGTTA	2855
2856	CCCCCGCAAGTGGAGGACTGATGACGCCCCGCTCtagggactgcagcccagccccagctt	2915
2916	cctcgcccaactcatttctagttccagccgcatttcagcagtgccttctgggtgtcccccc	2975
2976	acaccctgcttggccccagaggcgagggaccagtggaggtgccaggaggccccaggac	3035
3036	cctgtggtccctggctctgcctccccaccctgggtggggctccggcacctgtctt	3095
3096	gctcctatggagtcacataagccaacgcccagagccctccacctcaggccccagccctg	3155
3156	cctctccattatttatttgctctgcgtcaggaagcgacgtgacccctgccccagcttgg	3215
3216	acctggcagaggccttaggacccttcaagtgcactgcccggccaagccccagccctca	3275
3276	gacctgcgcctgagctgcatgcgcaccatttggcagcgtggcagtttgcaggggt	3335
3336	ggggccctcggcgtggggccatgccttctgtgttagtgcgtggatggccgg	3395
3396	gctcaataatgtttattcatgaaaaaaaaaaaaaa	3433

FIG. 17 CONT'D

FIG. 18

NUCLEOTIDE AND AMINO ACID SEQUENCE OF hVR3
INCLUDING THE 5'UTR (nt -684 TO nt 0), CODING REGION (nt1
TO 2889) AND 3'UTR (nt 2890 TO nt 3418)

-684	ttacgcgttaagaaaatacccaagcttatgcatacgcttggtagccgactcgatccact	-625
-624	agtaccgcggccaggctgtgctgaaattcaaggtagggagggcatggatcctggagc	-565
-564	gagtgtgtcaggccaggagggcttccagaggagcccagttgagcttggaaacaccagt	-505
-504	gggaggagttgaccagcaaaggtagcaggagggatcagcacattgcacttgcactgggagcagag	-445
-444	tttgtcactgggaagtcaactcaagtattggagccctcagttcctgttctgtaaaatg	-385
-384	ggttcatcatgacagtgttgatgagaaaaaggactgccgactacacagoaagtccaca	-325
-324	tggattttctgagcccttcgtgcctgaagcccacggtaatgggtctgccttagcagg	-265
-264	tgcttaccacgtgccaggcactgcacttgccactggactgcattgttctgtccatg	-205
-204	aggcttggatatccccatcttacagatcaggaagctgaggctatgaaatgtcgacttgct	-145
-144	caatgtcatggaatgactaagtgtggagcctggatttgaacttggctctggggctcca	-85
-84	aagctggcttcttggtcagcagtagggcttggatccaagtatgggtccacagcttgcac	-25
-24	cctgaagtccaccctcttcagctaATGCCAGGGTAGTTGGACCTGGGGCCAATTGTG	35
1	M P R V V G P G A N L C	12
36	TTTCCAGGTTCGTAAAGAGGGCTCCTGTTGCAGTTCCCGCCTGAGGCTGGCGGCCAACCA	95
13	F Q V R E R G S C C S S R L R L A A N H	32
96	CATCTGGGACTGGCCTCCCTGTGCCCCCTGTCAATTACAACGGTGGCTTGAAGCAGCTGGC	155
33	I W E W P P C A P V I T T V A L K Q L A	52
156	AGCACTGCTGCTTGTCACGTGGAGGGGCTTCTGGAGCCCCCGCCCCCTGGCCGGGTT	215
53	A L L L V H V G G G F L E P P P L A G F	72
216	CTGCCTGACTCCCCTTCATTCCCTTGAGGCTGAGCAGTGCAGACGGGCCTGGGCAGG	275
73	C L T P L S F P C R L S S A D G P G A G	92
276	CATGGCGGATTCCAGCGAAGGCCCGCGCGGGGGCCGGGAGGTGGCTGAGCTCCCCGG	335
93	M A D S S E G P R A G P G E V A E L P G	112
336	GGATGAGAGTGGCACCCAGGTGGGAGGCTTCTCTCTCCCTGGCCAATCTGTT	395
113	D E S G T P G G E A F P L S S L A N L F	132
396	TGAGGGGGAGGATGGCTCCCTTCGCCCTCACCGGCTGATGCCAGTCGCCCTGCTGGCC	455
133	E G E D G S L S P S P A D A S R P A G P	152
456	AGGCGATGGCGACCAAATCTGCGCATGAAGTCCAGGGCGCCTCCGCAAGGGGTGCC	515
153	G D G R P N L R M K F Q G A F R K G V P	172
516	CAACCCCCATCGATCTGCTGGAGTCCACCCATATGAGTCCTCGGTGGTGCCTGGGCCAA	575
173	N P I D L L E S T L Y E S S V V P G P K	192

576	GAAAGCACCCATGGACTCACTGTTGACTACGGCACCTATCGTCACCACCTCCAGTGACAA	635
193	K A P M D S L F D Y G T Y R H H S S D N	212
636	CAAGAGGTGGAGGAAGAAGATCATAGAGAACGAGCCGAGAGCCCCAAAGCCCCCTGCC	695
213	K R W R K K I I E K Q P Q S P K A P A P	232
696	TCAGCCGCCCATCCTCAAAGTCTTCAACCGGCCTATCCTCTTGACATCGTGTCCC	755
233	Q P P P I L K V F N R P I L F D I V S R	252
756	GGGCTCCACTGCTGACCTGGACGGGCTGCTCCATTCTGCTGACCCACAAGAAACGCC	815
253	G S T A D L D G L L P F L L T H K K R L	272
816	AACTGATGAGGAGTTCGAGAGCCATCTACGGGAAGACCTGCCTGCCAAGGCCTGCT	875
273	T D E E F R E P S T G K T C L P K A L L	292
876	GAACCTGAGCAATGGCCGCAACGACACCATCCCTGTGCTGCTGGACATCGCGAGCGCAC	935
293	N L S N G R N D T I P V L L D I A E R T	312
936	CGGCAACATGCGGGAGTTCACTAACCGCCCTCCGTGACATCTACTATCGAGGTAGAC	995
313	G N M R E F I N S P F R D I Y Y R G Q T	332
996	AGCCCTGCACATGCCATTGAGCGCTGCAAACACTACGTGGAACTTCTCGTGGCCA	1055
333	A L H I A I E R R C K H Y V E L L V A Q	352
1056	GGGAGCTGATGTCACGCCAGGCCGTGGCGCTTCTCCAGGCCAAGGATGAGGGGG	1115
353	G A D V H A Q A R G R F F Q P K D E G G	372
1116	CTACTTCTACTTGGGAGCTGCCCTGTCGCTGGCTGCCATGCCAACAGCCCCACAT	1175
373	Y F Y F G E L P L S L A A C T N Q P H I	392
1176	TGTCAACTACCTGACGGAGAACCCCCACAAGAAGGCGGACATGCGGCCAGGACTCGCG	1235
393	V N Y L T E N P H K K A D M R R Q D S R	412
1236	AGGCAACACAGTGCATGCCATGCCGTGGCATTGCTGACAACACCCGTGAGAACACCAA	1295
413	G N T V L H A L V A I A D N T R E N T K	432
1296	GTTGTTACCAAGATGTACGACCTGCTGCTCAAGTGTGCCCTCTTCCCCGACAG	1355
433	F V T K M Y D L L L K C A R L F P D S	452
1356	CAACCTGGAGGCCGTGCTCAACAACGACGCCCTCGCCCTCATGATGGCTGCCAAGAC	1415
453	N L E A V L N N D G L S P L M M A A K T	472
1416	GGCAAGATTGGATTTCAAGCACATCATCCGGGGAGGTGACGGATGAGGACACACG	1475
473	G K I G I F Q H I I R R E V T D E D T R	492
1476	GCACCTGTCCCGCAAGTCCAAGGACTGGGCCTATGGGCOAGTGTATTCTCGCTTATGA	1535
493	H L S R K S K D W A Y G P V Y S S L Y D	512
1536	CCTCTCCTCCCTGGACACGTGTGGGAAGAGGCCCTCGCTGGAGATCCTGGTGTACAA	1595
513	L S S L D T C G E E A S V L E I L V Y N	532
1596	CAGCAAGATTGAGAACGCCACGAGATGCTGGCTGTGGAGCCCATCAATGAACGTGCTGCG	1655
533	S K I E N R H E M L A V E P I N E L L R	552
1656	GGACAAAGTGGCGGAAGTCGGGCCGTCTCCTTCTACATCAACGTGGTCTCCTACCTGTG	1715
553	D K W R K F G A V S F Y I N V V S Y L C	572

FIG. 18 CONTD

1716	TGCCATGGTTATCTTCACTCTCACCGCCTACTACCAGCCGCTGGAGGGCACACCGCCGTA	1775
573	A M V I F T L T A Y Y Q P L E G T P P Y	592
1776	CCCTTACCGCACCACGGTGGACTACCTGC GGCTGGCTGGCGAGGT CATTACGCTCTTCAC	1835
593	P Y R T T V D Y L R L A G E V I T L F T	612
1836	TGGGGT CCTGTTCTTCACCAACATCAAAGACTTGTTCATGAAGAAATGCCCTGGAGT	1895
613	G V L F F F T N I K D L F M K K C P G V	632
1896	GAATTCTCTTCATTGATGGCTCCTTCCAGCTGCTCTACTTCATCTACTCTGTCCCTGGT	1955
633	N S L F I D G S F Q L L Y F I Y S V L V	652
1956	GATCGTCTCAGCAGCCCTCTACCTGGCAGGGATCGAGGCCTACCTGGCATGATGGTCTT	2015
653	I V S A A L Y L A G I E A Y L A M M V F	672
2016	TGCCCTGGT CCTGGCTGGATGAATGCCCTTACTTCACCCGTGGCTGAAGCTGACGGG	2075
673	A L V L G W M N A L Y F T R G L K L T G	692
2076	GACCTATAGCATCATGATCCAGAAGATTCTCTTCAGGACCTTCCGATTCTGCTCGT	2135
693	T Y S I M I Q K I L F K D L F R F L L V	712
2136	CTACTTGCTCTTCATGATGGCTACGCTTCAGCCCTGGTCTCCCTCCTGAACCCGTGTGC	2195
713	Y L L F M I G Y A S A L V S L L N P C A	732
2196	CAACATGAAGGTGTGCAATGAGGACCAGACCAACTGCACAGTGCCCACTTACCCCTCGTG	2255
733	N M K V C N E D Q T N C T V P T Y P S C	752
2256	CCGTGACAGCGAGACCTTCAGCACCTCCTGGACCTGTTAAGCTGACCATCGGCAT	2315
753	R D S E T F S T F L L D L F K L T I G M	772
2316	GGCGCACCTGGAGATGCTGAGCAGCACCAAGTACCCCGTGGTCTTCATCATCCTGCTGGT	2375
773	G D L E M L S S T K Y P V V F I I L L V	792
2376	GACCTACATCATCCTCACCTCTGTGCTCCTCAACATGCTCATTGCCCTCATGGCGA	2435
793	T Y I I L T S V L L L N M L I A L M G E	812
2436	GACAGTGGGCCAGGTCTCCAAGGAGAGCAAGCACATCTGGAAGCTGCAGTGGGCCACCAC	2495
813	T V G Q V S K E S K H I W K L Q W A T T	832
2496	CATCCTGGACATTGAGCGCTCCTCCCCGTATTCTGAGGAAGGCCCTCCGCTCTGGGA	2555
833	I L D I E R S F P V F L R K A F R S G E	852
2556	GATGGTCACCGTGGCAAGAGCTCGGACGGCACTCCTGACCGCAGGTGGTCTCAGGGT	2615
853	M V T V G K S S D G T P D R R W C F R V	872
2616	GGATGAGGTGAACGGTCTCACTGGAACAGAACCTGGCATCATCACGAGGACCCGGG	2675
873	D E V N W S H W N Q N L G I I N E D P G	892
2676	CAAGAATGAGACCTACCAAGTATTATGGCTTCTCGCATACCGTGGGCCCTCCGAGGG	2735
893	K N E T Y Q Y Y G F S H T V G R L R R D	912
2736	TCGCTGGT CCTCGGTGGTACCCCGCGTGGAACTGAACAAGAACCTCGAACCCGGACGA	2795
913	R W S S V V P R V V E L N K N S N P D E	932
2796	GGTGGTGGTGCCTCTGGACAGCATGGGAACCCCGCTGCGATGCCACCAGCAGGGTTA	2855
933	V V V P L D S M G N P R C D G H Q Q G Y	952

FIG. 18 CONT'D

2856	CCCCCGCAAGTGGAGGACTGATGACGCCCGCTCtagggactgcagcccagccccagctt	2915
953	P R K W R T D D A P L	963
2916	cctcgcccaactcattttagtccagccgcatttcagcagtgcctctgggtgtccccc	2975
2976	acaccctgcttggcccccagaggcgagggaccagtggaggtgccagggaggccccaggac	3035
3036	cctgtggtccccctggctctgcctccccaccctgggtggggctccggccacctgtctt	3095
3096	gctcctatggagtcacataagccaacgccagagccctccacctcaggccccagccctg	3155
3156	cctctccattatttatttgcgtctcaggaagcgacgtgaccctgccccagctgga	3215
3216	acctggcagaggccttaggaccccgttccaagtgcactgcccggccaagccccagcctca	3275
3276	gcctgcgcctgagctgcatgcgccaccatttggcagcgtggcagcttgcaagggct	3335
3336	ggggccctggcgtgggccatgcctctgtgtttctgttagtgtctggatttccgggt	3395
3396	gctcaataaatgtttattcattgaaaaaaaaaaaaaaa 3433	

FIG. 18_{CONT'D}

FIG. 19

AMINO ACID SEQUENCE OF hVR3

1 MPRVVGPGAN LCFQVRERGS CCGSRLRLAA NHIWEWPPCA PVITTVALKQ
51 LAALLLVHVG GGFLEPPPLA GFCLTPLSFP CRLSSADGPG AGMADSSEGP
101 RAGPGEVAEL PGDESGTPGG EAFLPLSSLAN LFEGEDGSLS PSPADASRPA
151 GPGDGRPMLNR MKFQGAFRKKG VPPNPIDLLES TLYESSVVPG PKKAPMDSL
201 DYGTYRHSS DNKRWRKKII EKQPQSPKAP APQPPPILKV FNRPILFDIV
251 SRGSTADLDG LLPFLLTHKK RLTDEEFREP STGKTCLPKA LLNLNSGRND
301 TIPVLLDIAE RTGNMREFIN SPFRDIYYRG QTALHIAIER RCKHYVELLV
351 AQGADVHAQA RGRFFQPKDE GGYFYFGELP LSLAACTNQP HIVNYLTEM
401 HKKADMRRQD SRGNTVLHAL VAIADNTREN TKFVTKMYDL LLLKCARLFP
451 DSNLEAVLNN DGLSPIMMAA KTGKIGIFQH IIRREVTDED TRHLSRKSKD
501 WAYGPVYSSL YDLSSLDTG EEASVLEILV YNSKIENRHE MLAVEPINEL
551 LRDKWRKFGA VSEYINVVSY LCAMVIEETTLEAYQPLEGTP PYPYRTTVDY
601 LRLACAEVITTEEVGVIEETTNEIK DLFMKKCP GVNSLFI DGS FQIILYEIYSV
651 EVIEVSAALYC AGIEAYLAMM VFAEVLGWMN ALIYETRGLKL TGTYSIMI QK
701 ILFKDIEFRFTEEVYELFMIGY ASALV SLLNP CANMKVCNED QTNCVPTYP
751 SCRDSETFST FLDDLFLKLTG GMGDILEMLSS TKYPVVFIIILIVTYIIETSV
801 IETTNNMTATM CETVGQVSKE SKHIWKLQWA TTILDIERSF PVFLRKAFRS
851 GEMVTVGKSS DGTPDRRWCF RVDEVNWSHW NQNLGIINED PGKNETYQYY
901 GFSHTVGRLR RDRWSSVVPR VVELNKNSNP DEVVVPLOSM GNPRCDGHQQ
951 GYPRKWRTDD APL

Key

Transmembrane domains

Ankyrin binding domains

FIG. 20

FULL-LENGTH hVR3 CLONED INTO (A) pBLUESCRIPT SK(+) (hVR3pBSK) AND (B) pCDNA3.1(+) (hVR1pCDNA3.1) VIA NotI/Xhol RESTRICTION SITES.

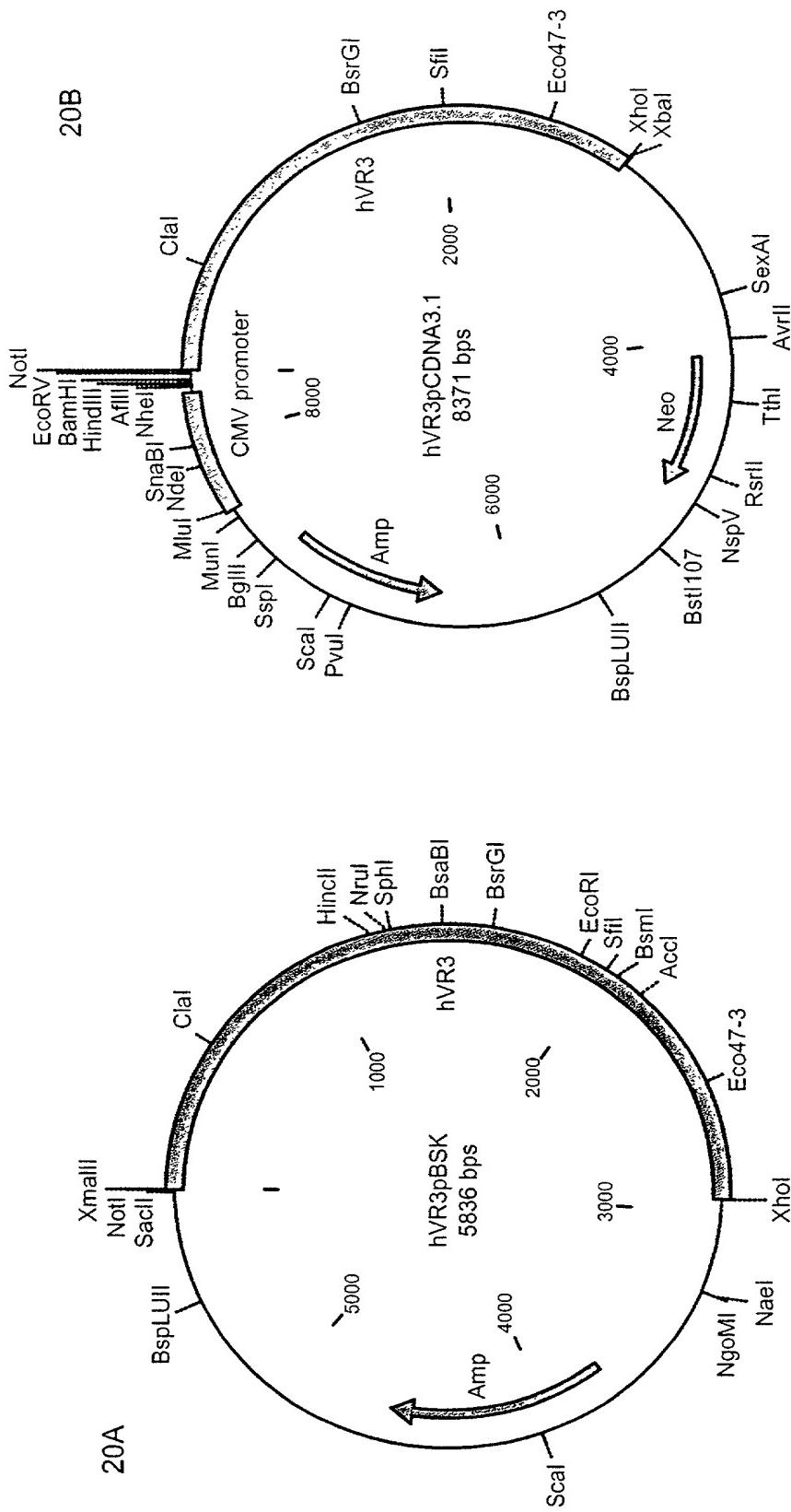


FIG. 21

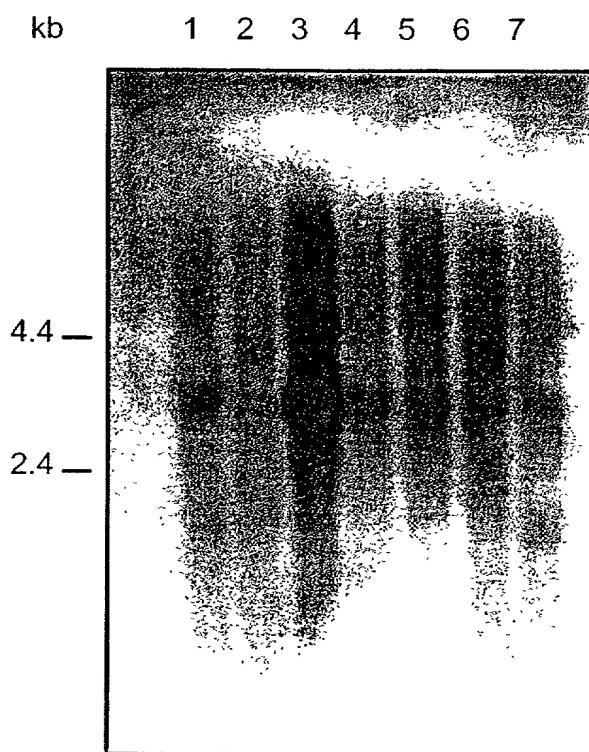
A MULTIPLE COMPARISON OF THE AMINO ACID SEQUENCES OF THE RAT VR1 AND THE HUMAN VANILLOID RECEPTORS, hVR1, hVRL-1 AND hRV3

	10	20	30	40	50
VR1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVR1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVRL-1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVR3	MPRVVGPGANLCFQVRERGSCCSSRLRLAANHIWEWPPCAPVITTVALKQ				
	60	70	80	90	100
VR1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVR1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVRL-1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVR3	LAALLLVHVGFFLEPPPLAGFCLTPLSFPCLRSSADGPGAGMADSSEGP				
	110	120	130	140	150
VR1	~~~~~	~~~~~	MEORASLDSEESESPPQENSCL		
hVR1	~~~~~	~~~~~	MKKWSSTDLGAAADPLQKDTCP		
hVRL-1	~~~~~	~~~~~	~~~~~		
hVR3	RAGPGEVAELPGDESGTPGGEAFPLSSLANLFEGEDGSLSPSPADASRPA				
	160	170	180	190	200
VR1	DPPDRDPNCKPPPVKPHIFTRSRTRLFG	KGDSEEA	SPLDCPYEEG	
hVR1	DPLDGDPNSSRPPPAKQQLSTAKSRTRLFG	KGDSEEA	FPVDCPHEEG	
hVRL-1	~~~~~	MTSPSSSPVFRLETLDGGQEDGSEADRGKLDF			
hVR3	GPGDGRPNLRMKFQGAFRKGVPNP	IDLLESTLYESSVVPGPKKAP		
	210	220	230	240	250
VR1	GLASCPIITVSSVLTIQRPGDGPASVRPSSQDSVSAG	.EKP.	PRLYDRRS		
hVR1	EILDSCPPIITVSPVITIQRPGDGP	TGARLLSQDSVA	STEKT.LRLYDRRS		
hVRL-1	GSGLPPM..ESQFOGEDRKFAPQIRVNLYRKGTGASQPDP	.NR.	FDRDR		
hVR3	MDSLFDYGTYRHSSDNKWRKKIIIEKQ	QSPKAPAPQOPP	PILKVFNRP	I	
	260	270	280	290	300
VR1	IEDAVAOQNCOLESLLPFLQRSKKRLTDSEFKDPE	TGKTCLIKAMLNH			
hVR1	IFEAVAQNQNCODLE	SLLLFLQSKKHLTDNEFKDPE	TGKTCLIKAMLNH		
hVRL-1	LENASRVGPEDLAGIPEYLSKTSKYLT	SEYTEGSTGKTCLIMKAVLNK			
hVR3	LEDIVSRGSTDLDGLLPFLLTHKKRLT	DEFRE	PSTGKTCLPKALLNL	S	
	310	320	330	340	350
VR1	NGQNDTIALLLDVARKTDSLQFVNASYTD	SYYKGQ	OTALHIAIERRNM	TT	
hVR1	DGQNTTIPLLEIARQTDDSLKE	LVNASYTD	SYYKGQ	OTALHIAIERRN	MAL
hVRL-1	DGVNACILPLLIQIDRDGSNPQPLVNA	QCTDDY	YRGHSA	LHIAIEKRSLQC	
hVR3	NGRNDTIPVLLDIAERTGNMREFINS	SPFRL	IYYRGQ	OTALHIAIERRCKHY	
	360	370	380	390	400
VR1	VILLVENGADVOAAANGDFFKTKGRPGFYFGE	PLSLA	ACTNOLAI	IVK	
hVR1	VILLVENGADVOAAAHGDF	FFKTKGRPGFYFGE	PLSLA	ACTNOLGIV	VF
hVRL-1	VKILLVENGANVHARACGRFFQKGQG	.TCFYFGE	PLSLA	ACTKQWDVVSY	
hVR3	VELLEVAQGADVHAQARGRFFQPKDEGGY	FYFGE	PLSLA	ACTNQPHIV	NY
	410	420	430	440	450
VR1	LLQNSWQADISARD	SVGNTVLHALVE	ADNTVDNTKFVT	SMYNE	ILILG
hVR1	LLQNSWQADISARD	SVGNTVLHALVE	ADNTADNTKFVT	SMYNE	ILILG
hVRL-1	LLENPHQ	PASLQATDSQGNTVLHALVM	ISDN	SAENIALV	TSMYDGLLQAG
hVR3	LLENPHKKADMRRQDSRGNTVLHALV	IA	ADNTRENTKFVT	MYD	LLLKC
	460	470	480	490	500
VR1	AKLHPTLKLEEITNRKGLTP	PLALAASS	SKIGV	YLAYT	LOREI
hVR1	AKLHPTLKLEEITNKKGMTPL	ALAAGT	CKIGV	YLAYT	LOREI
hVRL-1	ARICPTVQEDIRNLQDLTP	KLAAKE	GKIE	FRH	ILOREFS..GLSHES
hVR3	ARIFPDNSL	EA	VNLNDGL	SPLMM	AAKTGKIGIFQHIIREVTD

	510	520	530	540	550
VR1	RKFTEWAYGPVHSSLYDLSCIDTC.	EKNBVLEVIAYSSSETPNRHDMILLV			
hVR1	RKFTEWAYGPVHSSLYDLSCIDTC.	EKNBVLEVIAYSSSETPNRHDMILLV			
hVRL-1	RKFTEWCYGPVRVSLYDLASVDSC.	EENBVLEIIAF.HCKSPHRHRMVVL			
hVR3	RKSKDWAYGPVYSSLYDLSSLDTCGEEASVLEILVY.WSKIENRHEMLAV				
	560	570	580	590	600
VR1	EPLNRLLQDKWDRFVKRIFYFMFFVYCLYMIIFTAAAYYRPV..EGLPPY				
hVR1	EPLNRLLQDKWDRFVKRIFYFMFLVYCLYMIIFTMAAYYRPV..DGLPPF				
hVRL-1	EPLNKLLOAKWDLLIPK.FFLNFLCNLIYMFIFTAVAYHQPTLIKQOAPH				
hVR3	EPINELLRDKWRKEGAVSFYINVVSYLCAMVIFTLTAYYQPL..EGTPPY				
	610	620	630	640	650
VR1	KLKNTVGDYFRVTGEILSVGGVYFFFRCGIQ.YFLQRRPSLKSLFVDSYS				
hVR1	KMEN.IGDYFRVTGEILSVLGGVYFFFRCGIQ.YFLQRRPSMKTLFVDSYS				
hVRL-1	.LNAEVGNMSMLLTGHILILLGGIYLGVQLW.YFWRRHVFIWISFIDSYF				
hVR3	PYRTTV.DYLRLLAGEVITLFTGVLFFFTNIKDLFMMKCPGVNSLFIDGSF				
	660	670	680	690	700
VR1	EILFFVQSLFMLVSVVLYFSQRKEYVASMVFSLAMGWTNMLYYTRGFQQM				
hVR1	EMLFFLQSLFMLATVVLYFSHLKEYVASMVFSLALGWTNMLYYTRGFQQM				
hVRL-1	EILFLFQALLTVVSQVLCFLAIEWYLPLLVSVLGLWNLLYYTRGFQHT				
hVR3	QLLYIFIYSLVIVSAALYLAGIEAYLAMMVFAVLVLGWMNALYFTRGLKLT				
	710	720	730	740	750
VR1	GIYAVMIEKMLRDLCRFMFVYLVFLFGFSTAVVTLIEDGKNNSLP....				
hVR1	GIYAVMIEKMLRDLCRFMFVYIVFLFGFSTAVVTLIEDGKNDSLP....				
hVRL-1	GIYSVMIQKVILRDLLRFLLIYLVFLFGFAVALVSLSQEAWRPEAPTGPN				
hVR3	GTYSIMIQKILFKDLFRFLVYLLFIMIGYASALVSLINPCANMKVCNEDQ				
	760	770	780	790	800
VR1	MESTPHKCGRSACK.PGNSYNSLYSTCLELFKFTIGMDLEFTENYDFKA				
hVR1	SESTSHWRGPACRPPDSSYNLSYSTCLELFKFTIGMDLEFTENYDFKA				
hVRL-1	ATESVQPMEGQEDEGNGAQYRGILEASLELFKFTIGMDLEFTENYDFKA				
hVR3	TNCTVPTY..PSCR.DSETFSTFL...LDLFKLTIGMDLEFTENYDFKA				
	810	820	830	840	850
VR1	VFIILLLAYVILTYIILLNMLIALMGETVNKIAQESKNIKLORAITILD				
hVR1	VFIILLLAYVILTYIILLNMLIALMGETVNKIAQESKNIKLORAITILD				
hVRL-1	MVILLILAYVILTYIILLNMLIALMSETVNSVATDSWSIWKLOKAISVLE				
hVR3	VETILLVITYIILTSVILLNMLIALMGETVGQVSKEKHIWKLQWATTILD				
	860	870	880	890	900
VR1	TEKSFLKCMRKAFRSRGKLLQVGETPDGKDGYRWCFRDEVNWTTWNTNVG				
hVR1	TEKSFLKCMRKAFRSRGKLLQVGETPDGKDGYRWCFRDEVNWTTWNTNVG				
hVRL-1	MENGYWWC.RKKQAGVMLTVGTPDGSPDERWCFRVEVNWASWEQTLPM				
hVR3	IERSFPVFLRKAFRSGEMVTVGKSSDGTDPRRWCFRDEVNWASHWNQNLG				
	910	920	930	940	950
VR1	IINEDPGNCE.....GVKRTLFSLSLRSG.....RVSGRNWKNEALV				
hVR1	IINEDPGNCE.....GVKRTLFSLSLRSS.....RVSGRNWKNEALV				
hVRL-1	TLCEDPSGA.....GVPRTLENPVLAS.....PPKEDEDGASEENYVPV				
hVR3	IINEDPGKWEETYQYYGFSHTVGRLLRRDRWSSVPRVVELNKNSNPDEVVV				
	960	970	980	990	
VR1	PLLRASTRDRHATQOEVOLKHYTGSILKPEDAEVFKDSMVPGEN				
hVR1	PLLREASARDRQSAQPEEVYLRFQSGSLKPEDAEVFKSPAASGEN				
hVRL-1	QLLQSN~~~~~				
hVR3	PLDSMGNPRCDGHQQGYPRKWRTDDAPL~~~~~				

FIG. 21 CONT'D

FIG. 22A
HYBRIDISATION OF A NORTHERN BLOT WITH hVR3

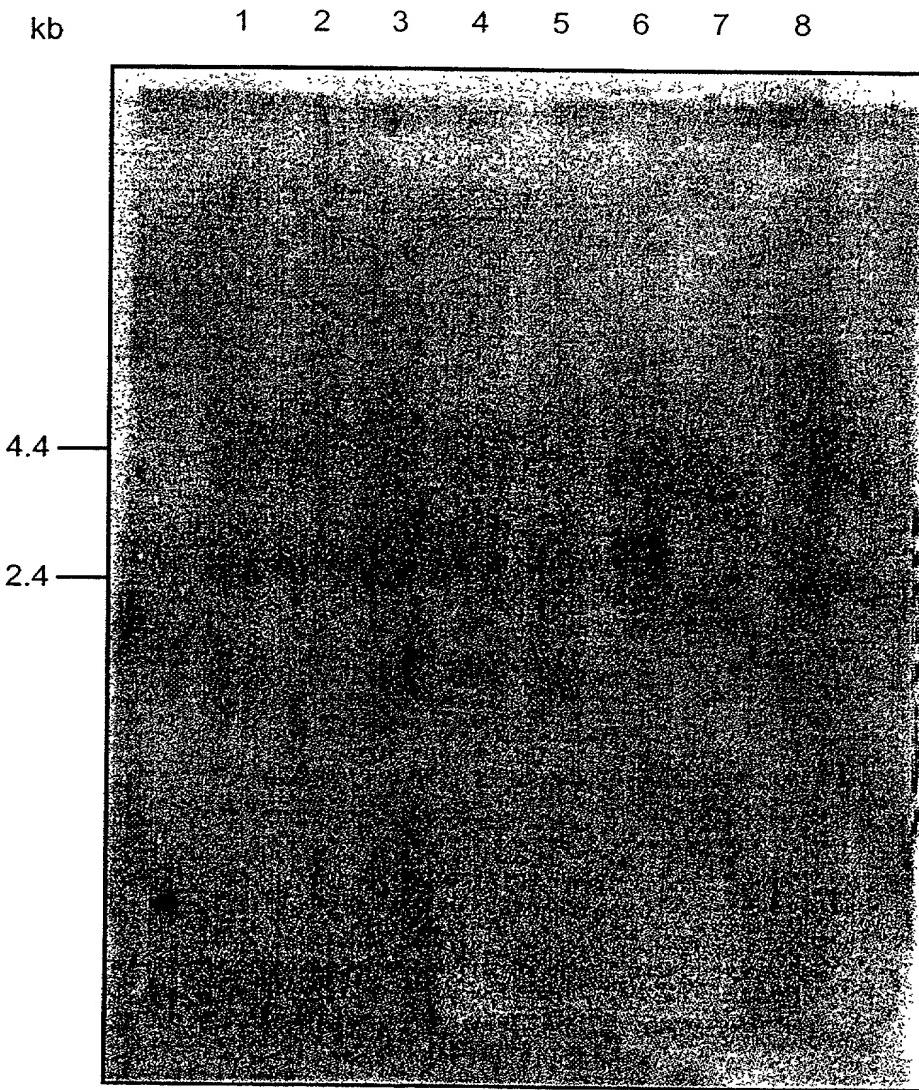


LANE 1: BONE MARROW	LANE 5: SPINAL CORD
LANE 2: ADRENAL GLAND	LANE 6: THYROID
LANE 3: TRACHEA	LANE 7: STOMACH
LANE 4: LYMPH NODE	

40 / 41

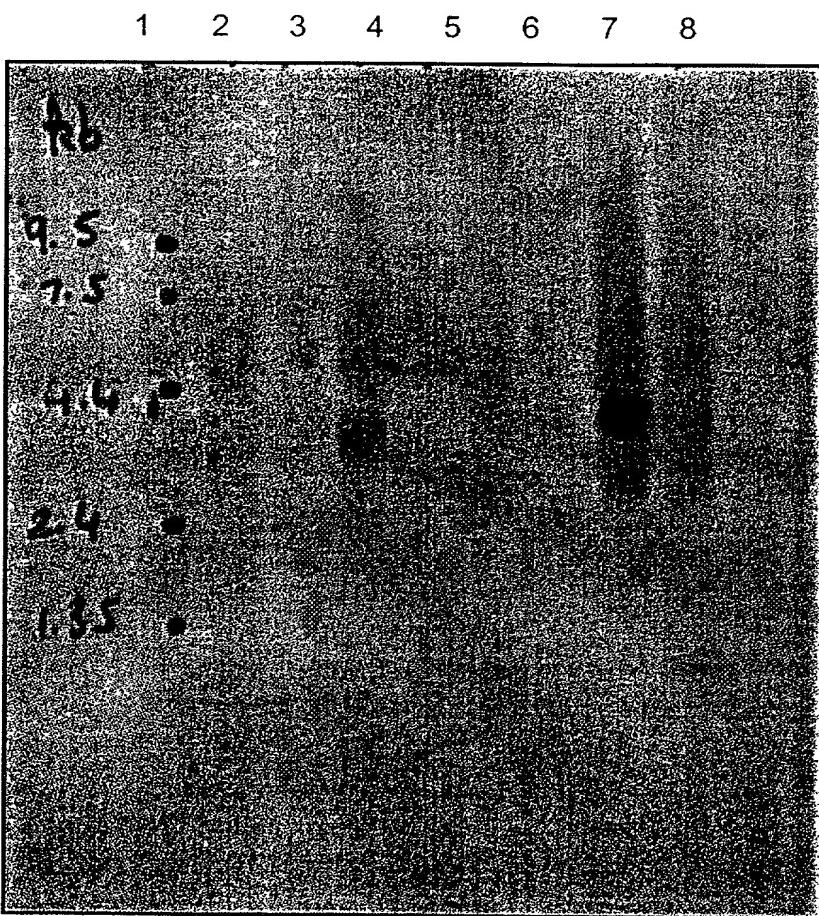
FIG. 22B

HYBRIDISATION OF NORTHERN BLOT WITH hVR3 PROBE



- LANE 1: PERIPHERAL BLOOD LEUKOCYTE
- LANE 2: COLON
- LANE 3: SMALL INTESTINE
- LANE 4: UTERUS
- LANE 5: TESTIS
- LANE 6: PROSTATE
- LANE 7: THYROID
- LANE 8: SPLEEN

FIG. 22C
HYBRIDISATION OF A MULTI-TISSUE NORTHERN
BLOT WITH THE hVR3 PROBE



LANE 1: HEART
LANE 2: BRAIN
LANE 3: PLACENTA
LANE 4: LUNG

LANE 5: LIVER
LANE 6: SKELETAL MUSCLE
LANE 7: KIDNEY
LANE 8: PANCREAS

SEQUENCE LISTING

<110> Glaxo Group Ltd
Tate, Simon N
Delany, Natalie S
Sanseau, P

<120> Novel Receptors

<130> PG3606

<140>
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<151> 1998-12-01

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ctgatgtatgt gtggaccgcgt tgcacagcag ggccgcagt gcggtgtggg tgtgggtggg 240

ccagtctctg ccgctcaccc tattccaggg acacagtctg cttggcttctt ctggactgag 300

ccatctcat caccgagatc ctccctgaat tcagcccacg acagccaccc cggccgtttt 360

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gctaggcctg ctcacctctg aggcctctgg ggtgagaggt tcagtcctgg aaacacttca 600

gttcttagggg gctggggca gcagcaagtt ggagtttgg ggtaccctgc ttacacaggc 660

ccttggcaag gagggcaggt ggggtctaag gacaaggcgt cttactttg ggagtcaacc 720

ccggcgttgtt ggctgctgca gttgcacac tggccacag agatccagc aagg atg 777

Met

1

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Lys Lys Trp Ser Ser Thr Asp Leu Gly Ala Ala Ala Asp Pro Leu Gln

5

10

15

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Lys Asp Thr Cys Pro Asp Pro Leu Asp Gly Asp Pro Asn Ser Arg Pro

20

25

30

cct cca gcc aag ccc cag ctc tcc acg gcc aag agc cgc acc cgg ctc 921

Pro Pro Ala Lys Pro Gln Leu Ser Thr Ala Lys Ser Arg Thr Arg Leu

35

40

45

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Phe Gly Lys Gly Asp Ser Glu Glu Ala Phe Pro Val Asp Cys Pro His

50

55

60

65

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Glu Glu Gly Glu Leu Asp Ser Cys Pro Thr Ile Thr Val Ser Pro Val

70

75

80

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Ile Thr Ile Gln Arg Pro Gly Asp Gly Pro Thr Gly Ala Arg Leu Leu
85 90 95

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Asp Arg Arg Ser Ile Phe Glu Ala Val Ala Gln Asn Asn Cys Gln Asp
115 120 125

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Leu Glu Ser Leu Leu Phe Leu Gln Lys Ser Lys Lys His Leu Thr
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Ala Met Leu Asn Leu His Asp Gly Gln Asn Thr Thr Ile Pro Leu Leu
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Leu Glu Ile Ala Arg Gln Thr Asp Ser Leu Lys Glu Leu Val Asn Ala
180 185 190

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Ser Tyr Thr Asp Ser Tyr Tyr Lys Gly Gln Thr Ala Leu His Ile Ala
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Cys Thr Asn Gln Leu Gly Ile Val Lys Phe Leu Leu Gln Asn Ser Trp
260 265 270

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Gln Thr Ala Asp Ile Ser Ala Arg Asp Ser Val Gly Asn Thr Val Leu
275 280 285

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His Ala Leu Val Glu Val Ala Asp Asn Thr Ala Asp Asn Thr Lys Phe
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Val Thr Ser Met Tyr Asn Glu Ile Leu Ile Leu Gly Ala Lys Leu His
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Pro Thr Leu Lys Leu Glu Glu Leu Thr Asn Lys Lys Gly Met Thr Pro
325 330 335

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340 345 350

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Ser Cys Ile Asp Thr Cys Glu Lys Asn Ser Val Leu Glu Val Ile Ala
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Ile Phe Tyr Phe Asn Phe Leu Val Tyr Cys Leu Tyr Met Ile Ile Phe
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Thr Met Ala Ala Tyr Tyr Arg Pro Val Asp Gly Leu Pro Pro Phe Lys
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Met Glu Lys Ile Gly Asp Tyr Phe Arg Val Thr Gly Glu Ile Leu Ser
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Val Leu Gly Gly Val Tyr Phe Phe Arg Gly Ile Gln Tyr Phe Leu
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Gln Arg Arg Pro Ser Met Lys Thr Leu Phe Val Asp Ser Tyr Ser Glu
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Met Leu Phe Phe Leu Gln Ser Leu Phe Met Leu Ala Thr Val Val Leu
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Tyr Phe Ser His Leu Lys Glu Tyr Val Ala Ser Met Val Phe Ser Leu
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Cys Arg Phe Met Phe Val Tyr Ile Val Phe Leu Phe Gly Phe Ser Thr
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gag tcc acg tcg cac agg tgg cgg ggg cct gcc tgc agg ccc ccc gat 2649
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Ser Ser Tyr Asn Ser Leu Tyr Ser Thr Cys Leu Glu Leu Phe Lys Phe
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Thr Ile Gly Met Gly Asp Leu Glu Phe Thr Glu Asn Tyr Asp Phe Lys
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675 680 685

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725 730 735

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740 745 750

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Val Lys Arg Thr Leu Ser Phe Ser Leu Arg Ser Ser Arg Val Ser Gly
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Arg His Trp Lys Asn Phe Ala Leu Val Pro Leu Leu Arg Glu Ala Ser
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Leu Phe Gly Lys Gly Asp Ser Glu Glu Ala Phe Pro Val Asp Cys Pro
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His Glu Glu Gly Glu Leu Asp Ser Cys Pro Thr Ile Thr Val Ser Pro
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Val Ile Thr Ile Gln Arg Pro Gly Asp Gly Pro Thr Gly Ala Arg Leu
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Leu Ser Gln Asp Ser Val Ala Ala Ser Thr Glu Lys Thr Leu Arg Leu
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Asp Leu Glu Ser Leu Leu Leu Phe Leu Gln Lys Ser Lys Lys His Leu
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Thr Asp Asn Glu Phe Lys Asp Pro Glu Thr Gly Lys Thr Cys Leu Leu
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Lys Ala Met Leu Asn Leu His Asp Gly Gln Asn Thr Thr Ile Pro Leu
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Ala Ser Tyr Thr Asp Ser Tyr Tyr Lys Gly Gln Thr Ala Leu His Ile
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Ala Ile Glu Arg Arg Asn Met Ala Leu Val Thr Leu Leu Val Glu Asn
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Gly Ala Asp Val Gln Ala Ala Ala His Gly Asp Phe Phe Lys Lys Thr
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Ala Cys Thr Asn Gln Leu Gly Ile Val Lys Phe Leu Leu Gln Asn Ser
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Trp Gln Thr Ala Asp Ile Ser Ala Arg Asp Ser Val Gly Asn Thr Val
275 280 285

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Phe Val Thr Ser Met Tyr Asn Glu Ile Leu Ile Leu Gly Ala Lys Leu
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His Pro Thr Leu Lys Leu Glu Glu Leu Thr Asn Lys Lys Gly Met Thr
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Pro Leu Ala Leu Ala Ala Gly Thr Gly Lys Ile Gly Val Leu Ala Tyr
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Lys Phe Thr Glu Trp Ala Tyr Gly Pro Val His Ser Ser Leu Tyr Asp
370 375 380

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Ala Tyr Ser Ser Ser Glu Thr Pro Asn Arg His Asp Met Leu Leu Val
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Glu Pro Leu Asn Arg Leu Leu Gln Asp Lys Trp Asp Arg Phe Val Lys
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Gln Met Gly Ile Tyr Ala Val Met Ile Glu Lys Met Ile Leu Arg Asp
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725 730 735

Asp Asp Tyr Arg Trp Cys Phe Arg Val Asp Glu Val Asn Trp Thr Thr
740 745 750

Trp Asn Thr Asn Val Gly Ile Ile Asn Glu Asp Pro Gly Asn Cys Glu
755 760 765

Gly Val Lys Arg Thr Leu Ser Phe Ser Leu Arg Ser Ser Arg Val Ser
770 775 780

Gly Arg His Trp Lys Asn Phe Ala Leu Val Pro Leu Leu Arg Glu Ala
785 790 795 800

Ser Ala Arg Asp Arg Gln Ser Ala Gln Pro Glu Glu Val Tyr Leu Arg
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Leu Phe Gly Lys Gly Asp Ser Glu Glu Ala Ser Pro Leu Asp Cys Pro

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Tyr Glu Glu Gly Gly Leu Ala Ser Cys Pro Ile Ile Thr Val Ser Ser

65

70

75

80

Val Leu Thr Ile Gln Arg Pro Gly Asp Gly Pro Ala Ser Val Arg Pro

85

90

95

Ser Ser Gln Asp Ser Val Ser Ala Gly Glu Lys Pro Pro Arg Leu Tyr

100

105

110

Asp Arg Arg Ser Ile Phe Asp Ala Val Ala Gln Ser Asn Cys Gln Glu

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Asp Ser Glu Phe Lys Asp Pro Glu Thr Gly Lys Thr Cys Leu Leu Lys

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Ala Met Leu Asn Leu His Asn Gly Gln Asn Asp Thr Ile Ala Leu Leu

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175

Leu Asp Val Ala Arg Lys Thr Asp Ser Leu Lys Gln Phe Val Asn Ala

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185

190

Ser Tyr Thr Asp Ser Tyr Tyr Lys Gly Gln Thr Ala Leu His Ile Ala

195

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Ile Glu Arg Arg Asn Met Thr Leu Val Thr Leu Leu Val Glu Asn Gly

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Ala Asp Val Gln Ala Ala Ala Asn Gly Asp Phe Phe Lys Lys Thr Lys

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Gly Arg Pro Gly Phe Tyr Phe Gly Glu Leu Pro Leu Ser Leu Ala Ala
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Cys Thr Asn Gln Leu Ala Ile Val Lys Phe Leu Leu Gln Asn Ser Trp
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Gln Pro Ala Asp Ile Ser Ala Arg Asp Ser Val Gly Asn Thr Val Leu
275 280 285

His Ala Leu Val Glu Val Ala Asp Asn Thr Val Asp Asn Thr Lys Phe
290 295 300

Val Thr Ser Met Tyr Asn Glu Ile Leu Ile Leu Gly Ala Lys Leu His
305 310 315 320

Pro Thr Leu Lys Leu Glu Glu Ile Thr Asn Arg Lys Gly Leu Thr Pro
325 330 335

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Leu Gln Arg Glu Ile His Glu Pro Glu Cys Arg His Leu Ser Arg Lys
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370 375 380

Ser Cys Ile Asp Thr Cys Glu Lys Asn Ser Val Leu Glu Val Ile Ala
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Tyr Ser Ser Ser Glu Thr Pro Asn Arg His Asp Met Leu Leu Val Glu
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Pro Leu Asn Arg Leu Leu Gln Asp Lys Trp Asp Arg Phe Val Lys Arg
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675 680 685

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725 730 735

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755 760 765

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785 790 795 800

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110 115 120

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140 145 150

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285 290 295

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Arg Asn Asp Thr Ile Pro Val Leu Leu Asp Ile Ala Glu Arg Thr Gly
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Asn Met Arg Glu Phe Ile Asn Ser Pro Phe Arg Asp Ile Tyr Tyr Arg
315 320 325

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365 370 375

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Glu Leu Pro Leu Ser Leu Ala Ala Cys Thr Asn Gln Pro His Ile Val
380 385 390

aac tac ctg acg gag aac ccc cac aag aag gcg gac atg cg_g cg_c cag 1912
Asn Tyr Leu Thr Glu Asn Pro His Lys Lys Ala Asp Met Arg Arg Gln
395 400 405

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410           415           420           425

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Leu Asn Asn Asp Gly Leu Ser Pro Leu Met Met Ala Ala Lys Thr Gly
460 465 470

aag att ggg atc ttt cag cac atc atc cgg cgg gag gtg acg gat gag 2152
Lys Ile Gly Ile Phe Gln His Ile Ile Arg Arg Glu Val Thr Asp Glu
475 480 485

gac aca cgg cac ctg tcc cgc aag tcc aag gac tgg gcc tat ggg cca 2200
Asp Thr Arg His Leu Ser Arg Lys Ser Lys Asp Trp Ala Tyr Gly Pro
490 495 500 505

gtg tat tcc tcg ctt tat gac ctc tcc ctg gac acg tgt ggg gaa 2248
Val Tyr Ser Ser Leu Tyr Asp Leu Ser Ser Leu Asp Thr Cys Gly Glu
510 515 520

gag gcc tcc gtg ctg gag atc ctg gtg tac aac agc aag att gag aac 2296
Glu Ala Ser Val Leu Glu Ile Leu Val Tyr Asn Ser Lys Ile Glu Asn
525 530 535

cgc cac gag atg ctg gct gtg gag ccc atc aat gaa ctg ctg cgg gac 2344
Arg His Glu Met Leu Ala Val Glu Pro Ile Asn Glu Leu Leu Arg Asp
540 545 550

aag tgg cgg aag ttc ggg gcc gtc tcc ttc tac atc aac gtg gtc tcc 2392
Lys Trp Arg Lys Phe Gly Ala Val Ser Phe Tyr Ile Asn Val Val Ser
555 560 565

tac ctg tgt gcc atg gtt atc ttc act ctc acc gcc tac tac cag ccg 2440
Tyr Leu Cys Ala Met Val Ile Phe Thr Leu Thr Ala Tyr Tyr Gln Pro
570 575 580 585

ctg gag ggc aca ccg ccg tac cct tac cgc acc acg gtg gac tac ctg 2488
Leu Glu Gly Thr Pro Pro Tyr Pro Tyr Arg Thr Thr Val Asp Tyr Leu
590 595 600

cgg ctg gct ggc gag gtc att acg ctc ttc act ggg gtc ctg ttc ttc 2536
Arg Leu Ala Gly Glu Val Ile Thr Leu Phe Thr Gly Val Leu Phe Phe
605 610 615

ttc acc aac atc aaa gac ttg ttc atg aag aaa tgc cct gga gtg aat 2584
Phe Thr Asn Ile Lys Asp Leu Phe Met Lys Lys Cys Pro Gly Val Asn
620 625 630

tct ctc ttc att gat ggc tcc ttc cag ctg ctc tac ttc atc tac tct 2632
Ser Leu Phe Ile Asp Gly Ser Phe Gln Leu Leu Tyr Phe Ile Tyr Ser
635 640 645

gtc ctg gtg atc gtc tca gca gcc ctc tac ctg gca ggg atc gag gcc 2680
Val Leu Val Ile Val Ser Ala Ala Leu Tyr Leu Ala Gly Ile Glu Ala
650 655 660 665

tac ctg gcc atg atg gtc ttt gcc ctg gtc ctg ggc tgg atg aat gcc 2728
Tyr Leu Ala Met Met Val Phe Ala Leu Val Leu Gly Trp Met Asn Ala
670 675 680

ctt tac ttc acc cgt ggg ctg aag ctg acg ggg acc tat agc atc atg 2776
Leu Tyr Phe Thr Arg Gly Leu Lys Leu Thr Gly Thr Tyr Ser Ile Met
685 690 695

atc cag aag att ctc ttc aag gac ctt ttc cga ttc ctg ctc gtc tac 2824
Ile Gln Lys Ile Leu Phe Lys Asp Leu Phe Arg Phe Leu Leu Val Tyr
700 705 710

ttg ctc ttc atg atc ggc tac gct tca gcc ctg gtc tcc ctc ctg aac 2872
Leu Leu Phe Met Ile Gly Tyr Ala Ser Ala Leu Val Ser Leu Leu Asn
715 720 725

ccg tgt gcc aac atg aag gtg tgc aat gag gac cag acc aac tgc aca 2920
Pro Cys Ala Asn Met Lys Val Cys Asn Glu Asp Gln Thr Asn Cys Thr
730 735 740 745

gtg ccc act tac ccc tcg tgc cgt gac agc gag acc ttc agc acc ttc 2968
Val Pro Thr Tyr Pro Ser Cys Arg Asp Ser Glu Thr Phe Ser Thr Phe
750 755 760

ctc ctg gac ctg ttt aag ctg acc atc ggc atg ggc gac ctg gag atg 3016
Leu Leu Asp Leu Phe Lys Leu Thr Ile Gly Met Gly Asp Leu Glu Met
765 770 775

ctg agc agc acc aag tac ccc gtg gtc ttc atc atc ctg ctg gtg acc 3064
Leu Ser Ser Thr Lys Tyr Pro Val Val Phe Ile Ile Leu Leu Val Thr
780 785 790

tac atc atc ctc acc tct gtg ctg ctc aac atg ctc att gcc ctc 3112
Tyr Ile Ile Leu Thr Ser Val Leu Leu Leu Asn Met Leu Ile Ala Leu
795 800 805

atg ggc gag aca gtg ggc cag gtc tcc aag gag agc aag cac atc tgg 3160
Met Gly Glu Thr Val Gly Gln Val Ser Lys Glu Ser Lys His Ile Trp
810 815 820 825

aag ctg cag tgg gcc acc acc atc ctg gac att gag cgc tcc ttc ccc 3208
Lys Leu Gln Trp Ala Thr Thr Ile Leu Asp Ile Glu Arg Ser Phe Pro
830 835 840

gta ttc ctg agg aag gcc ttc cgc tct ggg gag atg gtc acc gtg ggc 3256
Val Phe Leu Arg Lys Ala Phe Arg Ser Gly Glu Met Val Thr Val Gly
845 850 855

aag agc tcg gac ggc act cct gac cgc agg tgg tgc ttc agg gtg gat 3304
Lys Ser Ser Asp Gly Thr Pro Asp Arg Arg Trp Cys Phe Arg Val Asp
860 865 870

gag gtg aac tgg tct cac tgg aac cag aac ttg ggc atc atc aac gag 3352
Glu Val Asn Trp Ser His Trp Asn Gln Asn Leu Gly Ile Ile Asn Glu
875 880 885

gac ccg ggc aag aat gag acc tac cag tat tat ggc ttc tcg cat acc 3400
Asp Pro Gly Lys Asn Glu Thr Tyr Gln Tyr Gly Phe Ser His Thr
890 895 900 905

gtg ggc cgc ctc cgc agg gat cgc tgg tcc tcg gtg gta ccc cgc gtg 3448
Val Gly Arg Leu Arg Arg Asp Arg Trp Ser Ser Val Val Pro Arg Val
910 915 920

gtg gaa ctg aac aag aac tcg aac ccg gac gag gtg gtg gtg cct ctg 3496
Val Glu Leu Asn Lys Asn Ser Asn Pro Asp Glu Val Val Val Pro Leu
925 930 935

gac agc atg ggg aac ccc cgc tgc gat ggc cac cag cag ggt tac ccc 3544
Asp Ser Met Gly Asn Pro Arg Cys Asp Gly His Gln Gln Gly Tyr Pro
940 945 950

cgc aag tgg agg act gat gac gcc ccg ctc tag ggactgcagc ccagccccag 3597
Arg Lys Trp Arg Thr Asp Asp Ala Pro Leu
955 960

cttctctgcc cactcatttc tagtccagcc gcatttcagc agtgccttct ggggtgtccc 3657

cccacaccct gctttggccc cagaggcgag ggaccagtgg aggtgccagg gaggccccag 3717

gaccctgtgg tccccctggct ctgcctcccc accctgggtt gggggctccc gcccacctgt 3777

cttgctccta tggagtcaca taagccaacg ccagagcccc tccacctcag gccccagccc 3837

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ggaacctggc agaggcctta ggaccccggtt ccaagtgcac tgcccgccca agccccagcc 3957

tcagcctgcg cctgagctgc atgcgccacc attttggca gcgtggcagc tttgcaaggg 4017

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ggtgctcaat aaatgtttat tcattgaaaa aaaaaaaaaa a 4118

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<211> 963

<212> PRT

<213> Homo sapiens

<400> 5

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20 25 30

Ile Trp Glu Trp Pro Pro Cys Ala Pro Val Ile Thr Thr Val Ala Leu

35 40 45

Lys Gln Leu Ala Ala Leu Leu Leu Val His Val Gly Gly Gly Phe Leu

50 55 60

Glu Pro Pro Pro Leu Ala Gly Phe Cys Leu Thr Pro Leu Ser Phe Pro

65 70 75 80

Cys Arg Leu Ser Ser Ala Asp Gly Pro Gly Ala Gly Met Ala Asp Ser

85 90 95

Ser Glu Gly Pro Arg Ala Gly Pro Gly Glu Val Ala Glu Leu Pro Gly

100 105 110

Asp Glu Ser Gly Thr Pro Gly Gly Glu Ala Phe Pro Leu Ser Ser Leu

115 120 125

Ala Asn Leu Phe Glu Gly Glu Asp Gly Ser Leu Ser Pro Ser Pro Ala

130 135 140

Asp Ala Ser Arg Pro Ala Gly Pro Gly Asp Gly Arg Pro Asn Leu Arg
145 150 155 160

Met Lys Phe Gln Gly Ala Phe Arg Lys Gly Val Pro Asn Pro Ile Asp
165 170 175

Leu Leu Glu Ser Thr Leu Tyr Glu Ser Ser Val Val Pro Gly Pro Lys
180 185 190

Lys Ala Pro Met Asp Ser Leu Phe Asp Tyr Gly Thr Tyr Arg His His
195 200 205

Ser Ser Asp Asn Lys Arg Trp Arg Lys Lys Ile Ile Glu Lys Gln Pro
210 215 220

Gln Ser Pro Lys Ala Pro Ala Pro Gln Pro Pro Pro Ile Leu Lys Val
225 230 235 240

Phe Asn Arg Pro Ile Leu Phe Asp Ile Val Ser Arg Gly Ser Thr Ala
245 250 255

Asp Leu Asp Gly Leu Leu Pro Phe Leu Leu Thr His Lys Lys Arg Leu
260 265 270

Thr Asp Glu Glu Phe Arg Glu Pro Ser Thr Gly Lys Thr Cys Leu Pro
275 280 285

Lys Ala Leu Leu Asn Leu Ser Asn Gly Arg Asn Asp Thr Ile Pro Val
290 295 300

Leu Leu Asp Ile Ala Glu Arg Thr Gly Asn Met Arg Glu Phe Ile Asn
305 310 315 320

Ser Pro Phe Arg Asp Ile Tyr Tyr Arg Gly Gln Thr Ala Leu His Ile
325 330 335

Ala Ile Glu Arg Arg Cys Lys His Tyr Val Glu Leu Leu Val Ala Gln
340 345 350

Gly Ala Asp Val His Ala Gln Ala Arg Gly Arg Phe Phe Gln Pro Lys
355 360 365

Asp Glu Gly Gly Tyr Phe Tyr Phe Gly Glu Leu Pro Leu Ser Leu Ala
370 375 380

Ala Cys Thr Asn Gln Pro His Ile Val Asn Tyr Leu Thr Glu Asn Pro
385 390 395 400

His Lys Lys Ala Asp Met Arg Arg Gln Asp Ser Arg Gly Asn Thr Val
405 410 415

Leu His Ala Leu Val Ala Ile Ala Asp Asn Thr Arg Glu Asn Thr Lys
420 425 430

Phe Val Thr Lys Met Tyr Asp Leu Leu Leu Lys Cys Ala Arg Leu
435 440 445

Phe Pro Asp Ser Asn Leu Glu Ala Val Leu Asn Asn Asp Gly Leu Ser
450 455 460

Pro Leu Met Met Ala Ala Lys Thr Gly Lys Ile Gly Ile Phe Gln His
465 470 475 480

Ile Ile Arg Arg Glu Val Thr Asp Glu Asp Thr Arg His Leu Ser Arg
485 490 495

Lys Ser Lys Asp Trp Ala Tyr Gly Pro Val Tyr Ser Ser Leu Tyr Asp
500 505 510

Leu Ser Ser Leu Asp Thr Cys Gly Glu Glu Ala Ser Val Leu Glu Ile
515 520 525

Leu Val Tyr Asn Ser Lys Ile Glu Asn Arg His Glu Met Leu Ala Val
530 535 540

Glu Pro Ile Asn Glu Leu Leu Arg Asp Lys Trp Arg Lys Phe Gly Ala
545 550 555 560

Val Ser Phe Tyr Ile Asn Val Val Ser Tyr Leu Cys Ala Met Val Ile
565 570 575

Phe Thr Leu Thr Ala Tyr Tyr Gln Pro Leu Glu Gly Thr Pro Pro Tyr
580 585 590

Pro Tyr Arg Thr Thr Val Asp Tyr Leu Arg Leu Ala Gly Glu Val Ile
595 600 605

Thr Leu Phe Thr Gly Val Leu Phe Phe Thr Asn Ile Lys Asp Leu
610 615 620

Phe Met Lys Lys Cys Pro Gly Val Asn Ser Leu Phe Ile Asp Gly Ser
625 630 635 640

Phe Gln Leu Leu Tyr Phe Ile Tyr Ser Val Leu Val Ile Val Ser Ala
645 650 655

Ala Leu Tyr Leu Ala Gly Ile Glu Ala Tyr Leu Ala Met Met Val Phe
660 665 670

Ala Leu Val Leu Gly Trp Met Asn Ala Leu Tyr Phe Thr Arg Gly Leu
675 680 685

Lys Leu Thr Gly Thr Tyr Ser Ile Met Ile Gln Lys Ile Leu Phe Lys
690 695 700

Asp Leu Phe Arg Phe Leu Leu Val Tyr Leu Leu Phe Met Ile Gly Tyr
705 710 715 720

Ala Ser Ala Leu Val Ser Leu Leu Asn Pro Cys Ala Asn Met Lys Val
725 730 735

Cys Asn Glu Asp Gln Thr Asn Cys Thr Val Pro Thr Tyr Pro Ser Cys
740 745 750

Arg Asp Ser Glu Thr Phe Ser Thr Phe Leu Leu Asp Leu Phe Lys Leu
755 760 765

Thr Ile Gly Met Gly Asp Leu Glu Met Leu Ser Ser Thr Lys Tyr Pro
770 775 780

Val Val Phe Ile Ile Leu Leu Val Thr Tyr Ile Ile Leu Thr Ser Val
785 790 795 800

Leu Leu Leu Asn Met Leu Ile Ala Leu Met Gly Glu Thr Val Gly Gln
805 810 815

Val Ser Lys Glu Ser Lys His Ile Trp Lys Leu Gln Trp Ala Thr Thr
820 825 830

Ile Leu Asp Ile Glu Arg Ser Phe Pro Val Phe Leu Arg Lys Ala Phe
835 840 845

Arg Ser Gly Glu Met Val Thr Val Gly Lys Ser Ser Asp Gly Thr Pro
850 855 860

Asp Arg Arg Trp Cys Phe Arg Val Asp Glu Val Asn Trp Ser His Trp
865 870 875 880

Asn Gln Asn Leu Gly Ile Ile Asn Glu Asp Pro Gly Lys Asn Glu Thr
885 890 895

Tyr Gln Tyr Tyr Gly Phe Ser His Thr Val Gly Arg Leu Arg Arg Asp
900 905 910

Arg Trp Ser Ser Val Val Pro Arg Val Val Glu Leu Asn Lys Asn Ser
915 920 925

Asn Pro Asp Glu Val Val Pro Leu Asp Ser Met Gly Asn Pro Arg
930 935 940

Cys Asp Gly His Gln Gln Gly Tyr Pro Arg Lys Trp Arg Thr Asp Asp
945 950 955 960

Ala Pro Leu

<210> 6

<211> 764

<212> PRT

<213> Homo sapiens

<400> 6

Met Thr Ser Pro Ser Ser Pro Val Phe Arg Leu Glu Thr Leu Asp
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Gly Gly Gln Glu Asp Gly Ser Glu Ala Asp Arg Gly Lys Leu Asp Phe
20 25 30

Gly Ser Gly Leu Pro Pro Met Glu Ser Gln Phe Gln Gly Glu Asp Arg
35 40 45

Lys Phe Ala Pro Gln Ile Arg Val Asn Leu Asn Tyr Arg Lys Gly Thr
50 55 60

Gly Ala Ser Gln Pro Asp Pro Asn Arg Phe Asp Arg Asp Arg Leu Phe
65 70 75 80

Asn Ala Val Ser Arg Gly Val Pro Glu Asp Leu Ala Gly Leu Pro Glu
85 90 95

Tyr Leu Ser Lys Thr Ser Lys Tyr Leu Thr Asp Ser Glu Tyr Thr Glu
100 105 110

Gly Ser Thr Gly Lys Thr Cys Leu Met Lys Ala Val Leu Asn Leu Lys
115 120 125

Asp Gly Val Asn Ala Cys Ile Leu Pro Leu Leu Gln Ile Asp Arg Asp
130 135 140

Ser Gly Asn Pro Gln Pro Leu Val Asn Ala Gln Cys Thr Asp Asp Tyr
145 150 155 160

Tyr Arg Gly His Ser Ala Leu His Ile Ala Ile Glu Lys Arg Ser Leu
165 170 175

Gln Cys Val Lys Leu Leu Val Glu Asn Gly Ala Asn Val His Ala Arg
180 185 190

Ala Cys Gly Arg Phe Phe Gln Lys Gly Gln Gly Thr Cys Phe Tyr Phe
195 200 205

Gly Glu Leu Pro Leu Ser Leu Ala Ala Cys Thr Lys Gln Trp Asp Val
210 215 220

Val Ser Tyr Leu Leu Glu Asn Pro His Gln Pro Ala Ser Leu Gln Ala
225 230 235 240

Thr Asp Ser Gln Gly Asn Thr Val Leu His Ala Leu Val Met Ile Ser
245 250 255

Asp Asn Ser Ala Glu Asn Ile Ala Leu Val Thr Ser Met Tyr Asp Gly
260 265 270

Leu Leu Gln Ala Gly Ala Arg Leu Cys Pro Thr Val Gln Leu Glu Asp
275 280 285

Ile Arg Asn Leu Gln Asp Leu Thr Pro Leu Lys Leu Ala Ala Lys Glu
290 295 300

Gly Lys Ile Glu Ile Phe Arg His Ile Leu Gln Arg Glu Phe Ser Gly
305 310 315 320

Leu Ser His Leu Ser Arg Lys Phe Thr Glu Trp Cys Tyr Gly Pro Val
325 330 335

Arg Val Ser Leu Tyr Asp Leu Ala Ser Val Asp Ser Cys Glu Glu Asn
340 345 350

Ser Val Leu Glu Ile Ile Ala Phe His Cys Lys Ser Pro His Arg His
355 360 365

Arg Met Val Val Leu Glu Pro Leu Asn Lys Leu Leu Gln Ala Lys Trp
370 375 380

Asp Leu Leu Ile Pro Lys Phe Phe Leu Asn Phe Leu Cys Asn Leu Ile
385 390 395 400

Tyr Met Phe Ile Phe Thr Ala Val Ala Tyr His Gln Pro Thr Leu Lys
405 410 415

Lys Gln Ala Ala Pro His Leu Lys Ala Glu Val Gly Asn Ser Met Leu
420 425 430

Leu Thr Gly His Ile Leu Ile Leu Leu Gly Gly Ile Tyr Leu Leu Val
435 440 445

Gly Gln Leu Trp Tyr Phe Trp Arg Arg His Val Phe Ile Trp Ile Ser
450 455 460

Phe Ile Asp Ser Tyr Phe Glu Ile Leu Phe Leu Phe Gln Ala Leu Leu
465 470 475 480

Thr Val Val Ser Gln Val Leu Cys Phe Leu Ala Ile Glu Trp Tyr Leu
485 490 495

Pro Leu Leu Val Ser Ala Leu Val Leu Gly Trp Leu Asn Leu Leu Tyr
500 505 510

Tyr Thr Arg Gly Phe Gln His Thr Gly Ile Tyr Ser Val Met Ile Gln
515 520 525

Lys Val Ile Leu Arg Asp Leu Leu Arg Phe Leu Leu Ile Tyr Leu Val
530 535 540

Phe Leu Phe Gly Phe Ala Val Ala Leu Val Ser Leu Ser Gln Glu Ala
545 550 555 560

Trp Arg Pro Glu Ala Pro Thr Gly Pro Asn Ala Thr Glu Ser Val Gln
565 570 575

Pro Met Glu Gly Gln Glu Asp Glu Gly Asn Gly Ala Gln Tyr Arg Gly
580 585 590

Ile Leu Glu Ala Ser Leu Glu Leu Phe Lys Phe Thr Ile Gly Met Gly

595 600 605

Glu Leu Ala Phe Gln Gln Glu Leu His Phe Arg Gly Met Val Leu Leu

610 615 620

Leu Leu Leu Ala Tyr Val Leu Leu Thr Tyr Ile Leu Leu Leu Asn Met

625 630 635 640

Leu Ile Ala Leu Met Ser Glu Thr Val Asn Ser Val Ala Thr Asp Ser

645 650 655

Trp Ser Ile Trp Lys Leu Gln Lys Ala Ile Ser Val Leu Glu Met Glu

660 665 670

Asn Gly Tyr Trp Trp Cys Arg Lys Lys Gln Arg Ala Gly Val Met Leu

675 680 685

Thr Val Gly Thr Lys Pro Asp Gly Ser Pro Asp Glu Arg Trp Cys Phe

690 695 700

Arg Val Glu Glu Val Asn Trp Ala Ser Trp Glu Gln Thr Leu Pro Thr

705 710 715 720

Leu Cys Glu Asp Pro Ser Gly Ala Gly Val Pro Arg Thr Leu Glu Asn

725 730 735

Pro Val Leu Ala Ser Pro Pro Lys Glu Asp Glu Asp Gly Ala Ser Glu

740 745 750

Glu Asn Tyr Val Pro Val Gln Leu Leu Gln Ser Asn

755 760

<210> 7
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<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 7
atttaggta cactata 18

<210> 8
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 8
taatacgact cactata 20

<210> 9
<211> 19
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 9
ggaaacagct atgaccat 19

<210> 10
<211> 17
<212> DNA
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<220>
<223> Description of Artificial Sequence: Primer

<400> 10
gtaaaaacgac ggccagt 17

<210> 11
<211> 20
<212> DNA
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<220>
<223> Description of Artificial Sequence: Primer

<400> 11
aatttaaccct cactaaaggg 20

<210> 12
<211> 20
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<220>
<223> Description of Artificial Sequence: Primer

<400> 12
tctacttcgg tgaactgcc 20

<210> 13
<211> 20
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<220>
<223> Description of Artificial Sequence: Primer

<400> 13
acggcaggga gtcattcttc

20

<210> 14
<211> 19
<212> DNA
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<220>
<223> Description of Artificial Sequence: Primer

<400> 14
ctgcagaact cctggcaga

19

<210> 15
<211> 20
<212> DNA
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<220>
<223> Description of Artificial Sequence: Primer

<400> 15
gtcaccacccg ctgtggaaaa

20

<210> 16
<211> 21
<212> DNA
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<220>
<223> Description of Artificial Sequence: Primer

<400> 16
tcctctggct tccaaaccgt t

21

<210> 17
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<212> DNA
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<220>
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<400> 17
gaactggcca gaaagtgcct

20

<210> 18
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 18
ctggagttag ggtctccatc c

21

<210> 19
<211> 43
<212> DNA
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<220>
<223> Description of Artificial Sequence: Primer

<400> 19
gtcatagcgccgcgaccatgaagaatggagcagcac

43

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<211> 20
<212> DNA
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<220>
<223> Description of Artificial Sequence: Primer

<400> 20
aggcccactc ggtgaacttc

20

<210> 21
<211> 20
<212> DNA
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<220>
<223> Description of Artificial Sequence: Primer

<400> 21
gacgagcatgtacaatgaga

20

<210> 22
<211> 20
<212> DNA
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<220>
<223> Description of Artificial Sequence: Primer

<400> 22
gtcaccacccg ctgtggaaaaa 20

<210> 23
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 23
tgtggacagc tacagtgaga 20

<210> 24
<211> 32
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 24
tgcaactgaat tcgagcactg gtgttccctc ag 32

<210> 25
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 25
tgtggacagc tacagtgaga

20

<210> 26
<211> 19
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 26
gtggaaaacc cgaacaaga

19

<210> 27
<211> 23
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<220>
<223> Description of Artificial Sequence: Synthetic sequence

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Asp Ser Glu Glu Ala Ser Cys

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<211> 21

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Synthetic sequence

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Cys Gly Ser Leu Lys Pro Glu Asp Ala Glu Val Phe Lys Asp Ser Met

1

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15

Val Pro Gly Glu Lys

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<210> 29

<211> 20

<212> DNA

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<223> Description of Artificial Sequence: Primer

<400> 29

atggccacca gcagggttac

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<210> 30

<211> 18

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 30

tctgccaggt tccagctg

18

<210> 31

<211> 41

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 31

gtcatagcg cccgcgcgcca ccatgccca ggttagttggaa c

41

<210> 32

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 32

cacctcttgt tgtcactggaa

20

<210> 33

<211> 23

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 33

caaatctgcg catgaagtgc cag

23

<210> 34
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 34
gccacgagaa gttccacgt a gtg

23

<210> 35
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 35
gctgctcccc ttcttgctga

20

<210> 36
<211> 32
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 36
tgcactctcg agaaaatgagt gggcagagaa gc

32

<210> 37
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 37
atggccacca gcagggttac

20

<210> 38
<211> 18
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 38
tctgccaggt tccagctg

18

<210> 39
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Primer

<400> 39
acaagaaggc ggacatgcgg

20

<210> 40

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 40

atctcgtggc ggttctcaat

20

ABSTRACT

HUMAN VANILLOID RECEPTORS AND THEIR USES

The invention provides novel human vanilloid receptor (hVR) proteins, in particular hVR1 and hVR3, nucleotide sequences encoding for the novel hVR proteins, and hVR proteins for use in a method for screening for agents useful in the treatment or prophylaxis of disorders which are responsive to modulation of hVR activity in a human patient. The invention also provides expression vectors comprising said nucleotide sequences, stable cell lines comprising said expression vectors, antibodies specific for the novel hVR proteins, methods for the identification of compounds which exhibit hVR modulating activity, compounds identifiable and identified by such methods, and methods of treatment or prophylaxis of disorders which are responsive to modulation of hVR activity in a human patient.

PCT/US00/12522

DECLARATION FOR "371" APPLICATION

**COMBINED DECLARATION FOR UTILITY OR DESIGN PATENT
APPLICATION WITH POWER OF ATTORNEY**

() Declaration submitted with initial filing or

(X) Declaration submitted after initial filing (surcharge required 37CFR1.16(e))

ATTORNEY'S DOCKET
PG3606USWFirst Names Inventor:
DELANYComplete if known:
App No.:Filing Date
Concurrently herewith
Group Art Unit:

As below named inventor. I hereby declare that:

23347

My residence, post office address and citizenship are as stated below next to my name. U.S. PATENT TRADEMARK OFFICE

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

HUMAN VANILLOID RECEPTORS AND THEIR USES

the specification of which (check only one item below):

[] is attached hereto.

OR

[X] was filed on _____ as United States application Serial No. _____ or PCT International

Application Number EP99/09284 filed 11/30/1999 and was amended on (MM/DD/YYYY) _____ (if applicable)

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment specifically referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR §1.56.

I hereby claim foreign priority benefits under 35, U.S.C. §119 (a)-(d) or §365(b) of any foreign applications(s) for patent or inventor's certificate or 365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate or of any PCT international application having a filing date before that of the application on which priority is claimed:

PRIOR FOREIGN AND ANY PRIORITY CLAIMS UNDER 35 U.S.C. 119:

Prior Foreign Application Number (s)	Country	Foreign Filing Date (MM/DD/YYYY)	PRIORITY CLAIMED
1. GB9826359.3	GB	12/01/1998	x
2.			
3.			
4.			
5.			

I hereby claim the benefit under Title 35, United States Code §119(e) of any United States provisional application(s) listed below:

Application No.	Filing Date (MM/DD/YYYY)

DECLARATION FOR "371" APPLICATION

**COMBINED DECLARATION FOR UTILITY or DESIGN
PATENT APPLICATION WITH POWER OF ATTORNEY** Continued

 ATTORNEY'S DOCKET NUMBER
 PG3606USW

I hereby claim the benefit under 35, U.S.C. §120 of any United States application or §365(c) of any PCT international application designating the United States of America that is listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. §112, I acknowledge the duty to disclose information which is material to patentability as defined in 37 C.F.R. §1.56 which became available between the filing date of the prior application(s) and the national or PCT international filing date of this application:

PRIOR U.S. PARENT APPLICATION or PCT PARENT APPLICATION

		STATUS (Check one)		
U.S. Parent Application or PCT Parent Number	Parent Filing Date (MM/DD/YYYY)	PATENTED	PENDING	ABANDONED

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the U.S. Patent and Trademark Office connected therewith. (List name and registration number)

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under 18 U.S.C. 1001, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

1. 02	FULL NAME OF INVENTOR <u>DELANY</u>	FAMILY NAME <u>DELANY</u>	FIRST GIVEN NAME <u>Natalie</u>	SECOND GIVEN NAME/INITIAL <u>Samantha</u>
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0	INVENTOR'S SIGNATURE <u>J Scangeau</u>			DATE: <u>19/6/01</u>
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0	INVENTOR'S SIGNATURE <u>Simon Tate</u>			DATE: <u>24/6/01</u>
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